

Estimating Latest Cretaceous and Tertiary Atmospheric CO₂ Concentration from Stomatal Indices

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The role of atmospheric CO₂ in determining climate is important for understanding patterns in the geologic record and predicting future climate. Because of the recent anthropogenic rise in temperatures, the role of CO₂ in globally warm periods is particularly important for any predictions of future climate change. Here, I reconstruct the concentration of atmospheric CO₂ using patterns of stomatal index in the plant cuticles of *Ginkgo* and *Metasequoia* for two globally warm intervals, the latest Cretaceous to early Eocene (66-53 Ma) and the middle Miocene (17-15 Ma).

Most vascular C₃ plants show an inverse relationship between the stomatal index in their leaves and the level of atmospheric CO₂. Stomatal index (SI) has been shown experimentally and in the field to be largely independent of every environmental factor (e.g., water stress) except CO₂. There is a potential, then, to use patterns in SI as a proxy for ancient levels of atmospheric CO₂. One drawback to SI-CO₂ relationships is that they are generally species-specific. Modern *Ginkgo* and *Metasequoia* are unusual because both have morphologically identical forms back to the Early and Late Cretaceous, respectively. In addition, at least in the case *Ginkgo*, its sedimentological and floral associations remain stable throughout the Cenozoic. Thus, both *Ginkgo* and *Metasequoia* represent excellent taxa for applying this CO₂ proxy back into the Cretaceous.

Based upon dated herbaria sheets, which document the anthropogenic rise in CO₂, and saplings grown in greenhouses at ambient and elevated CO₂ (350-800 ppmV), the SI-CO₂ relationships for *Ginkgo* and *Metasequoia* were determined. These relationships were then applied to fossil cuticles of *Ginkgo* and *Metasequoia* from 30 sites in order to reconstruct CO₂. The resulting reconstruction indicates near present-day levels of CO₂ (300-450 ppmV) for both the early Paleogene and middle Miocene. This suggests that other radiative forcings were more important during these intervals than they are today, and that these intervals may not serve as good analogs for understanding future climate change.

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from Stomatal Indices**

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As the old cliché goes, I am standing on the shoulders of giants. Estimates of paleo-CO₂ are not meaningful unless they can be placed into a temporal framework. My work therefore relies upon robust pre-existing stratigraphic frameworks. Secondly, and more obviously, stomatal-based estimates of paleo-CO₂ are not possible unless there are fossil cuticles to be measured. My work therefore also relies upon museum collections of cuticles and well-scouted field areas for which I can visit and have reasonable chances of success for extracting fossil cuticle. Both of these requirements were met in spades in the Bighorn Basin. It is not a coincidence that the bulk of my data come from this basin. I am deeply indebted to the countless workers of stratigraphy, biostratigraphy (vertebrate paleontology, paleobotany, and palynology), magnetostratigraphy, and chronostratigraphy who, over the last 100+ years, have mapped out this basin to a degree perhaps unmatched in the world. I also thank the paleobotanists of the Bighorn Basin (in particular, Leo Hickey and Scott Wing) who have brought back truckloads of plant fossils and detailed fieldnotes to return for more. I think that my study serves as an example of what can be achieved in areas where the stratigraphy and “stamp book collecting” (i.e., the hard stuff, in my opinion) have been well done.

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NOTE ON FORMATTING

All of my dissertation chapters have been published elsewhere. Care has been taken to balance any standardization of formatting with preserving the content of the chapters. The published references for the chapters are as follows:

Chapter 1

Royer, D.L., 2001. Stomatal density and stomatal index as indicators of paleoatmospheric CO₂ concentration. *Review of Palaeobotany and Palynology*, 114: 1-28.

Chapter 2

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Chapter 3

Royer, D.L., Wing, S.L., Beerling, D.J., Jolley, D.W., Koch, P.L., Hickey, L.J., Berner, R.A., 2001. Paleobotanical evidence for near present-day levels of atmospheric CO₂ during part of the Tertiary. *Science*, 292: 2310-2313.

Chapter 4

Royer, D.L., 2002. Estimating latest Cretaceous and Tertiary PCO₂ from stomatal indices. *Geological Society of America Special Paper* (in review).

INTRODUCTION

It is the purpose of this study to reconstruct the concentration of paleoatmospheric CO₂ from patterns in the stomatal index (SI) in the fossil cuticles of *Ginkgo adiantoides* and *Metasequoia occidentalis*. CO₂ is an important greenhouse gas, and so understanding the role(s) CO₂ plays in regulating global climate lends insight into understanding the dynamics of the biosphere in the geologic rock record and aids in predicting future climate change.

This study is divided into four discrete chapters. In Chapter One I analyze a compilation of published stomatal responses (both stomatal density and stomatal index) to changes in atmospheric CO₂. Several confounding factors that may influence the stomata-CO₂ relationship have been suggested in the past, and I discuss these in light of this new compilation. The general reliability of this relationship in terms of its application into the geologic past as a CO₂ proxy is discussed in detail. In addition, this chapter serves as a general introduction into stomatal densities and stomatal indices and why they respond to changes in atmospheric CO₂.

Chapter Two is a paleoecological study of Tertiary *Ginkgo*. This ecological dataset includes most of the sites for which I measured stomatal indices from *Ginkgo* cuticle. In addition to some peculiar ecological patterns that may shed light on pre-angiospermous Mesozoic paleoecology, this study provides important constraints on the reliability of *Ginkgo* as a paleo-CO₂ indicator. These constraints are discussed more fully in the final two chapters.

In Chapter Three I present the training sets of *Ginkgo biloba* and *Metasequoia glyptostroboides*. These data stem both from dated herbaria sheets from the last 145 years where CO₂ has risen exponentially from 280 ppmV to (as of 2001) 370 ppmV, and from plant saplings growing in CO₂-controlled greenhouses ranging from 350 ppmV to 800 ppmV. I then inverted the regressions in these training sets and applied them to measurements of SI from fossil *Ginkgo* and *Metasequoia* dating to the early Paleogene (58-53 Ma) and middle Miocene (17-15 Ma). I conclude by discussing the implications of my CO₂ reconstruction for these two intervals, both of which are considered to represent periods of global warmth relative to today.

In Chapter Four I expand on both the data of Chapter Three and its implications for ancient and future climates. I present SI data back to the latest Cretaceous (66 Ma), and then discuss how the combined CO₂ reconstruction bears on the role CO₂ played during these globally warm periods.

CHAPTER 1

Stomatal density and stomatal index as indicators of paleoatmospheric CO₂ concentration

1.1 Introduction

The increase in atmospheric CO₂ concentration since industrialization (Friedli et al., 1986; Keeling et al., 1995) and the predicted continued increase into the near future (Houghton et al., 1995) forces the need to understand how the biosphere operates under elevated (relative to pre-industrial) CO₂ levels. The geologic record affords a wealth of such information. Fundamental to the use of the geologic record, however, is a reliable estimate of CO₂ concentration throughout the intervals of interest. The results of a computer-based model for the Phanerozoic (Berner, 1994; see Fig. 1.1), based on rates of Ca-Mg silicate weathering and burial as carbonates, weathering and burial of organic carbon, CO₂ degassing, vascular land plant evolution, and solar radiation, have gained wide acceptance (e.g., Retallack, 1997; Kump et al., 1999). Proxy data are still crucial, however, for both testing and refining this model. Currently used proxies include $\delta^{13}\text{C}$ from pedogenic carbonates (Cerling, 1991, 1992; Mora et al., 1991, 1996; Ekart et al., 1999), $\delta^{13}\text{C}$ from trace carbonates contained within goethite (Yapp and Poths, 1992, 1996), $\delta^{13}\text{C}$ from phytoplankton (Freeman and Hayes, 1992; Pagani et al., 1999a, 1999b), and $\delta^{11}\text{B}$ from planktonic foraminifera (Pearson and Palmer, 1999). To a first approximation, these proxies largely support the model of Berner (1994) (Fig. 1.1). A

discrepancy exists during the late Carboniferous and early Permian between the pedogenic carbonate-derived data of Ekart et al. (1999) and the model of Berner (1994). However, this discrepancy disappears if the $\delta^{13}\text{C}$ values for marine carbonates of Popp et al. (1986) are used during this time interval instead of those of Veizer et al. (1999) in calculating CO_2 from the data of Ekart et al. (1999) (R.A. Berner, unpublished data; see Fig. 1.1B).

Another emerging proxy relies on the plant species-specific inverse relationship between atmospheric CO_2 concentration and stomatal density and/or stomatal index. Concerns have been raised regarding this method's reliability (Körner, 1988; Poole et al., 1996), and it is the purpose of this chapter to address these concerns via an extensive analysis of the literature. Analysis includes stomatal responses from fossil observations as well as short-term (experimental, natural CO_2 springs, altitudinal transects, and herbaria) observations, as responses from the latter category are often used to generate standard curves for estimating CO_2 from fossil observations (van der Burgh et al., 1993; Beerling et al., 1995; Kürschner, 1996; Kürschner et al., 1996; Rundgren and Beerling, 1999; Wagner et al., 1999). Specifically, the utility of stomatal indices will be examined, an approach not analyzed in previous reviews (Beerling and Chaloner, 1992, 1994; Woodward and Kelly, 1995).

1.2 Mechanism controlling stomatal density

Stomata are pores on leaf surfaces through which plants exchange CO_2 , water vapor, and other constituents with the atmosphere. They form early in leaf development, and typically mature by the time the leaf reaches 10-60% of its final leaf size (Tichá,

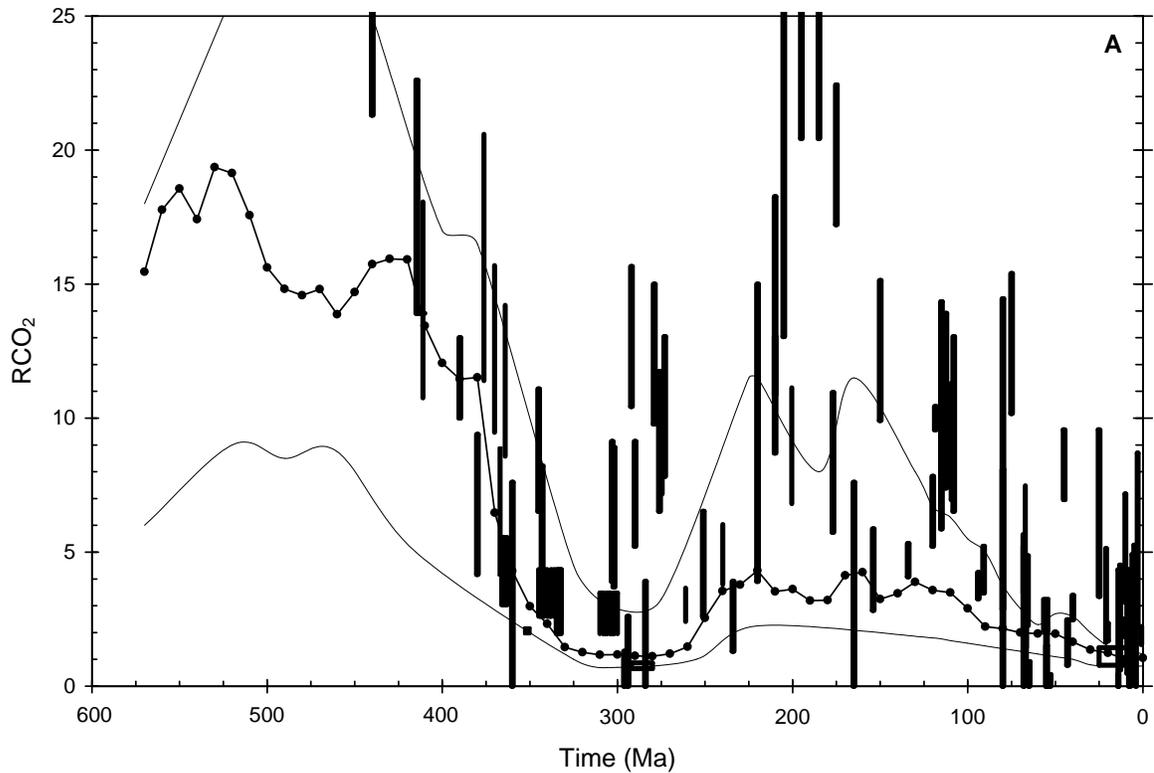


Figure 1.1. Atmospheric CO₂ versus time for the Phanerozoic. RCO₂ = ratio of mass of paleo-CO₂ to time-averaged pre-industrial value (230 ppmV, the mean CO₂ over at least the last 400 k.y. (Petit et al., 1999)). The centerline joining filled circles (10 m.y. time steps) represents the best estimate from the model of Berner (1994, 1998). The two straddling lines represent error estimates based on sensitivity analyses. Boxes in (A) represent 100 non-stomatal-based proxy estimates of varying RCO₂ resolution (data from Suchocky et al., 1988; Platt, 1989; Cerling, 1991, 1992; Freeman and Hayes, 1992; Koch et al., 1992; Muchez et al., 1993; Sinha and Stott, 1994; Andrews et al., 1995; Ghosh et al., 1995; Mora et al., 1996; Yapp and Poths, 1996; King, 1998; Ekart et al., 1999; Elick et al., 1999; Lee, 1999; Lee and Hisada, 1999; Pagani et al., 1999a, 1999b; Pearson and Palmer, 1999; Cox et al., 2001; Ghosh et al., 2001).

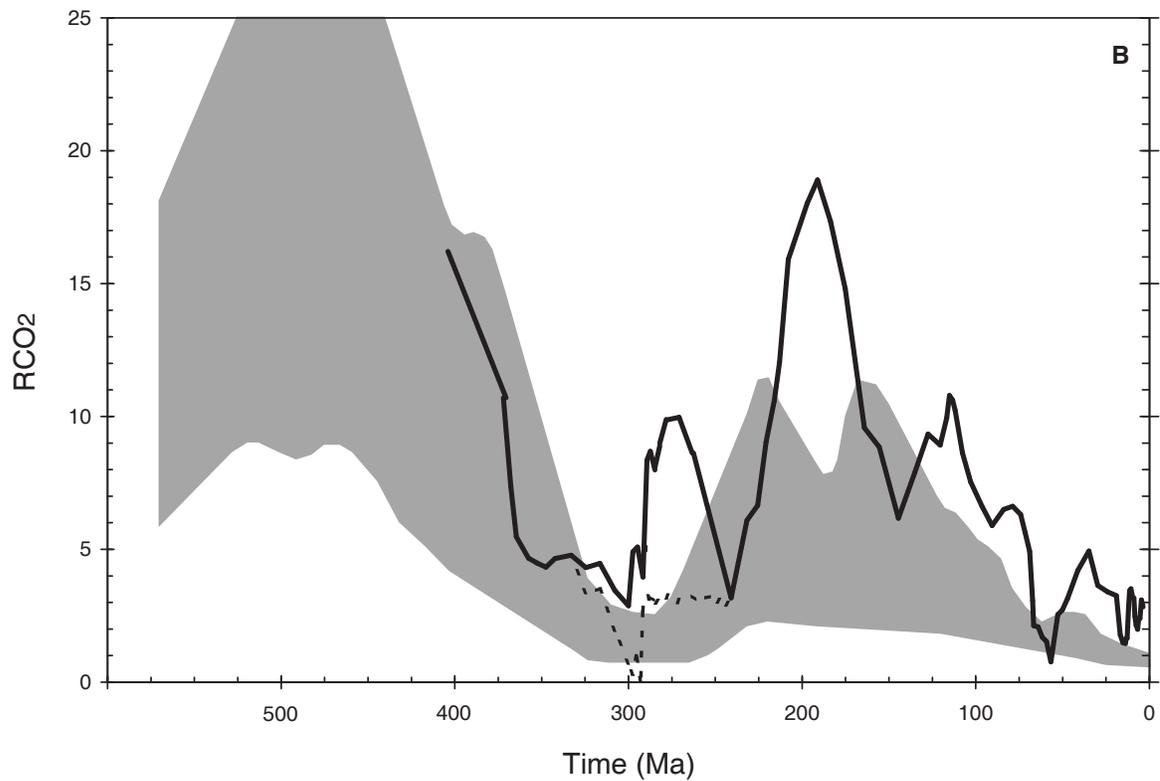


Fig. 1.1 con't. The heavy line in (B) is a five point running average of the mean RCO₂ of every box in (A). This approach smoothes short-term CO₂ fluctuations and is more directly comparable with the model of Berner (1994, 1998). The dashed line in (B) is a five point running average incorporating a recalculation of Ekart et al. (1999) data during the late Carboniferous and early Permian using the marine carbonate $\delta^{13}\text{C}$ data of Popp et al. (1986) (see text for details).

1982). Thus, the timing for the mechanism(s) of stomatal initiation lies early in leaf ontogeny (Gay and Hurd, 1975; Schoch et al., 1980). Currently, no mechanism or combination of mechanisms adequately explains the expression of stomatal initiation, although genetic work may provide insights in the near future (e.g., Berger and Altmann, 2000). Proposed mechanisms include irradiance (Gay and Hurd, 1975; Schoch et al., 1980), humidity (Salisbury, 1927), and PCO_2 (Woodward, 1986; Beerling and Chaloner, 1992; Woodward and Kelly, 1995; Beerling and Woodward, 1996).

A common theory for why CO_2 should (partially) control stomatal initiation is as follows (e.g., Woodward, 1987). Water vapor and CO_2 constitute the two main fluxes across the leaf epidermis. It is generally advantageous for plants to conserve water loss while maximizing CO_2 uptake, two typically antithetical processes. As CO_2 rises for a given water budget, for example, a plant can ‘afford’ to reduce its stomatal conductance without suffering a reduction in carbon assimilation rates. Two main pathways driving this response are smaller stomatal pores (Bettarini et al., 1998) and a reduction in stomatal numbers (Woodward, 1987). Conversely, a drop in CO_2 requires an increase in stomatal conductance to maintain assimilation rates, but at the cost of increased water loss.

1.2.1 Stomatal index

Stomatal density (SD) is a function of both the number of stomata plus the size of the epidermal cells. Thus, SD is affected both by the initiation of stomata and the expansion of epidermal cells. This expansion is a function of many variables (e.g., light, temperature, water status, position of leaf on crown, and intra-leaf position), and can overprint the signal reflective of stomatal initiation. As it turns out, CO_2 plays a stronger

role in stomatal initiation than in epidermal cell expansion (this is discussed in detail below). Salisbury (1927) introduced the concept of stomatal index (SI), which normalizes for the effects of this expansion (i.e., density of epidermal cells). It is defined as:

$$SI(\%) = \frac{\text{stomatal density}}{\text{stomatal density} + \text{epidermal cell density}} \times 100$$

where stomata consist of the stomatal pore and two flanking guard cells.

1.2.2 C₄ plants

The fundamental photosynthetic differences between C₃ and C₄ plants have consequences for stomatal-based CO₂ reconstructions. Carbon in C₃ plants is fixed within the spongy and palisade mesophyll where CO₂ concentrations (c_i) are approximately 70% of the atmospheric value. As atmospheric CO₂ fluctuates, so too does c_i to maintain this ~0.7 ratio (Polley et al., 1993; Ehleringer and Cerling, 1995; Beerling and Woodward, 1996; Bettarini et al., 1997). Thus the stomatal pore area is sensitive to changing atmospheric CO₂ levels. C₄ plants, in contrast, fix carbon within their bundle sheath cells. The endodermis enclosing these bundle sheath cells is highly impervious to CO₂, and consequently CO₂ concentrations within these cells can reach 1000-2000 ppmV (Lambers et al., 1998). One would therefore anticipate, based on the proposed mechanism between CO₂ and stomatal initiation discussed above, that even moderate changes in atmospheric CO₂ have little influence on stomatal pore area and, by extension, stomatal density and stomatal index (Raven and Ramsden, 1988). Of the 9 responses derived from C₄ plants documented here, only 1 inversely responds to CO₂ (see Appendix 1.1). This marked insensitivity in C₄ plants lends indirect support for the proposed mechanism. Because of

the above physiological reasons, none of the analyses considered here include responses from C₄ plants.

1.3 Stomatal density and stomatal index as CO₂ indicators

A database consisting of 285 SD responses and 145 SI responses to variable CO₂ concentrations was compiled to elucidate salient patterns (Appendices 1.1-1.3). 176 species are represented. This database is an expansion of previous reviews (Beerling and Chaloner, 1994; Woodward and Kelly, 1995) and includes, for the first time, stomatal indices.

Each response was first placed in one of three categories: experimental, subfossil, and fossil. Experimental responses stem from experimentally controlled CO₂ environments, typically in greenhouses, which last from 14 days to 5 years in length. For studies that measured SD and/or SI at several different times and/or CO₂ levels, typically only the response corresponding to the longest exposure time and highest CO₂ level was used. Most subfossil responses stem from dated herbarium specimens (from the last 240 years), where corresponding CO₂ concentrations are known from ice core data (Neftel et al., 1985; Friedli et al., 1986). Data from altitudinal transects and natural CO₂ springs are also placed in the subfossil category, as this category represents the closest match in terms of CO₂ exposure time. Finally, fossil responses consist of well-dated fossil material. Methods for obtaining reference CO₂ concentrations for the fossil responses are discussed below.

Each response was assigned as either increasing ($P < 0.05$), decreasing ($P < 0.05$), or remaining the same ($P > 0.05$) relative to controls. Where P -values were not reported, a test for overlapping standard deviations was used, which typically yields a conservative estimate for statistical significance (relative to the $\alpha = 0.05$ level).

1.3.1 Experimental responses

127 SD and 74 SI responses from a pool of 68 species are represented here. For stomatal density, 40% of the experimental responses inversely respond (at the $\alpha = 0.05$ level) to CO₂; the proportion for stomatal indices is similar (36%) (Table 1.1).

Plants exposed to subambient CO₂ are more likely to inversely respond than plants exposed to elevated CO₂ for both SD (50% vs. 39%; $P = 0.36$) and SI (89% vs. 29%; $P < 0.001$). These results support previous claims that plants more strongly express the CO₂-SD/SI inverse relationship when exposed to subambient versus elevated CO₂ concentrations (Woodward, 1987; Woodward and Bazzaz, 1988; Beerling and Chaloner, 1993a; Kürschner et al., 1997). A common explanation for this CO₂ ‘ceiling’ phenomenon is that plants today have not experienced elevated CO₂ levels (350+ ppmV) for at least the entire Quaternary and possibly longer (Pagani et al., 1999a; Pearson and Palmer, 1999). Thus, for short time scales where only plant plasticity is tested, plants respond more favorably to CO₂ conditions which they most recently experienced, namely subambient concentrations (Woodward, 1988; Beerling and Chaloner, 1993a). The implication for stomatal-based CO₂ reconstructions is that experimental evidence based on elevated CO₂ treatments may not reflect the reliability of the method. Over 85% of the experimental

Table 1.1
Statistical summary of stomatal responses to changing CO₂ concentrations

	Experimental				Subfossil				Fossil				Combined			
	% ^c	SD ^a (n)	%	SI ^b (n)	%	SD (n)	%	SI (n)	%	SD (n)	%	SI (n)	%	SD (n)	%	SI (n)
total	40	(127)	36	(74)	50	(133)	34	(35)	88	(25)	94	(36)	49	(285)	50	(145)
elevated CO ₂ ^d	39	(109)	29	(65)	-	-	-	-	100	(13)	96	(24)	45	(232)	41	(116)
subambient CO ₂	50	(18)	89	(9)	-	-	-	-	89	(9)	89	(9)	75	(40)	88	(25)
opposite response ^e	9	(127)	4	(74)	11	(133)	9	(35)	12	(25)	3	(36)	11	(285)	5	(145)
hypostomatous ^f	59	(27)	65	(17)	50	(80)	38	(24)	-	-	-	-	56	(121)	69	(70)
amphistomatous ^g	36	(90)	27	(55)	49	(49)	25	(8)	-	-	-	-	44	(149)	32	(71)
abaxial	40	(45)	24	(29)	41	(22)	-	-	-	-	-	-	41	(68)	21	(33)
adaxial	29	(42)	31	(26)	55	(22)	-	-	-	-	-	-	38	(65)	33	(30)
experiments using OTCs ^h	13	(31)	13	(24)	-	-	-	-	-	-	-	-	-	-	-	-
experiments using greenhouses	48	(95)	48	(50)	-	-	-	-	-	-	-	-	-	-	-	-
herbarium studies only	- ^j	-	-	-	57	(93)	89	(9)	-	-	-	-	-	-	-	-
repeated species ⁱ	-	-	-	-	-	-	-	-	-	-	-	-	57	(28)	55	(11)

^a stomatal density

^b stomatal index

^c percentage of responses inversely correlating with CO₂

^d CO₂ concentrations are higher than controls

^e percentage of responses positively correlating with CO₂

^f leaves with stomata only on abaxial (lower) side

^g leaves with stomata on both surfaces

^h OTC = open-top chamber; typically cone-shaped with an open top

ⁱ for species with multiple responses with ≥1 inversely correlating with CO₂, percentage that consistently inversely correlate

^j not applicable or sample size too small for meaningful comparison

responses analyzed here stem from elevated CO₂ treatments. Another related concern raised with experimental results is that CO₂ is shifted in one step in contrast to the smoother, longer-term trend in nature (Beerling and Chaloner, 1992; Kürschner et al., 1997).

An alternative explanation for the CO₂ ceiling is that while CO₂ is limiting for photosynthesis at CO₂ concentrations below present day levels, it is not limiting at elevated levels. Therefore, for example, if CO₂ decreases in a subambient CO₂ regime (where CO₂ is limiting for photosynthesis), a mechanism exists to increase stomatal pore area and, by extension, CO₂ uptake. The same may not be true at elevated CO₂ concentrations if CO₂ is not limiting for photosynthesis under such conditions (Wagner et al., 1996; Kürschner et al., 1998). Empirical data do not strongly support this alternative hypothesis. While assimilation rates generally decrease at subambient CO₂ levels (Polley et al., 1992; Robinson, 1994), they also typically increase in response to CO₂ concentrations of at least 700 ppmV (Long et al., 1996; Curtis and Wang, 1998). CO₂ therefore usually continues to limit photosynthesis in most plants above present day CO₂ levels, even if the effects of this excess CO₂ are partially mediated by a reduction in photorespiration and enhancement in RuBP regeneration (the primary substrate used to fix CO₂ in C₃ plants), and so only affect photosynthesis indirectly. Therefore, there is no reason to expect a CO₂ ceiling coincident with current CO₂ levels. It is likely, however, that the rate of change in assimilation rates is reduced at elevated CO₂ concentrations (Farquhar et al., 1980), which could reduce the sensitivity of SD and SI responses under such conditions.

Experimental manipulations are usually conducted in either enclosed greenhouses or open-top chambers (OTCs). Most OTCs have less control over humidity and

temperature. Significant ‘chamber effects’ have been detected for stomatal parameters (Knapp et al., 1994; Apple et al., 2000), and results generated here support such claims. Plants in OTCs inversely respond to CO₂ in far fewer cases than greenhouse grown plants for both SD (13% vs. 48%; $P < 0.001$) and SI (13% vs. 48%; $P < 0.01$). Thus, it appears OTCs introduce confounding factors and should be avoided in stomatal density/index work.

Although the proportion of experimental responses inversely responding to CO₂ may appear low (40% and 36% for SD and SI, respectively), in part from the factors discussed above, it is important to note that the percentage of responses showing a positive relationship ($P < 0.05$) is very low (9% and 4% for SD and SI, respectively). Thus, the vast majority of plants either respond inversely to experimental exposure to CO₂ or do not respond at all.

1.3.2 Subfossil responses

133 SD and 35 SI responses from a pool of 95 species are represented here. For stomatal density, 50% of the subfossil responses inversely relate (at the $\alpha = 0.05$ level) to CO₂. Thus, subfossil responses, which are based on longer exposure times, more often inversely relate to CO₂ than do experimental responses (50% vs. 40%; $P = 0.11$). For stomatal index, only 34% of the responses show a significant inverse relationship, however the sample size is disproportionally small ($n = 35$) (Table 1.1).

As outlined above, three types of studies comprise the subfossil responses: altitudinal transects, natural CO₂ springs, and herbaria. If only herbarium responses are analyzed ($n = 93$ and $n = 9$ for SD and SI, respectively), the proportion showing an

inverse response to CO₂ improves to 57% and 89%, respectively. Responses from altitudinal transects and natural CO₂ springs may therefore be of less value for paleo-CO₂ reconstructions. This dichotomy in response fidelity may be an expression of the CO₂ ceiling phenomenon discussed above. As CO₂ levels rose to current levels over the last 240+ years, the majority of plants (57% and 89% for SD and SI, respectively) responded with significant decreases in SD and/or SI. At higher CO₂ levels, however, as expressed near natural CO₂ springs, a smaller proportion of plants responded with lower SD (30%; $n = 30$) and/or SI (16%; $n = 25$). If, on the other hand, current CO₂ concentrations do *not* represent a true genetic ceiling for plants, than these data show that the majority of plants cannot adapt to CO₂ levels above today's within the special residence time near natural CO₂ springs (10^2 - 10^3 years?).

In accordance with the experimental responses, a very small proportion of the subfossil observations positively respond to CO₂ (11% and 9% for SD and SI, respectively). Most plants either inversely respond to CO₂ or do not respond at all. If CO₂ exerts any influence on stomatal initiation, it must be of an inverse behavior.

1.3.3 Fossil responses

25 SD and 36 SI responses from a pool of 28 species are represented here. For stomatal density, 88% of the observations show an inverse relationship (at the $\alpha = 0.05$ level) to CO₂; for stomatal index, the proportion is 94% (Table 1.1). Only 12% and 3% of the observations positively respond to CO₂ for SD and SI, respectively. Thus, the robustness of the SD/SI method improves with increased CO₂ exposure time (Fig. 1.2), supporting earlier hypotheses (Beerling and Chaloner, 1992, 1993a).

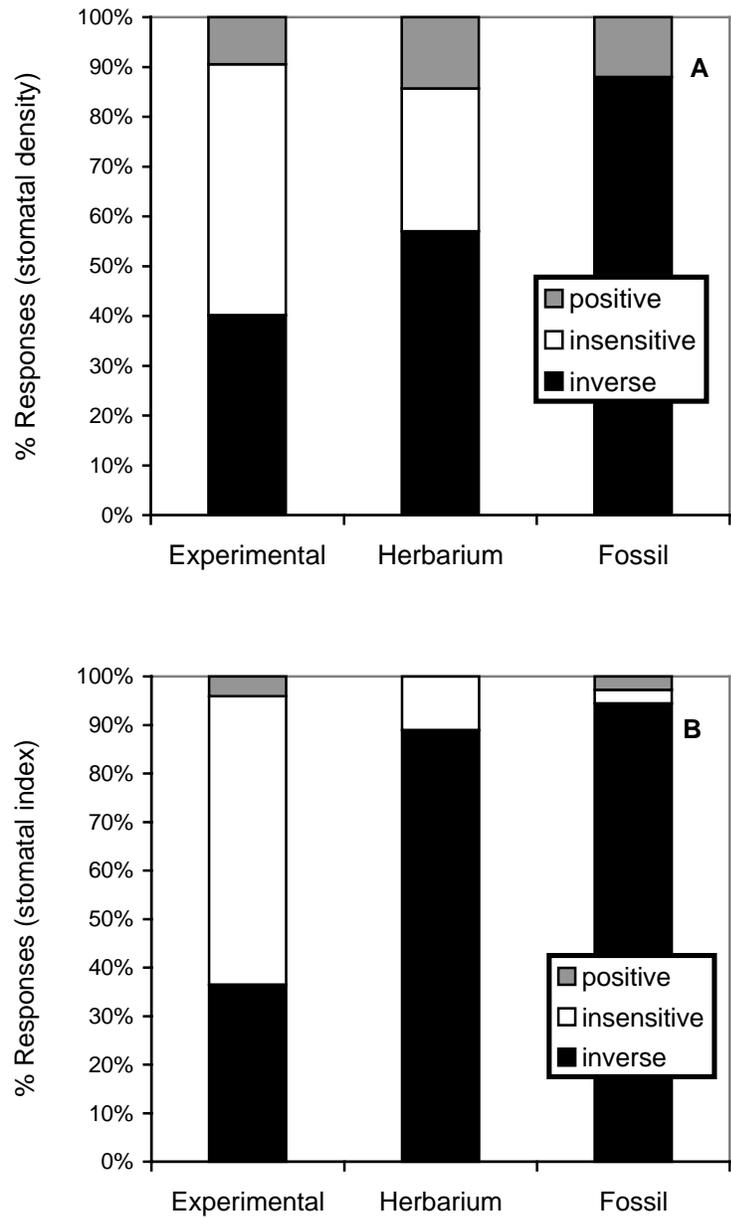


Figure 1.2. The percentage of responses for (A) stomatal density and (B) stomatal index that inversely relate to CO₂ ('inverse'), show no significant change to CO₂ ('insensitive'), or response positively to CO₂ ('positive') in each of three categories. Note that only herbarium responses compose the subfossil category.

Qualitatively, the transition between dominance of stomatal response by plasticity within a given gene pool and genetic adaptation appears to occur for most plants between 10^2 and 10^3 years (i.e., intermediate between CO_2 exposure times typical for subfossil and fossil responses). This conclusion hinges on the assumption that CO_2 exerts a consistent genetic pressure on stomatal initiation, and given sufficient exposure time will overprint the smaller scale plastic responses (including changes in individual stomatal pore size). The fact that the increase in responses showing an inverse relationship to CO_2 as a function of exposure time comes at the expense of insensitive responses (Fig. 1.2) supports this assumption. 10^2 to 10^3 years is slightly longer than previous estimates (Beerling and Chaloner, 1993a), and should give rise to some caution in using experimental and subfossil responses in paleo- CO_2 reconstructions (i.e., comparing responses due mainly to plasticity versus genetic adaptation).

The fossil data cast doubt on the notion that stomata cannot respond to CO_2 concentrations above present day levels. The proportion of fossil responses showing an inverse relationship based on subambient CO_2 exposure are nearly equal to those fossil observations based on elevated CO_2 exposure for both SD (89% and 100%, respectively) and SI (89% and 96%, respectively), although sample sizes are fairly small (Table 1.1). Some groups of plants respond to CO_2 levels of at least 2700 ppmV (McElwain and Chaloner, 1995; Appendix 1.3). This result does not discount, however, that stomatal parameters may be less *sensitive* at elevated than at subambient (relative to today) CO_2 levels. The CO_2 ceiling observed in experimental responses therefore appear to stem from the short-term inability of plants to respond to elevated CO_2 , not a long-term genetic limit. Interestingly, Woodward (1988) noted that plants with short generation times (e.g., annuals) are often capable of decreasing their stomatal densities when experimentally

exposed to elevated CO₂ levels (for ≥ 1 year), probably because of their quicker genetic adaptation rates (Woodward, 1988). This suggests that the exposure time required to mitigate the CO₂ ceiling may not be much beyond typical experimental exposure times, and in fact may not exist at all for some plants.

Caution is urged with regard to several features concerning the fossil responses. First, in several studies stomatal comparisons between fossil and modern plants were made with two separate but ecologically equivalent sets of species (McElwain and Chaloner, 1995, 1996; McElwain, 1998; McElwain et al., 1999). In addition to the long-term influence of CO₂ on SD and SI for a given species, it has also been shown, for example, that high CO₂ selects for groups of plants with lower mean stomatal densities/indices (Beerling and Woodward, 1996; Beerling and Woodward, 1997) (Fig. 1.3). Thus, it is not particularly surprising that stomatal densities and indices from times of high CO₂ are lower than for ecologically equivalent modern species. Ideally, these two effects should be kept separate.

Second, estimates of CO₂ for the fossil responses are invariably not as accurate as those estimates for experimental and subfossil responses. Ice core derived data are used for the last 150 k.y., and the model of Berner (1994) or other proxy data are most often used for pre-ice core responses. In particular, estimates from Berner's curve are highly approximate due to its sizable error envelope and coarse 10 m.y. time resolution (see Fig. 1.1); brief but large CO₂ excursions discernable with the various proxy methods are probably too temporally constrained to influence Berner's model (Montañez et al., 1999). In cases where experimental and subfossil responses are used to generate a standard curve upon which CO₂ concentrations are directly calculated from fossil responses, ice core data (Beerling et al., 1995; Wagner et al., 1999; Rundgren and Beerling, 1999) or the presence

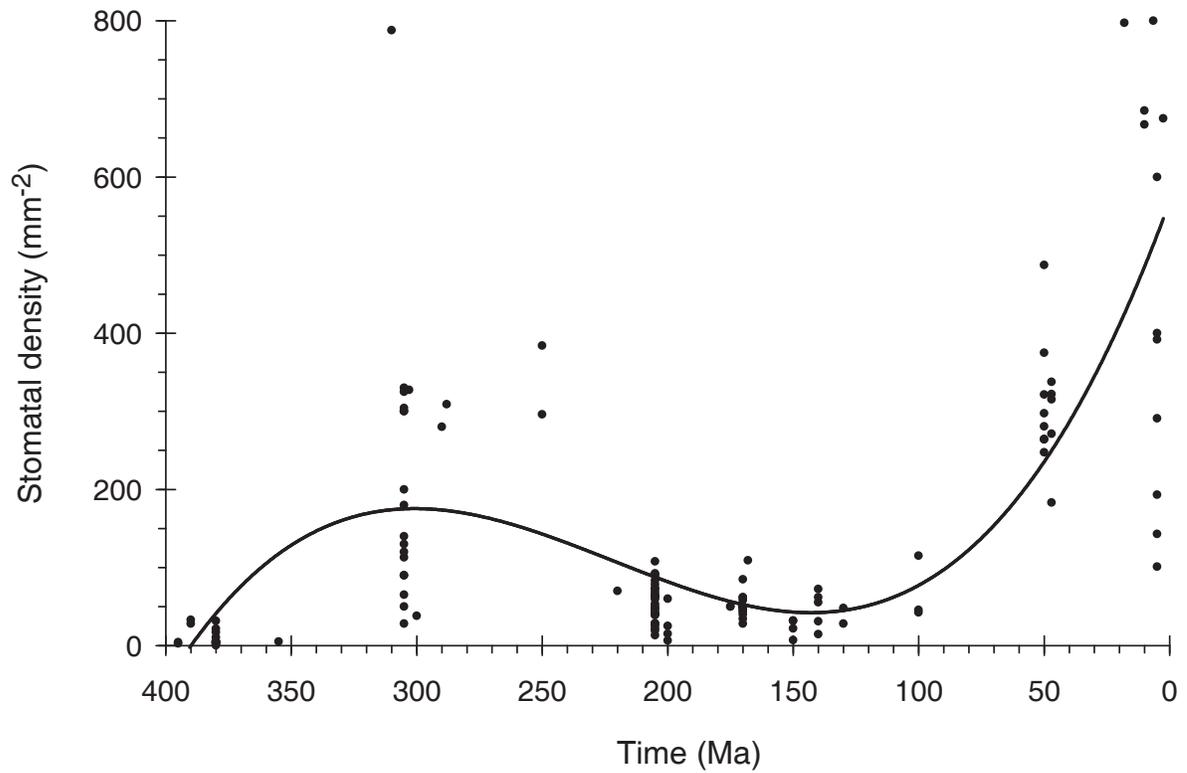


Fig. 1.3. Stomatal density versus time for the Phanerozoic. Redrawn from Beerling and Woodward (1997), with additional data plotted from McElwain and Chaloner (1996), Edwards et al. (1998), McElwain (1998), Cleal et al. (1999), and McElwain et al. (1999). Regression is a third order polynomial ($r^2 = 0.57$; $n = 132$). Compare trend with Fig. 1.1.

of temperature excursions (van der Burgh et al., 1993; Kürschner, 1996; Kürschner et al., 1996) are used to corroborate the stomatal-based estimates.

1.3.4 Combined data set

Based on the combination of the above three categories, both SD and SI inversely correlate with CO₂ c. 50% of the time ($n = 285$ and 145 , respectively) (Table 1.1). Very rarely do the responses positively correlate with CO₂ (11% and 5% for SD and SI, respectively). For species that have been analyzed repeatedly by different researchers, those that inversely respond to CO₂ tend always to respond in such a way (57% ($n = 28$) and 55% ($n = 11$) for SD and SI, respectively). Woodward and Kelly (1995) reported a similar behavior, where 76% of their sensitive species consistently responded.

Thus, although response times differ (see above and Fig. 1.2), CO₂ is highly negatively correlated with stomatal initiation. A scatterplot of all data shows an overall inverse relationship between stomatal density/index and CO₂ (Fig. 1.4A). Although the overall regression is not robust ($r^2 = 0.26$; $n = 420$), this principally stems from equivocal experimental and natural CO₂ spring data. The fossil data, when regressed independently, yield an r^2 of 0.68 ($n = 59$) (Fig. 1.4B). Given the species-specific and probable multi-mechanistic nature of the relationship, this regression is surprisingly robust.

Curiously, based solely on the combined results, it appears SD is equally reliable as SI as a CO₂ indicator (Table 1.1). The implications are tempting, as epidermal cells are often difficult to resolve in fossil material (Beerling et al., 1991; McElwain and Chaloner, 1996). This issue is discussed in the section below.

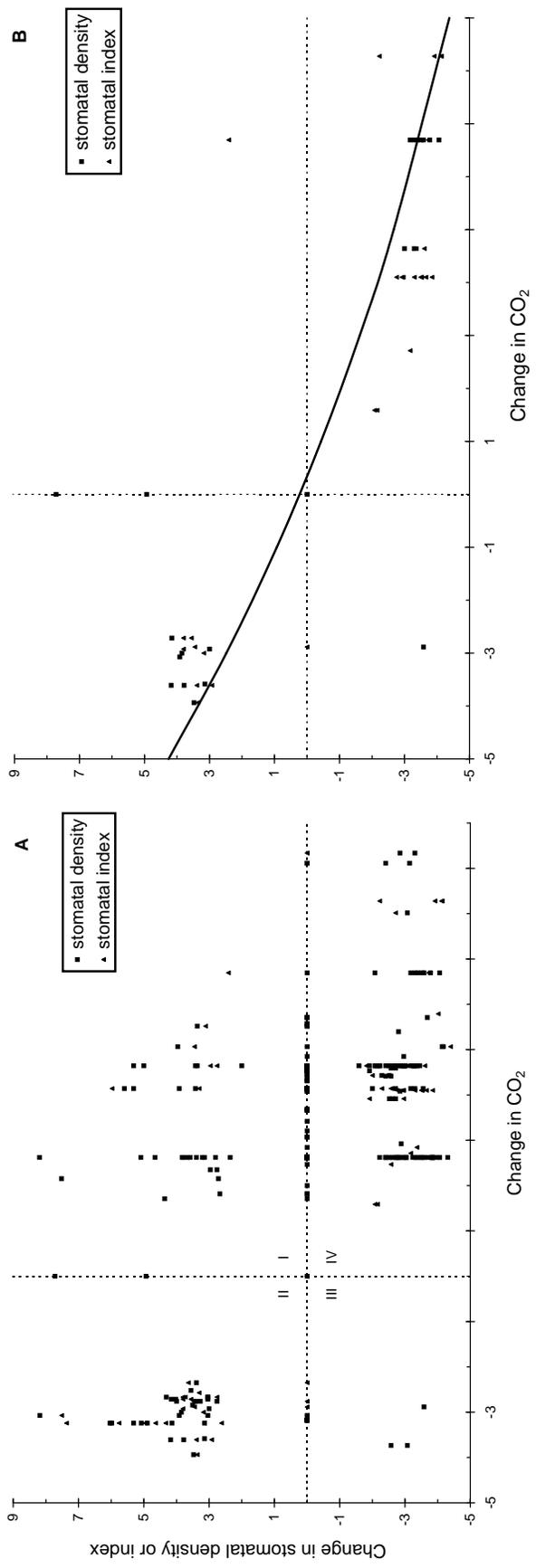


Fig. 1.4. (A) Scatterplot of all data ($r^2 = 0.26$; $n = 420$) showing the cube-root transform of percentage change in stomatal density and stomatal index in response to percentage change in atmospheric CO₂ concentration. Responses in quadrants II and IV inversely relate to CO₂ while responses in quadrants I and III positively relate. (B) Similar scatterplot for fossil data only. Regression equation of untransformed data: $y = 112.43 \times \exp(-0.0026x) - 100$. ($r^2 = 0.68$; $n = 59$).

Most vascular land plants have stomata on either both surfaces (amphistomatous) or only the abaxial (lower) surface (hypostomatous). Woodward and Kelly (1995) reported no strong differences in responses between the two leaf types, although in experimental responses amphistomatous species appeared more likely to inversely relate to CO₂. Results here indicate hypostomatous species more often inversely respond to CO₂ for both SD (56% vs. 44%; $P < 0.03$) and SI (69% vs. 32%; $P < 0.001$). For amphistomatous species, neither the abaxial nor adaxial (upper) surface yield statistically different responses (Table 1.1).

1.4 Potential confounding factors

CO₂ is likely not the sole factor determining stomatal density and stomatal index. As discussed above, SD is sensitive to both stomatal initiation and epidermal cell expansion, while SI is sensitive only to stomatal initiation. The influence of natural variability, water stress, irradiance, temperature, and other factors on stomatal parameters are briefly discussed below. More thorough reviews are given by Salisbury (1927), Tichá (1982) and Woodward and Kelly (1995).

1.4.1 Natural variability

In general, stomatal density increases from leaf base to tip (Salisbury, 1927; Sharma and Dunn, 1968, 1969; Tichá, 1982; Smith et al., 1989; Ferris et al., 1996; Zacchini et al., 1997; Stancato et al., 1999). SD also often increases from leaf midrib to

margin (Salisbury, 1927; Sharma and Dunn, 1968; Smith et al., 1989), although sometimes the differences are not significant (Sharma and Dunn, 1969; Tichá 1982). In contrast, very little intra-leaf variation in SI is present (Salisbury, 1927; Rowson, 1946; Sharma and Dunn, 1968, 1969; Rahim and Fordham, 1991), although Poole et al. (1996) found significant intra-leaf variation in *Alnus glutinosa*. For amphistomatous species, the distribution of stomata are generally more uniform on the abaxial surface (Rowson, 1946; Sharma and Dunn, 1968, 1969), and so for all species typically the mid-lamina of the abaxial surface yields the least variation.

Stomatal density also increases from the basal to distal regions of the plant (Salisbury, 1927; Gay and Hurd, 1975; Tichá, 1982; Oberbauer and Strain, 1986; Zacchini et al., 1997), primarily as a consequence of decreased water potential. Decreased water potential stimulates xerophytic traits, which include smaller epidermal cells, which in turn promote closer packing of stomata, and thus increased stomatal density. Little effect is reported for SI (Rowson, 1946), although evergreen species may exhibit a significant gradient (W.M. Kürschner, personal communication, 2000). Conflated with this trend are the differences between sun and shade leaves. Again, SD is consistently higher in sun leaves while SI values remain conservative (Salisbury, 1927; Poole et al., 1996; Kürschner, 1997; Wagner, 1998) with the exception of the study of Poole et al. (1996), who found a small 7% decline in SI for shade versus sun leaves. For fossil studies, since sun leaves in allochthonous assemblages are preferentially preserved (Spicer, 1981), this issue is often not a concern even for SD-based work. For example, Kürschner (1997) observed that 90% of his Miocene *Quercus pseudocastanea* leaves were sun morphotypes.

1.4.2 Water stress

In general, water stress correlates with increased SD (Salisbury, 1927; Sharma and Dunn, 1968, 1969; Tichá, 1982; Abrams, 1994; Estiarte et al., 1994; Clifford et al., 1995; Heckenberger et al., 1998; Pääkkönen et al., 1998). Some studies, however, report no response (Estiarte et al., 1994; Dixon et al., 1995; Pritchard et al., 1998; Centritto et al., 1999). No studies report a decrease. SI consistently appears insensitive to water stress (Salisbury, 1927; Sharma and Dunn, 1968, 1969; Estiarte et al., 1994; Clifford et al., 1995).

Salisbury (1927) proposed humidity as a mechanism for controlling stomatal initiation, and thus SI. Increased humidity slightly increased SI ($P > 0.05$) for *Scilla nutans*, however Tichá (1982) concluded that humidity may actually reduce stomatal index. Sharma and Dunn (1968, 1969) found no effect. Thus, the current data are equivocal.

1.4.3 Irradiance

Not surprisingly, light intensity usually positively correlates with SD (Sharma and Dunn, 1968, 1969; Gay and Hurd, 1975; Tichá, 1982; Oberbauer and Strain, 1986; Solárová and Pospíšilová, 1988; Stewart and Hoddinott, 1993; Ashton and Berlyn, 1994; Furukawa, 1997; Zacchini et al., 1997). This response is driven (partially) by enhanced water stress. Light intensity may also positively affect stomatal index (Sharma and Dunn, 1968, 1969; Furukawa, 1997), although some report no response (Salisbury, 1927; Sharma and Dunn, 1968, 1969) and Rahim and Fordham (1991) observed a small

decrease with increasing irradiance. In the case of Sharma and Dunn (1968, 1969), they speculated that the low irradiance levels required to depress SI could not sustain plants in a competitive environment.

In experimental manipulations, photoperiod strongly affects both SD and SI (Schoch et al., 1980; Zacchini et al., 1997). Schoch et al. (1980) observed that even one day of low irradiance levels during the critical period of stomatal initiation could affect SD and SI. Given that SI is typically conservative in deciduous species within a given crown, it is possible the effects of photoperiod on SI observed in experiments are minimal in nature.

1.4.4 Temperature

Temperature appears positively correlated with stomatal density (Ferris et al., 1996; Reddy et al., 1998; Wagner, 1998; but see Apple et al., 2000), a likely consequence of enhanced water stress. Temperature may also affect stomatal index (Ferris et al., 1996; Wagner, 1998), suggesting an influence on stomatal initiation. Reddy et al. (1998), however, found no response. The influence of temperature on stomatal initiation may be inconsequential, though, as most plants partially normalize for fluctuating temperatures by adjusting the timing of leaf development, and so the temperature at which stomata form remains fairly constant (Wagner, 1998).

1.4.5 Canopy CO₂ gradient

If CO₂ concentrations within canopies deviate significantly from ambient concentrations, CO₂ estimates based on stomatal parameters could be skewed. Empirical evidence, however, does not suggest such large deviations. Hourly measurements of CO₂ at 8 different heights (0.3, 0.8, 4.5, 7.5, 12.7, 18.3, 24.1, and 29.0 m above the ground surface) have been recorded for several consecutive years from an atmospheric tower in the Harvard Forest (data available at <http://www-as.harvard.edu/chemistry/hf/profile/profile.html>). This forest, in north central Massachusetts, USA, consists of mixed hardwoods and conifers. As shown in Fig. 1.5A, above 4.5 m, where the bulk of leaves from mature trees form, canopy CO₂ levels are virtually indistinguishable from ambient levels. Furthermore, all deviations diminish during the middle of the day (Fig. 1.5B), a period when cell respiration and division is highest. Thus, CO₂ gradients within canopies are likely not strong enough to influence stomatal initiation rates.

1.4.6 Paleotaxonomy

Paleobotanical species identification via morphological comparison with modern representatives is often tenuous, particularly with pre-Neogene material. There are methods, however, to bolster confidence in such morphologically-based species identification. These include comparing the sedimentological and ecological contexts with the proposed extant representative. For example, if a strictly swamp margin fossil species is morphologically identical to a modern representative that is also restricted to swamp

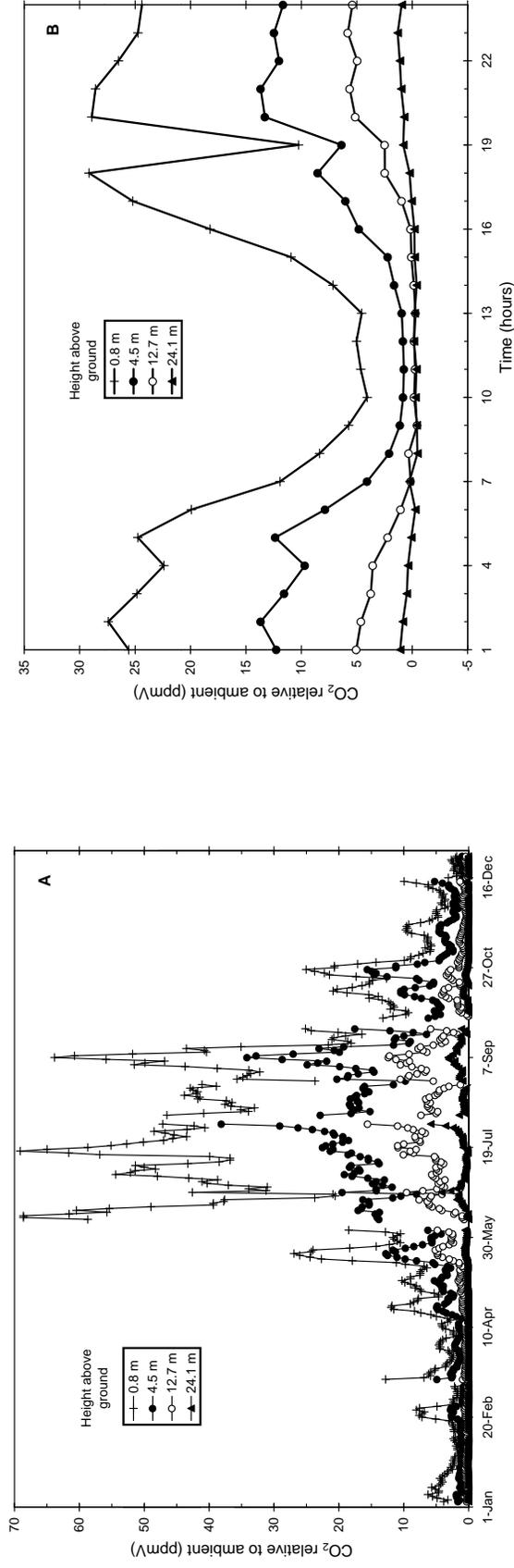


Fig. 1.5. (A) CO₂ relative to ambient concentrations for four heights within a tree canopy in 1996. Canopy height is c. 24 m. Ordinate represents seven day running average of daily averages of hourly measurements at each height ($n = 5311$ for each height). Measurements at 29.0 m height taken as ambient value (mean for time interval at this height = 370 ppmV). (B) Diurnal trend of CO₂ relative to ambient concentrations (data from 9 April-13 July 1996). Ordinate represents mean for each hour at each height ($n = 1388$ for each height). Standard errors approximate size of symbols. Raw data used with permission of S. Wofsy.

margins, then one can be more confident that the two are identical species. Independent of species identification, however, it is also possible that a single species may develop, for example, different stomatal behaviors through time. This, in turn, could affect paleo-CO₂ reconstructions. One way to circumvent this problem is through the study of the species' closest extant sister group (e.g., de Queiroz and Gauthier, 1990). If the stomata in both extant species show a similar response to CO₂, then it can be assumed that this stomatal behavior in both species is conservative in time back to at least when the species branched.

1.4.7 Other potential confounding factors

Through the comparison of 100 species, neither growth form (woodiness vs. non-woodiness; trees vs. shrubs), habitat (cool vs. warm), nor taxonomic relatedness strongly correlated with SD (Woodward and Kelly, 1995). Habitat has also been found not to affect SI (Rowson, 1946). Analysis of the data set presented here shows that for genera represented by >1 species, only 19% ($n = 16$) and 14% ($n = 7$) of these genera are internally consistent ($\alpha = 0.05$) with respect to SD and SI, respectively. These results provide further support for the taxonomic independence of stomatal responses to CO₂.

An increase in ploidy level is associated with lower SD (Rowson, 1946; Mishra, 1997). No clear trend is found in SI (based on two studies), with Rowson (1946) reporting a decrease and Mishra (1997) no change. Given the widespread variability of ploidy levels in the fossil record (Masterson, 1994), this may have important consequences for stomatal-based CO₂ reconstructions.

Elevated levels of ozone increase SD in *Betula pendula* (Frey et al., 1996; Pääkkönen et al., 1998), *Fraxinus excelsior* (Wiltshire et al., 1996) and *Olea europaea* (Minnocci et al., 1999). Effects on SI were not reported.

Although largely untested, atmospheric oxygen may influence SD and SI. Elevated O₂/CO₂ ratios increase photorespiration in C₃ plants, suppressing CO₂ assimilation rates. One pathway to mediate this decline is increasing SD and/or SI. Experimental work on *Hedera helix* and *Betula pubescens* show slightly higher stomatal indices in a 35% versus 21% O₂ atmosphere (Beerling et al., 1998b). If correct, this factor may be particularly important during the Carboniferous and early Permian when O₂ concentrations are modeled to exceed 30% (Berner and Canfield, 1989; Berner et al., 2000).

1.5 Summary

Based on the data presented here, nearly all species appear responsive on the time scales inherent in most fossil CO₂ reconstructions (>10² years) (Fig. 1.2; Table 1.1). Only 40-50% of species are responsive over the time scales of experimental and subfossil studies (~10⁻²-10² years), and so those conducting studies requiring such responses must take care in choosing sensitive species (Appendices 1.1-1.2). Another potential weakness in utilizing experimental and subfossil responses is that they are more reflective of plasticity within given gene pools, and may display different behaviors than their respective fossil responses (which are more reflective of genetic adaptation).

SD and SI are both equally likely to inversely relate to CO₂. Stomatal density, however, is sensitive to factors affecting cell expansion such as water stress, temperature, and irradiation. Stomatal index, in contrast, is sensitive only to factors affecting cell initiation, of which CO₂ appears to be one factor. Thus, even if SD and SI show similar responses for a given species (e.g., both positive or negative), stomatal index should yield more accurate CO₂ estimates.

1.5.1 Applications of method

Although experimental work has been carried out for many years, Woodward (1987) was the first to document the inverse CO₂-SD/SI relationship over longer time scales (200 years). Beerling et al. (1991, 1993) extended the applicability of the method to 140 k.y. with *Salix herbacea*, where stomatal densities showed a general inverse relationship with ice core reconstructed CO₂ concentrations. This method has also proven successful with 9.2 ka *Salix cinerea* (McElwain et al., 1995), 13 ka *Betula nana* (Beerling, 1993), and 28 ka *Pinus flexilis* (van de Water et al., 1994).

While the above studies validate the relationship over time scales useful for fossil studies, they do not generate independent estimates of paleo-CO₂. For this, fossil responses must be fitted to a standard curve based on experimental, subfossil, and fossil responses (from the last 400 k.y., for which ice core data exist; e.g., Petit et al., 1999) of the *same* species. This approach has been successful in the Holocene with *Salix herbacea* (Beerling and Chaloner, 1993a; Beerling et al., 1995; Rundgren and Beerling, 1999) and *Betula pubescens* and *B. pendula* (Wagner et al., 1999), and in the Miocene with *Quercus petraea* and *Betula subpubescens* (van der Burgh et al., 1993; Kürschner, 1996; Kürschner

et al., 1996). While this approach produces the most accurate CO₂ reconstructions, it is limited by its requirement to compare identical species (or highly similar species within a genus; Wagner et al., 1999). There are, however, single species spanning most or all of the late Cretaceous and Tertiary (e.g., *Ginkgo adiantoides/biloba*, several members of Taxodiaceae), and so CO₂ estimates for this interval are possible.

One clear advantage of the stomatal method over other proxies and models is its high temporal resolution. The temporal resolution of late Quaternary fossil material often exceeds that of ice cores (Beerling et al., 1995; Wagner et al., 1999), and similar high resolution data have been used to document CO₂ excursions across the Allerød/Younger Dryas (Beerling et al., 1995) and Triassic/Jurassic (McElwain et al., 1999) boundaries. Another advantage over other proxies and models is its higher level of precision (compare Fig. 1.1A with Fig. 1.6).

Estimating pre-Cretaceous CO₂ levels proves more difficult. McElwain and Chaloner (1995) developed a technique comparing responses of fossil species to those of their Nearest Living Equivalents (NLEs). NLEs are defined ecologically, not taxonomically, and represent the ecologically closest living analog to the fossil species. SI ratios of the fossils:NLEs were calculated, and in order to estimate CO₂ the Carboniferous:NLE stomatal ratio was normalized to the Phanerozoic CO₂ curve of Berner (1994), with the remainder of the ratios scaled accordingly in a linear fashion. Given that this method assumes a linear response and is not a true independent CO₂ indicator, reconstructed CO₂ concentrations from the Devonian, Carboniferous, Permian, and Jurassic all matched Berner's values remarkably well (McElwain and Chaloner, 1995, 1996). Later (McElwain, 1998), in order to reduce the method's dependence on Berner (1994), data (including new material from the Eocene) were plotted assuming a 1:1

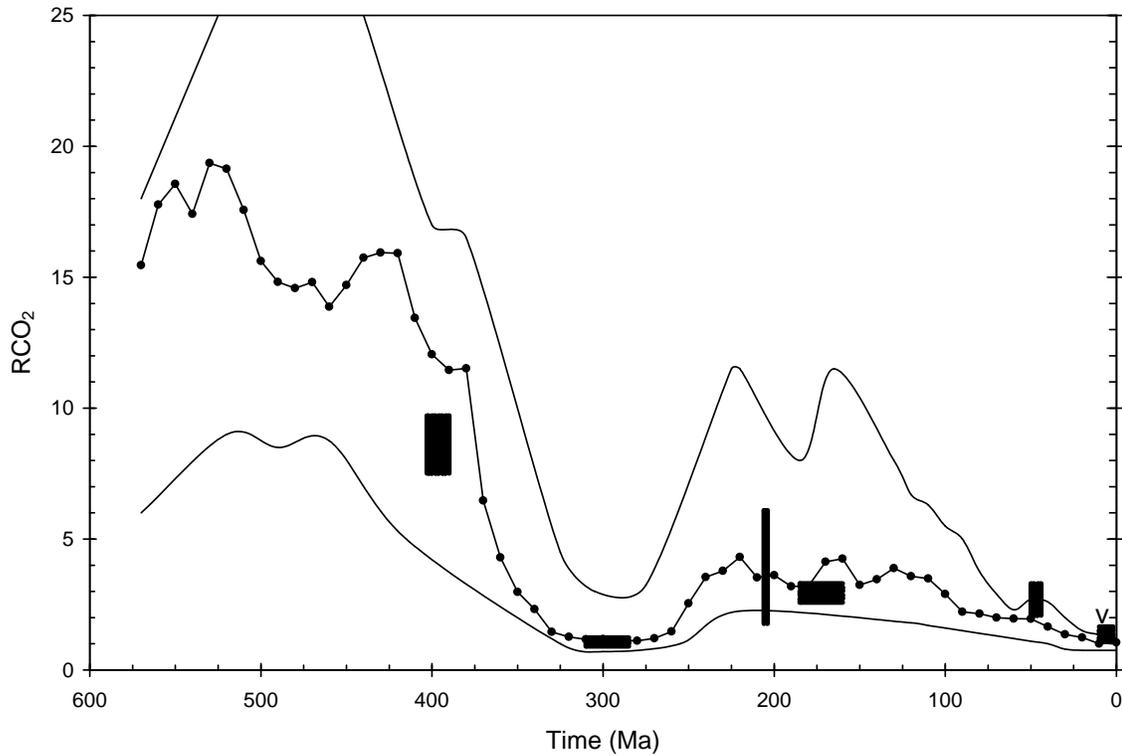


Fig. 1.6. Estimates of CO_2 for the Devonian, Carboniferous-Permian, Triassic, Jurassic, and Eocene (unmarked boxes) calculated from the stomatal ratio technique of McElwain and Chaloner (1995) superimposed over the CO_2 curve and corresponding error envelope of Berner (1994, 1998). Stomatal ratio scale calibrated to RCO_2 scale using a 1:1 correspondence; this is the 'Recent standard' of McElwain (1998). RCO_2 and stomatal ratio defined in Fig. 1.1 and text, respectively. Data from McElwain (1998) and McElwain et al. (1999). Estimates of CO_2 for the Miocene ("v") calculated from a herbaria-based training set. Data from van der Burgh et al. (1993) and Kürschner et al. (1996).

correspondence between stomatal ratios and RCO_2 (RCO_2 = ratio of mass of paleo- CO_2 to pre-industrial value; see Fig. 1.1). Using this more independent technique, all but the Devonian material agreed well with the estimates of Berner (1994). Recent changes in Berner's model, however, have pushed back the sharp drop in Paleozoic CO_2 ~40 m.y. (Berner, 1998), resulting in closer agreement between the two methods for the Devonian (Fig. 1.6).

There is growing interest in quantifying Tertiary CO_2 concentrations (Cerling et al., 1997; Pagani et al., 1999a, 1999b; Pearson and Palmer, 1999), primarily fueled by the question of whether CO_2 and temperature are coupled during this interval. Estimates from stomatal indices have the potential to help resolve this question. As for pre-Cretaceous estimates, the less quantitative stomatal ratio method of McElwain and Chaloner (1995) remains the best option.

CHAPTER 2

Ecological conservatism in the “living fossil” *Ginkgo*

2.1 Introduction

Members of the Ginkgoalean clade remained a moderately important to minor associate of mid- to high latitude paleofloras from the mid-Mesozoic to the mid-Cenozoic (Tralau, 1967, 1968; Vakhrameev, 1991). However, reconstructing the ecology of the group is problematic, given the extreme taxonomic isolation (Chamberlain, 1935; Gifford and Foster, 1989) and highly relictual distribution of what might well be only semi-wild stands of *Ginkgo biloba* Linnaeus, the sole survivor of this once moderately diverse clade (Vasilevskaya, 1963; Tralau, 1968). The objective of this study was to determine whether some aspects of *Ginkgo*'s paleoecology could be recovered from: 1) an interpretation of the sedimentary context in which its fossils occur; 2) the autecology of the modern descendants of its important fossil associates; and 3) a contextual study of the fragmentary ecology of its surviving species.

According to Tralau (1968), the Order Ginkgoales consists of six families and 19 genera. Ginkgoales first appeared in the Permian and achieved maximum diversity during the Jurassic and Early Cretaceous. Its most plausible ancestral group is the pteridosperms (“seed ferns”) (Thomas and Spicer, 1987), and especially the Peltaspermales (Meyen, 1987, p. 146). However, the clade is so isolated evolutionarily that efforts to establish its closest extant sister group have remained highly controversial. Nevertheless, there is a

growing consensus, favored by molecular data, that cycads are the most plausible closest living relative (Meyen, 1984; Thomas and Spicer, 1987; Raubeson and Jansen, 1992; Chaw et al., 1993, 1997; Rothwell and Serbet, 1994; Boivin et al., 1996; Hickey, 1996; Hasebe, 1997) rather than conifers (Chamberlain, 1935; Pant, 1977; Stewart, 1983; Crane, 1985; Doyle and Donoghue, 1986, 1987).

Undoubted remains of the Genus *Ginkgo* (*G. digitata* [Brongniart] Heer) first appeared in the Early Jurassic (Tralau, 1968: but see Vasilevskaya and Kara-Murza [1963] for a questionable attribution to the Late Triassic), making it the oldest extant genus among seed plants (Arnold, 1947). At least a dozen species have been assigned to *Ginkgo*, which achieved its greatest diversity during the Early Cretaceous (Tralau, 1968). Of particular interest for this study is *G. adiantoides* (Unger) Heer, which is morphologically identical to *G. biloba* (Seward, 1919, p. 29; Shaparenko, 1935; Manum, 1966; Tralau, 1968; Mösle et al., 1998; see Fig. 2.1). These similarities have led some authors to consider *G. adiantoides* conspecific with *G. biloba* (Seward, 1919; Tralau, 1967, 1968, p. 87). *Ginkgo adiantoides* first appeared in the Early Cretaceous and was relatively common during the Late Cretaceous and Paleogene (Tralau, 1968; Vakhrameev, 1991).

Although several additional Northern Hemisphere species of Cenozoic *Ginkgo* have been formally established, all but one of these are morphologically identical, or nearly so, to *G. adiantoides* (and *G. biloba*). One example is *G. beckii* Scott, Barghoorn, & Prakash, a species of Miocene wood associated with *G. adiantoides* foliage, which shows a striking similarity to *G. biloba* wood, with the possible exception of fewer pits per unit length on the radial walls of its tracheids (Scott et al., 1962; Mastrogiuseppe et al., 1970). The only Cenozoic form that possibly merits recognition as a separate species

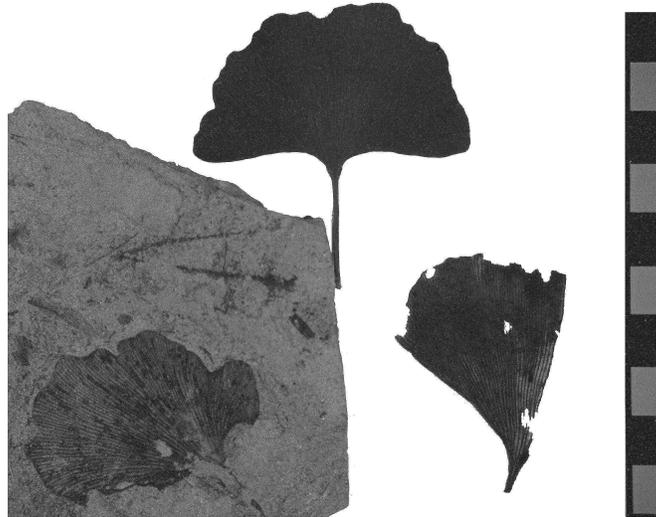


Fig. 2.1. Comparison of the leaves of modern *Ginkgo biloba* (top), middle Miocene *G. adiantoides* (right; only half of leaf present; from site 51), and middle Paleocene *G. adiantoides* (left; from site 18; University of Alberta specimen S51314). Scale bar = 1 cm.

is *G. gardneri* Florin (Tralau, 1968), which has more prominent papillae and less sinuous adaxial epidermal cells than does *G. adiantoides* (Manum, 1966). *G. gardneri* is found only in the late Paleocene deposits on the Isle of Mull, Scotland (e.g., Boulter and Kvaček, 1989). Given this lack of morphological diversity, it is possible that Ginkgoales has been monospecific (or nearly so) in the Northern Hemisphere for the entire Cenozoic. In the Southern Hemisphere a different, more strongly digitate type of *Ginkgo* leaf persists into the Eocene, but we lack data on its occurrence and do not discuss it further here.

A number of Mesozoic species of *Ginkgo* closely match the extant species as well. For example, Mösle et al. (1998) found strong similarities between the cuticles of the Early Cretaceous *G. coriacea* Florin and *G. biloba*. Villar de Seoane (1997) reported similar results for the cuticle of Early Cretaceous *G. tigrensis* Archangelsky from Argentina. Zhou (1993) compared the megaspore membranes of middle Jurassic *G. yimaensis* Zhou & Zhang with *G. biloba*, and noted few morphological differences. Van Konijnenburg-van Cittert (1971) concluded that the pollen of middle Jurassic *G. huttoni* (Sternberg) Heer was morphologically identical to *G. biloba*. Thus, *Ginkgo* is a highly conservative genus morphologically, with the long-ranging fossil species *G. adiantoides* indistinguishable from modern *G. biloba*.

2.1.1 Ecology of *Ginkgo biloba*

Ginkgo biloba (Fig. 2.2) has been cultivated for more than 2000 years in China and for some 1000 years in Japan as a source of food, shade, and beauty (Li, 1956; He et al., 1997); small stands are sometimes present within the forests customarily preserved adjacent to Buddhist and Taoist temples (Li, 1956). However, the natural ecology of *G.*

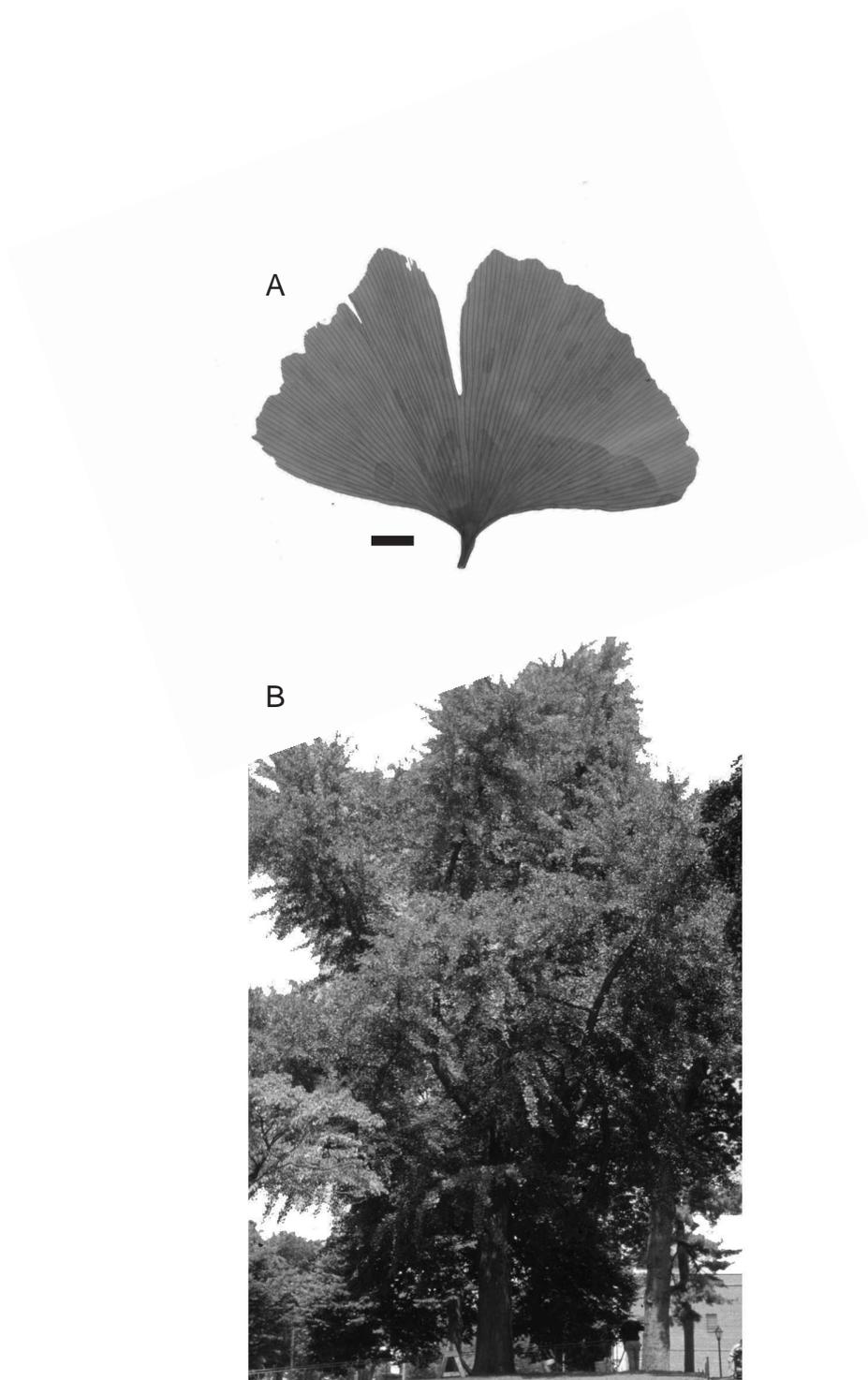


Fig. 2.2. (A) Representative leaf of *Ginkgo biloba* (scale bar = 1 cm) and (B) Mature *Ginkgo biloba* tree growing on Yale University grounds (New Haven, CT).

biloba is largely unknown because no unequivocally natural stands remain today (see discussions in Li, 1956; Franklin, 1959; Del Tredici et al., 1992; He et al., 1997). The last remaining natural populations are (or were) largely in the low coastal and interior mountains straddling the Yangtze River. This is the region where *G. biloba* was first cultivated and also harbors relictual stands of other taxa with long fossil records such as *Cryptomeria*, *Liriodendron*, *Metasequoia*, *Nothotaxus* and *Pseudolarix*. The area lies at about 30 °N and has a warm temperate mesic climate (mean annual temperature = 9-18 °C; mean annual precipitation = 600-1500 mm) (Li, 1956; Del Tredici et al., 1992; He et al., 1997). In cultivation *Ginkgo* tolerates a wide variety of seasonal climates, ranging from Mediterranean to cold temperate, where winter temperature minimums can reach –20 °C. Interestingly, nearly all fossil occurrences of *Ginkgo* lie poleward of 40° and, concomitant with cooling and increased seasonality during the Tertiary, its geographical range progressively constricted towards 40 °N, and it disappeared all together from the southern hemisphere (Tralau, 1967). Thus, both its broad modern range of climatic tolerance and the paleobotanical data suggest that the climatic parameters of its current (semi-)natural range at 30 °N are anomalous in terms of the long-term history of *Ginkgo*.

Ginkgo is a tree of medium height, reaching approximately 30 m (Del Tredici et al., 1992; He et al., 1997), and can live as long as 3500 years (Zhang et al., 1992). Although *Ginkgo* is frequently described as slow growing, under favorable conditions with warm summers it exhibits growth rates of up to 30 cm/yr for the first 30 years or so of its life (Del Tredici, 1989). During this “bolting” phase of their growth, seedlings and saplings have a straight main axis, with sparse, excurrent branches. After that, growth slows markedly—100-year-old trees may have attained only two-thirds of their mature height (Santamour et al., 1983; He et al., 1997)—and the young tree begins to fill out its

crown (Del Tredici, 1989). Plants do not produce viable seeds for 20 to 30 years (Wyman, 1965; Santamour et al., 1983; He et al., 1997), but individual trees can remain fertile for more than 1000 years (S.-A. He, personal communication, 2000). *Ginkgo*'s fusiform seeds are large, ranging from 1-2 cm in length.

In addition to sexual reproduction, *Ginkgo* can reproduce clonally from embedded buds, called chichi, that occur near the base of the trunk. If the soil in the immediate environment is disturbed, as by excessive erosion, positive geotropic growth is stimulated within its chichi. Once the apex of a chichi penetrates below ground, it forms rhizomatous tissue that can generate both aerial shoots and adventitious roots (Del Tredici, 1992). In a modern 'semi-wild' stand in the coastal mountains near the Yangtze River studied by Del Tredici et al. (1992), 40% of the larger trees had at least two primary stems but few saplings were present, a clear indication of the reproductive importance of chichi. In older individuals, these buds also form within secondary stems and are called aerial chichi. As with basal chichi, aerial chichi develop in response to a disturbance (e.g., severe crown damage) and can lead to successful clonal reproduction (Fujii, 1895; Del Tredici et al., 1992).

Modern *Ginkgo* is shade intolerant, growing best on exposed sites (He et al., 1997). Del Tredici et al. (1992) observed that seeds of *G. biloba* in a 'semi-wild' stand require an open canopy for growth and development; Jiang et al. (1990) report a similar requirement in another 'semi-wild' stand. Many of the individual trees studied by Del Tredici et al. (1992, p. 202) were growing in disturbed microsites such as "stream banks, steep rocky slopes, and the edges of exposed cliffs." However, Del Tredici (1989) also suggests that *Ginkgo* may play a role as a gap opportunist and persist in the shady understory of the forest until a gap occurs that allows it to shoot up into the canopy.

At least in cultivation, *Ginkgo* grows best in “sandy loam soils...along dams...or at the foot of hill slopes” (He et al., 1997, p. 377), environments that are well-watered and well-drained. *G. biloba* is very successful as an urban street tree (e.g., Handa et al., 1997), owing in part to its tolerance to air pollution (Kim and Lee, 1990, Kim et al. 1997), high resistance to insects (Major, 1967; Kwon et al., 1996; Ahn et al., 1997; Honda, 1997), fungi (Major et al., 1960; Major, 1967; Christensen and Sproston, 1972), bacteria (Mazzanti et al., 2000) and viruses (Major, 1967), as well as its preference for exposed sites. *Ginkgo*'s resistance to disease is due, in part, to its high production of secondary defense compounds (e.g., Yoshitama, 1997).

2.1.2 Implications for fossil *Ginkgo*

While the close morphological similarity of *Ginkgo adiantoides* to *G. biloba* makes it tempting to infer the paleoecology of the fossil species from what is known of the living, the extreme relictual nature of the modern form suggests that such surmises should be made cautiously and only with corroboration from the fossil record (cf. Hickey, 1977). Uemura (1997) thought that *Ginkgo* was ecologically conservative through time, and required moist environments. Kovar-Eder et al. (1994) studied Neogene occurrences in central Europe, and considered *G. adiantoides* an accessory element in riparian communities. Spicer and Herman (2001) reported similar depositional associations for mid-Cretaceous *G. adiantoides* in northern Alaska. In this chapter, I will provide a quantitative description of the sedimentological contexts and floral associates of fossil *Ginkgo*, concentrating primarily on *G. adiantoides*, the putative conspecific of *Ginkgo biloba*, and ranging in age from latest Cretaceous to middle Miocene. Through the

interpretation of *Ginkgo*'s sedimentological contexts and the modern ecology of the nearest living relatives of its floral associates, two independent lines of evidence will be used to interpret the paleoecology of the genus.

2.2 Methods

2.2.1 Sources of data

Sedimentological and floral data were obtained through field observations and the literature. Most data are from the Fort Union and Willwood Formations (Bighorn Basin) and the Hell Creek Formation (Williston Basin). The geographic extent of our sampling is shown in Fig. 2.3. Data sources are as follows: Williston Basin—Johnson (in press) for sedimentology and floral data, and Hicks et al. (in press) for dating, except for the Almont site, for which all data are taken from Crane et al. (1990); Bighorn Basin—Hickey (1980), Wing et al. (1995) and field observations for sedimentology and floral data, and Gingerich (2000) and Age Model 2 of Wing et al. (2000) for dating; Denver Basin (site DMNH 2360)—R. S. Barclay (unpublished data) for sedimentology, and Reynolds et al. (2001, p. 25) for dating; Basilika site—Manum (1963) for sedimentology and floral data, and Kvaček et al. (1994) for dating; south-central Alberta—Speirs (1982) and Hoffman and Stockey (1999) for sedimentology and floral data, and Fox (1990) for dating; Ardtun Head site—Boulter and Kvaček (1989) for sedimentology and floral data, and Chapter 2 for dating; Stenkul Fiord site—field observations for sedimentology and floral data; Kalkreuth et al. (1996) for dating; Beaver Creek site—K. R. Johnson (unpublished data)

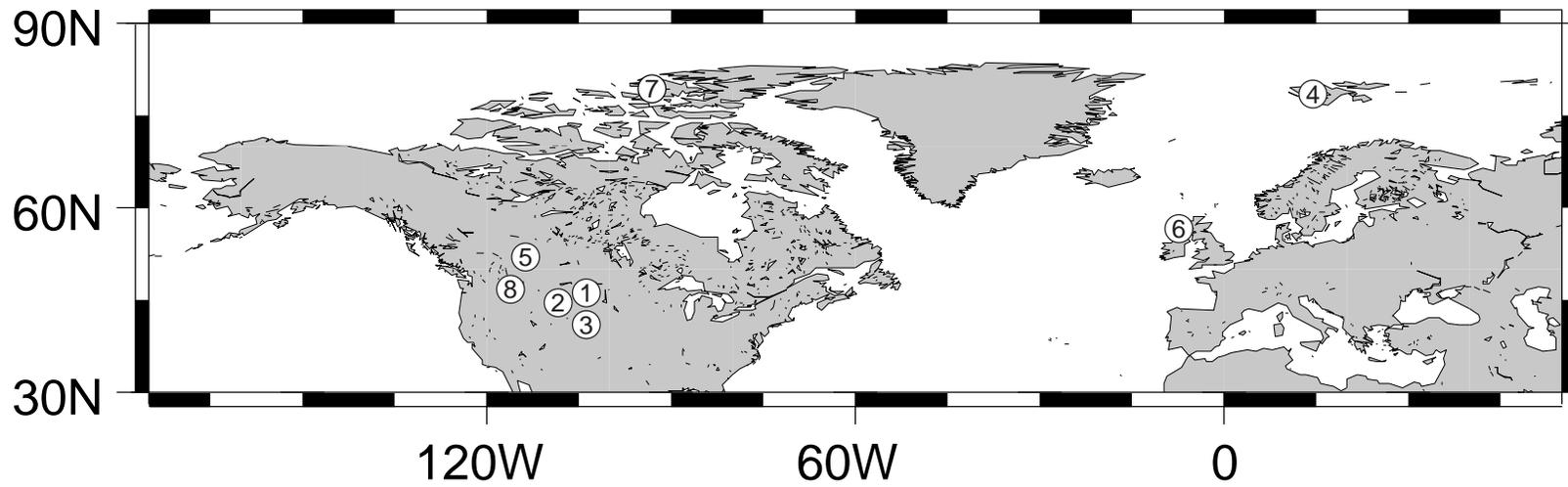


Fig. 2.3. Map showing sampling areas. Numbers refer to the following field areas: 1. Williston Basin; 2. Bighorn Basin; 3. Denver Basin; 4. Spitsbergen; 5. south-central Alberta; 6. Isle of Mull; 7. Axel Heiberg Island; 8. north-central Idaho.

for sedimentology, and C. N. Miller (unpublished data) for dating; Juliaetta site—field observations for sedimentology and floral data, and Reidel and Fecht (1986) for dating.

2.2.2 Recognition of sedimentological contexts

Sites were assigned to environments of deposition based upon the lithology, grain size, primary stratification, overall cross-sectional and plan-view shape of their deposits, along with their organic content and nature of their contacts with surrounding sediments, using criteria developed by J. R. L. Allen (1964, 1965), Hickey (1980), Wing (1984) and Miall (1992). We will use the following terms for the environments of deposition that we recognize here: relief / abandoned channel, crevasse splay, active trunk channel, backswamp, and distal floodplain.

Relief channels are represented by thin, shallow-lenticular, laterally restricted bodies of silt to sand-size sediment that fill minor stream axes used in time of flood. Abandoned channels represent a segment of a stream or river that was intermittently or permanently cut off from its main channel, whose original depth can be reconstructed from the thickness of the total sedimentary package. They often have clay- or mudstone in their axes. The basal contacts of both types of channels are frequently down-cut, concave upward, and veneered with a coarse (often pebble-sized) basal lag deposit. Plant remains within these deposits are typically parautochthonous (Wing, 1984; Gastaldo et al., 1989). These channel deposits are characterized by relatively thin (<5 cm for relief channels and <1 m for abandoned channels) couplets of coarser sandstone and siltstone with finer siltstone and mudstone that represent the alternating periods of flooding or channel reactivation and slackwater or subareal phases (Wing, 1984). Intervals within abandoned

channels often show evidence of standing water, such as aquatic plants and invertebrates, and successions of small-scale, fining-upwards sequences (Wing, 1984).

Crevasse splay deposits are floodplain deposits formed by the breaching of a levee, typically during flood events. They are characterized by sand- or silt-size sediments with generally tabular beds that are sometimes cross-bedded or cross-laminated. These often overlie finer-grained, massive floodplain-sediments. Proximal to the break-through point, downcut contacts with concave upward bedding and scour and fill structures are more common, but distal to the break-through point basal contacts tend to be more parallel with one another. Beds of sandstone in splays represent multiple reactivation events and range in thickness from centimeters to more than a meter. Often these grade upward into progressively finer sediments that represent low energy floodplain conditions. The modal grain size of the sediments in these reactivation events usually coarsens upwards due to the progressive lateral shift of the stream axis towards a given point in the crevasse splay (Miall, 1992). Crevasse splay deposits are generally more laterally extensive than relief channels, but are not traceable for long ($>10^2$ m) distances. A detailed microstratigraphic section at a site that we interpret as a crevasse splay is illustrated in Fig. 2.4.

Other sedimentological contexts distinguished here include backswamps, which are dominated by carbonaceous shale; active trunk channels, which are characterized by massive cross-bedded sandstone; and ponds / lakes, which are distinguished by laminated mudstone and the presence of aquatic plants and animals. Because of the convergence of their properties, we assigned large abandoned channels to the pond / lake category.

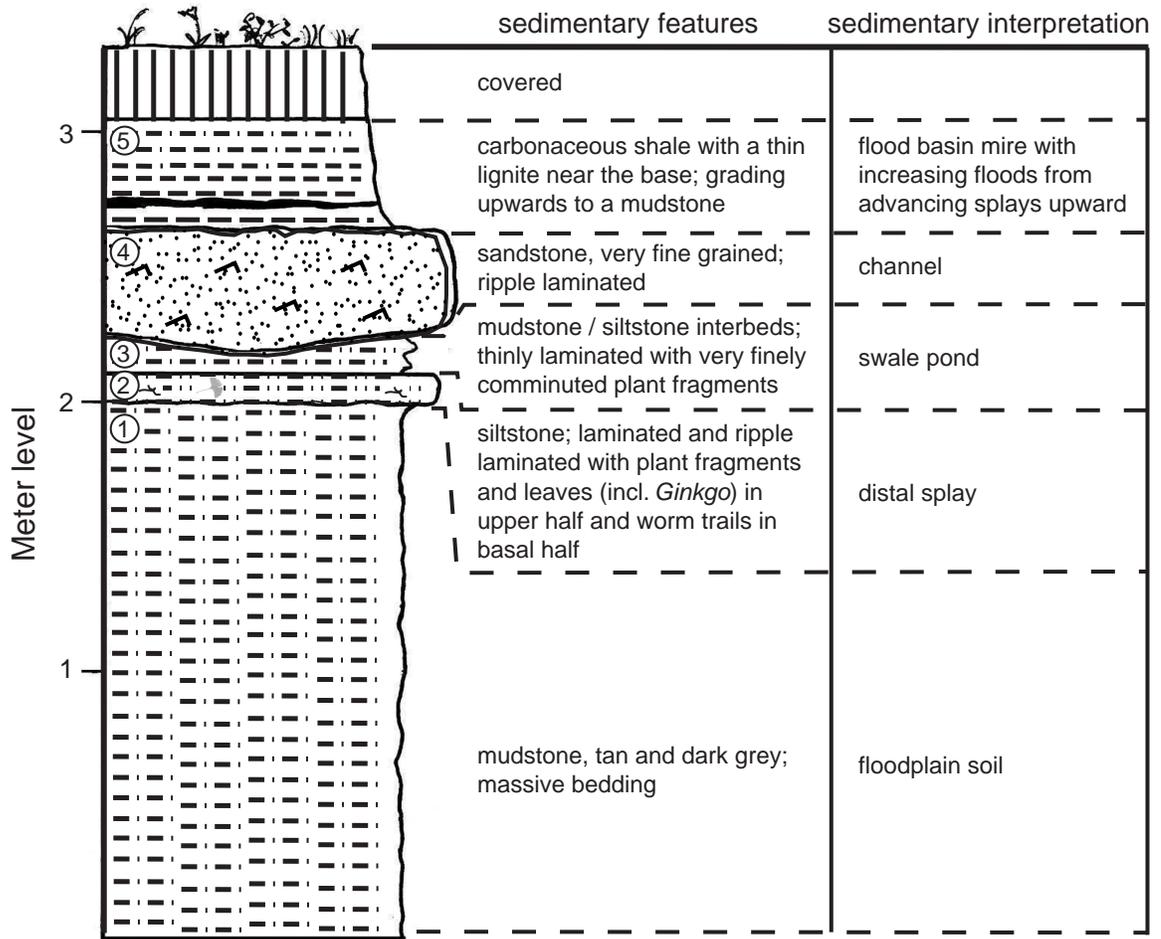


Fig. 2.4. Microstratigraphic section for Chance locality (site 23). Section is near Chance, MT (45° 00' 29.2" N, 109° 04' 53.0" W).

2.2.3 Floral associates

We generated floral lists for most sites. When possible, we limited these lists to the individual beds within a deposit where *Ginkgo* was found. In general, we follow the taxonomic terminology of Wing et al. (1995). In two cases, we combined closely related species of the family Platanaceae into single counting units. The first of these units consists of the broadly trilobed *Platanus raynoldsii* Newberry and *P. guillelmae* Goeppert, and the second of the narrowly five-lobed *Macginitiea gracilis* (Lesquereux) Wolfe & Wehr and *M. brownii* (Berry) Wolfe & Wehr.

All but two of our sites contain *Ginkgo adiantoides*. *G. spitsbergensis*, found at one site, is considered conspecific with *G. adiantoides* by Tralau (1968), while *G. gardneri*, found at one site on the Isle of Mull, may be a distinct species, as discussed above.

2.3 Results

2.3.1 Sedimentological contexts

Based upon our sampling of 48 sites spanning the latest Cretaceous to middle Miocene, *Ginkgo* is most often found in relief / abandoned channels (44% of all sites) and crevasse splays (38%) (Table 2.1; raw data presented in Appendix 2.1). In most relief / abandoned channel deposits, *Ginkgo* occurs within the coarser-grained intervals. *Ginkgo* is rarely preserved in backswamps (4%). Certain depositional environments are preserved

Table 2.1. Summary of the distribution of sedimentological contexts and floral associates. Description of columns are as follows: 1) % occurrence at all *Ginkgo*-bearing sites; 2) % occurrence at *Ginkgo*-bearing sites that overlap with the datasets of Johnson (in review) and Wing et al. (1995); 3) % occurrence at all sites (including *Ginkgo*-bearing sites) from the datasets of Johnson (in review) and Wing et al. (1995); 4) % difference between the two restricted sets (i.e., column [2]-[3]); 5) Level of significance between the two restricted sets (G-test of independence; Sokol and Rohlf, 1995, p. 729). Since the sedimentological contexts are not independent of one another (i.e., columns add up to 100%), these *P*-values were adjusted using the Dunn-Šidák method in the following manner: $a' = 1 - (1 - a)^{1/k}$, where $k = 6 =$ the number of significance tests (Sokol and Rohlf, 1995, p. 239).

	<i>Ginkgo</i> sites (%)		All sites from restricted set (weighted %)	Difference in restricted set (%)	<i>P</i>
	all	restricted set			
sedimentological contexts	(<i>n</i> = 48)	(<i>n</i> = 41)	(<i>n</i> = 288)		
relief channel / abandoned channel	43.8	41.5	14.3	27.2	0.004
crevasse splay	37.5	41.5	17.8	23.7	0.012
channel	8.3	9.8	5.5	4.3	0.98
backswamp	4.2	4.9	53.2	-48.4	<0.001
distal floodplain	4.2	4.9	0.8	4.0	0.60
other (mostly stable ponds / lakes)	4.2	0.0	8.4	-8.4	0.08
floral associates	(<i>n</i> = 47)	(<i>n</i> = 41)	(<i>n</i> = 289)		
(<i>Ginkgo adiantoides</i>)			8.4		
<i>Cercidiphyllum genatrix</i>	59.6	63.4	32.9	30.5	<0.001
<i>Metasequoia occidentalis</i>	44.7	46.3	27.7	18.7	0.022
<i>Platanus raynoldsii / guillelmae</i>	42.6	48.8	23.3	25.5	0.002
<i>Glyptostrobus europaeus</i>	29.8	31.7	44.3	-12.6	0.19

more commonly than others or are more likely to preserve fossil plants. The high proportion of relief / abandoned channels and crevasse splays might therefore simply reflect the numerical abundance of these plant fossil-containing deposits. To normalize for this potential bias, we determined the distribution of sedimentological contexts for all fossil plant sites (including *Ginkgo*-bearing sites) documented by Johnson (in press) ($n = 157$ sites) and Wing et al. (1995) ($n = 131$). Together, these datasets span the interval from the latest Cretaceous to the early Eocene and include the late Cretaceous Hell Creek Formation and earliest Paleocene part of the Fort Union Formation in the Williston Basin and the Paleocene through early Eocene Fort Union and Willwood Formations in the Bighorn Basin. We considered only our *Ginkgo*-bearing sites included in these datasets ($n = 41$ sites), and weighted the full datasets of Johnson (in press) and Wing et al. (1995) to reflect the distribution of these *Ginkgo*-bearing sites (22% from the Williston Basin, 78% from the Bighorn Basin).

For these two geographic regions, *Ginkgo* is preferentially found in both relief / abandoned channel (42% for *Ginkgo* localities vs. 14% for all plant localities) and crevasse splay (42% vs. 18%) deposits (Table 2.1; Fig. 2.5). Equally striking is that the likelihood of finding *Ginkgo* in a backswamp is far less than the background percentage of backswamp deposits (5% vs. 53%). The differences for all three of these sedimentological contexts are significant at the $\alpha = 0.02$ level (G-test of independence).

2.3.2 *Floral associates of Ginkgo*

Based upon a sampling of 47 *Ginkgo*-bearing sites spanning the latest Cretaceous to middle Miocene, *Ginkgo* most commonly is fossilized with *Cercidiphyllum genatrix*

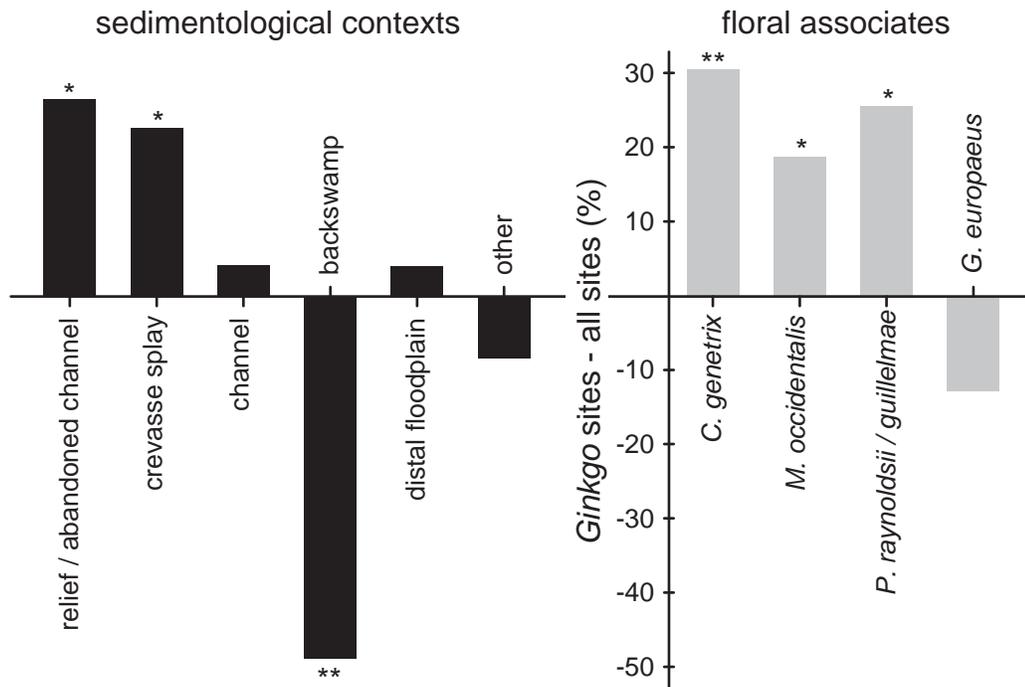


Fig. 2.5. Deviations of the proportion of sedimentological contexts and floral associates at *Ginkgo*-bearing sites from the background occurrence rate of these sedimentological contexts and floral associates. Data taken from second-to-last column in Table 2.1.

Methodology of calculation discussed in text. Asterisks represent whether the differences in proportionality are significant at the $\alpha = 0.05$ level (*) or $\alpha = 0.001$ level (**) (G-test of independence).

(Newberry) Hickey (co-occurs at 60% of all *Ginkgo* sites), *Metasequoia occidentalis* (Newberry) Chaney (45%), *Platanus raynoldsii / guillelmae* (43%), and *Glyptostrobus europaeus* (Brongniart) Heer (30%) (Table 2.1; raw data presented in Appendix 2.2). All four of these associates have temporal ranges that span the latest Cretaceous to middle Miocene. Analogous to the potential sedimentological taphonomic bias, it is possible that these floral associates simply dominate the megafloral record during this interval, which, if true, would diminish the paleoecological significance of our results. As with the data on sedimentary environments, then, we compared a weighted average of the overall floral distribution patterns for the Hell Creek and Fort Union Formations in the Williston Basin ($n = 158$ sites) (Johnson, in press) and the Fort Union and Willwood Formations in the Bighorn Basin ($n = 131$) (Wing et al., 1995) with the *Ginkgo*-bearing sites that overlap with these studies ($n = 41$). *C. genatrix* and *P. raynoldsii / guillelmae* occur more often with *Ginkgo* relative to their overall occurrence rate (63% vs. 33% and 49% vs. 23%, respectively) (Table 2.1; Fig. 2.5). These differences are significant at the $\alpha = 0.003$ level (G-test of independence). *Ginkgo* also preferentially associates with *M. occidentalis* (46% vs. 28%; $P = 0.02$; G-test of independence). The background occurrence rate of *G. europaeus*, however, is higher than its co-occurrence rate with *Ginkgo* (44% vs. 32%) (Table 2.1; Fig. 2.5), suggesting that it is not useful in determining the paleoecology of *Ginkgo*.

2.3.3 Microstratigraphy: a detailed example

The Chance locality (site 23) in the Tiffanian portion of the Fort Union Formation of the Bighorn Basin displays sedimentological and floral characteristics common to

many *Ginkgo*-bearing sites examined in this study. Here, fossil *Ginkgo* leaves are found in the upper half of a 10 cm-thick unit of interbedded mudstone and siltstone (unit 2 in Fig. 2.4) that is inferred to represent the distal portion of a crevasse splay. In addition to *G. adiantoides*, leaves of *Zizyphoides flabella* Crane, Manchester & Dilcher, *Platanus raynoldsii*, *Acer silberlingii* Brown and *Wardiaphyllum daturaefolia* (Ward) Hickey also occur. The fragmentary nature of many of these leaves and their occurrence in the laminated siltstone of a splay indicate that this flora has been subjected to some transport, in this case inferred to have been from the nearby slopes of the stream levee. The fossil bearing unit overlies a massive, flood-basin mudstone (unit 1) that shows very weakly developed soils features (inceptisol). Following the deposition of the *Ginkgo*-bearing splay siltstone, deposition of a thinly laminated siltstone-mudstone unit indicates that the area became a swale-pond (unit 3). This in turn was replaced by a sandstone lens that is inferred to represent a trunk channel. This is overlain by sediments representing a distal floodplain (units 4 and 5).

2.4 Discussion

2.4.1 Paleoecology of Ginkgo

The sedimentological features at the majority of *Ginkgo* localities indicate that it grew primarily in disturbed environments along stream margins and the distal sides of levees. These environments are typically well-watered (due to a shallow water table and frequent flood events) yet well-drained (due to their primarily coarse-grained substrate).

Due to their high disturbance rate, these environments typically host open-canopy forests. Although *Ginkgo* leaves tend to sort with silt-sand size particles in flume experiments (Spicer and Greer, 1986), this would not explain their distribution in both coarse-grained, typically cross-stratified sediments (representing crevasse splays, relief channels, and rejuvenated abandoned channels) and fine-grained, parallel-laminated sediments (e.g., abandoned channels).

As reported above, *Cercidiphyllum genatrix* and a form of *Platanus* delimited by the pair *P. raynoldsii* and *P. guillelmae* are two of the more common floral associates of *Ginkgo*. *C. japonicum* Siebold & Zuccarini, the nearest living relative (NLR) of *C. genatrix*, grows in the wild today in Japan and China. It usually occurs in small gaps or along stream margins (Seiwa and Kikuzawa, 1996). It is a fast growing pioneer species, often reproduces by sprouting, and prefers mesic, disturbed sites (Ishizuka and Sugawara, 1989). Interestingly, at mid-elevations (1450-3500 m) in the mountains of south-central China, *C. japonicum* occurs on scree slopes (Tang and Ohsawa, 1997) and in other highly disturbed habitats (Wang, 1961). Although plant remains are rarely preserved in alluvial fan facies, at Ardtun Head (Isle of Mull, Scotland) *Ginkgo* is found in fine-grained sediments within an alluvial fan deposit (see Appendix 2.1). As noted above, modern *Ginkgo* has been observed growing on steep rocky slopes and along edges of exposed cliffs (Del Tredici et al., 1992). It is possible, then, that disturbance is the overriding ecological filter for *Ginkgo*, and the fluvial deposits where *Ginkgo* is most commonly preserved represent only a small fraction of its potential habitat range.

The NLR's of *P. raynoldsii* and *P. guillelmae* are *P. occidentalis* Linnaeus, *P. orientalis* Linnaeus, and their hybrid *P. × acerifolia* Willdenow. These living species are most commonly found in riparian habitats (Tang and Kozlowski, 1982; Ware et al., 1992;

Thomas and Anderson, 1993; Atzmon and Henkin, 1998; Everson and Boucher, 1998); for example, *P. occidentalis* often grows on the lowest stream terrace (McClain et al., 1993) where flood frequency is highest (Bell, 1980). *P. occidentalis* can develop adventitious roots and lenticels in response to flooding (Tsukahara and Kozłowski, 1985), however it appears to require aerated soils during the growing season (Everson and Boucher, 1998).

Metasequoia occidentalis, another common *Ginkgo* associate (Table 2.1), has remained largely unchanged morphologically throughout the Cenozoic (Chaney, 1951; Christophel, 1976; Liu et al., 1999) and is represented today by *M. glyptostrobooides* H. H. Hu & Cheng. Like *Ginkgo*, the geographic range of *Metasequoia* has been severely restricted, however areas of near-natural stands still grow in one mountain valley in south-central China. It occurs there along stream banks and at seepages at the bases of slopes (Chu and Cooper, 1950). These modern observations contrast in part with the paleobotanical observations that show *Metasequoia* most often occurring in swamp, swamp margin and distal splay environments (e.g., Hickey, 1980; Momohara, 1994; Wing et al., 1995; Falder et al., 1999). This discrepancy may result either from the fact that most of the valley floor where *Metasequoia* makes its last stand has been converted to agriculture and habitat space has been lost, or from a shift in the ecological niche of *Metasequoia* during the Cenozoic.

In summary, the sedimentological record strongly indicates that *Ginkgo* most commonly grew along streamsides and on the proximal slope of levees. The ecology of the NLR's of two genera of *Ginkgo*'s common floral associates (*Cercidiphyllum* and *Platanus*) support these sedimentological interpretations. *Ginkgo*'s association with *Metasequoia* proves problematic, however, as the latter most commonly grew in swamps

and distal levees during the Tertiary. It is possible then, that either *Ginkgo* also grew in more stable distal floodplain settings or that *Metasequoia* was a minor associate in disturbed levee and riparian-type environments. This latter option is supported, albeit weakly, by modern ecological observations of *Metasequoia*.

In general, NLR-derived paleoecological interpretations are less reliable than sediment-derived interpretations because a given lineage of plants can shift its ecological tolerances over geologic time (e.g., Hickey, 1977; Wolfe, 1977), and because few late Cretaceous and early Tertiary plants have close modern relatives. In contrast, the sedimentology of an autochthonous fossil plant site retains a primary paleoecological signal. Only recently have paleobotanists begun to analyze the sedimentological contexts of their assemblages in an effort to extract paleoecological information (e.g., Hickey, 1980; Spicer, 1980; DiMichele and Wing, 1988; Burnham, 1994; Wing et al., 1995; Spicer and Herman, 2001). The results of this study highlight the potential of applying sedimentological data from a large number of fossil plant sites, and we hope that the application of such data becomes more common in the future.

2.4.2 Implications for the relationship between G. biloba and G. adiantoides

Analysis of our data indicate that there were no striking changes in the sedimentological context of *Ginkgo* during the latest Cretaceous to early Eocene (Fig. 2.6A). *Ginkgo* consistently associated with unstable crevasse splay and relief / abandoned channel environments. Although there is a shift to more relief / abandoned channel localities in the early Eocene, this is driven by a drying trend in the Bighorn Basin that

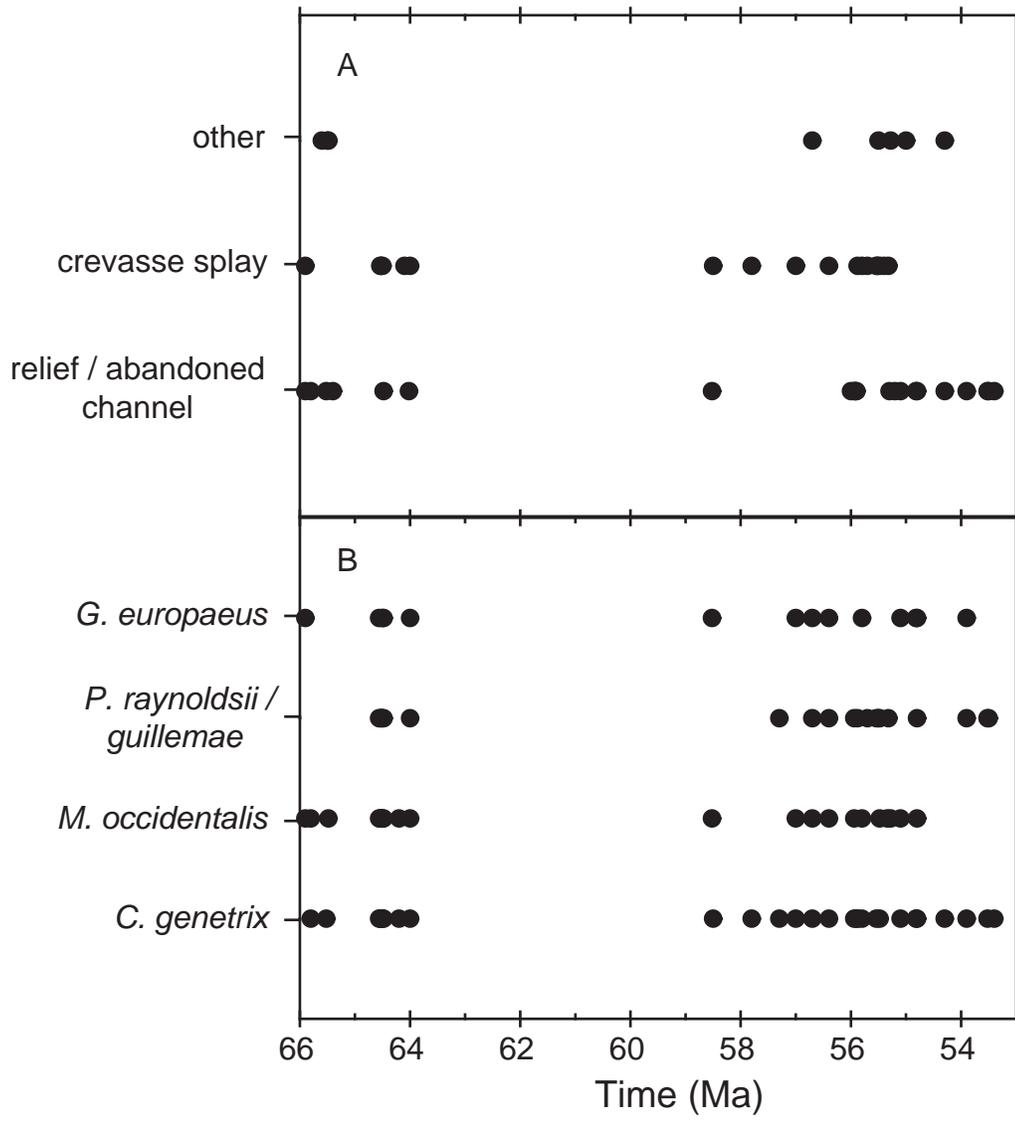


Fig. 2.6. Temporal distribution of (A) sedimentological contexts and (B) principle floral associates for latest Cretaceous-early Eocene *Ginkgo*-bearing sites.

restricted the preservation of fossil plants to these environments and to tabular carbonaceous layers representing swampy floodbasins (Wing, 1984).

Ginkgo's floral associates also remain stable through the early Paleogene (Fig. 2.6B). This stability is striking considering the large number of factors that influence plant migrations and evolution. The one minor change in this time series is the lack of *Metasequoia* during the early Eocene, although this may be due to the paucity of early Eocene backswamp deposits relative to the late Paleocene in the Bighorn Basin (Wing et al., 1995). If correct, this observation provides additional support for inferring *Ginkgo*'s preference for streamside and levee habitats.

Data are generally lacking for the Neogene. However, at the middle Miocene Juliaetta site in Idaho, *Ginkgo* occurs in sandy delta foreset beds (Appendix 2.1). Although the overall depositional setting is lacustrine, *Ginkgo* is only found in this high-energy delta environment, and so perhaps grew in a nearby riparian setting and was carried by the river to the lakeshore. *Amentotaxus gladifolia* (Ludwig), Ferguson, Jähnichen, & Alvin is the single floral associate of *Ginkgo* at the Juliaetta site. *A. gladifolia* is also present in the late Paleocene Ardtun Head site, some forty million years earlier. Its nearest living relative, *A. formosana* Li, grows along streams on mountain flanks (Page, 1990).

On the basis of our sedimentological and floral data, *Ginkgo* appears to have been ecologically conservative throughout the Cenozoic. The environmental preferences of the modern *G. biloba* match the paleoecological interpretations generated here. This marked stability in environmental preferences supports previous work, based upon morphology alone, that *G. adiantoides* and *G. biloba* are conspecific.

2.4.3 *The ecological paradox of Ginkgo and a possible mechanism for its decline*

Woody plants successful in highly disturbed habitats today usually share a suite of traits described as the competitive-ruderal strategy, sensu Grime (2001). These may include an early successional habit, shade intolerance, high growth rates, ability for clonal reproduction (Everitt, 1968; Eriksson, 1993), early reproductive maturity, small seed size (Harper et al., 1970; Westoby et al., 1992), the production of few secondary defense compounds (Coley et al., 1985), short life span, and a rapid rate of evolution (Eriksson and Bremer, 1992). *Ginkgo* shares some of these characteristics, namely shade intolerance, rapid early growth, and a clonal habit, but many of its life history traits counter those considered beneficial in disturbed environments. *Ginkgo* requires > 20 years to reach reproductive maturity, produces large seeds needing a protracted period for fertilization and embryo development, manufactures an impressive array of secondary defense compounds, can live for > 3000 years, and has an extremely slow rate of evolution.

Ginkgo therefore represents an ecological paradox: it appears to favor disturbed habitats, and has likely done so for more than 65 million years, yet the living species has few of the life history traits typical of plants that prosper in disturbed settings. One solution to this paradox is to accept a non-uniformitarian view that plants in the geologic past operated in a different ecological regime. Angiosperms dominate the canopy today in most riparian and crevasse splay-type environments, and are the principle taxon from which modern ecological concepts for disturbed habitats are derived. The Ginkgoalean clade first appeared in the Permian and the Genus *Ginkgo* in the early Jurassic, making it possible that, prior to the radiation of angiosperms early in the Early Cretaceous (Doyle

and Hickey, 1976), certain competitive-ruderal ecological strategies were less highly developed than in the present.

It is difficult to test this hypothesis directly because detailed sedimentological data for pre-angiospermous Mesozoic plant fossil-bearing sites are rare, and many lineages that may have dominated these disturbed habitats are extinct. However, from at least the Jurassic, the Ginkgoales show a strong tendency to be associated with sandy channel deposits (Hughes, 1976; Falcon-Lang et al., 2001; Spicer and Herman, 2001; L. J. Hickey, unpublished field data from the Jurassic in Yorkshire, England). In the subtropical Lower Cretaceous Wealden beds of northwest Germany, *Baiera* F. Braun is restricted to barrier sands, while *Ginkgoites* Seward is found in environments ranging from muddy floodplains to backswamps (Pelzer et al., 1992).

Prior to the flowering plants, ferns dominated the early seral stages in unstable, fluvial settings (Hughes, 1976; Hickey and Doyle, 1977; Taylor and Hickey, 1996) and probably provided the chief competition for *Ginkgo* from germination to the young sapling stage. In competition with the ferns, *Ginkgo*'s large seed reserves and "bolting" habit are inferred to have been sufficient to carry it beyond the herbaceous pteridophyte canopy. Among arborescent plants, *Ginkgo*'s chief competitors for crown space before the rise of the angiosperms were probably Bennettitales and various pteridosperms and tree ferns, plants whose sedimentary context, growth-habit, wood, and leaf structure suggest that they produced a relatively low, open canopy on disturbed sites (Hughes, 1976; Crane, 1985). The growth strategy of living *Ginkgo*, which undergoes rapid vertical elongation to a height of 10 m before elaborating lateral branches, would have been adaptive in such a situation.

Many of the early flowering plant lineages appear to have evolved in disturbed riparian habitats (Taylor and Hickey, 1996), and so could have been directly competing with *Ginkgo* for resources. For example, studies of the mid-Cretaceous (Albian-Cenomanian) of northern Alaska (Spicer and Herman, 2001) and the Antarctic Peninsula (Falcon-Lang et al., 2001) both indicate ginkgo-angiosperm-fern communities in riparian depositional environments. If the combination of rapid life cycles, high dispersal potential, and abundant foliage production in angiosperms was sufficient to out-compete *Ginkgo* in these settings, then it is possible that they played a role in the decline of *Ginkgo*.

There is also a striking temporal correlation between the rise of relative diversity in angiosperms (Ligard and Crane, 1990; Lupia et al., 1999) and the decline in Ginkgoalean diversity (Tralau, 1968) (Fig. 2.7). This comparison is not ideal as one must assume that all Ginkgoales possessed adaptive strategies similar to those of *G. adiantoides*. However, even within the genus *Ginkgo* there is a large drop in diversity between the Early and Late Cretaceous (Tralau, 1968). The relative diversity of ‘other seed plants’ (see Fig. 2.7), consisting of Ginkgoales, Bennettitales, Caytoniales, Cycadales, Czekanowskiales and Gnetales, but not conifers, also declines concomitantly with the rise of angiosperm diversity (Fig. 2.7; Ligard and Crane, 1990). The relative diversity of ferns, some of which preferred disturbed streamside environments (Taylor and Hickey, 1996), drops sharply as well in the mid-Cretaceous, while the diversity of conifers, which preferred backswamp environments, remains largely unchanged (Ligard and Crane, 1990; Lupia et al., 1999). These observations further suggest that angiosperms restructured the ecology of disturbed floodplain environments during the Cretaceous.

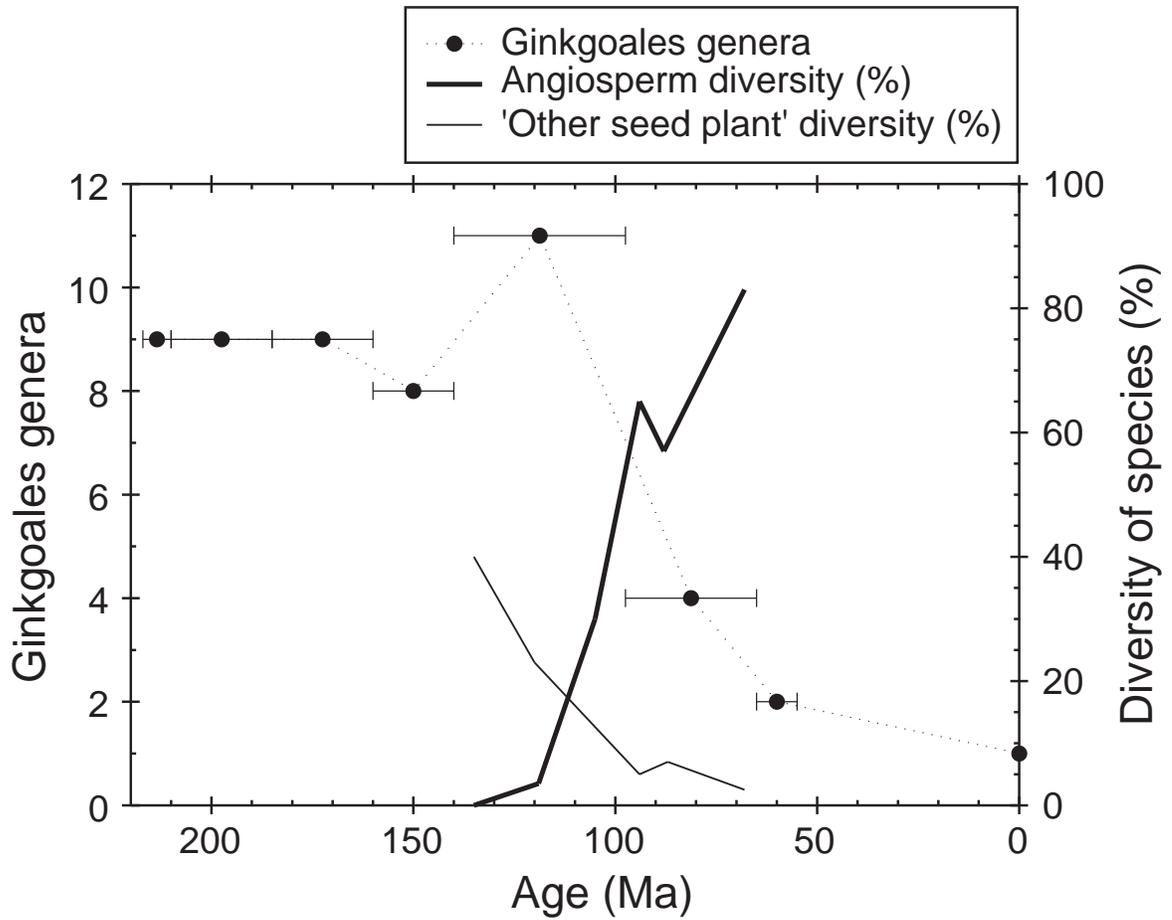


Fig. 2.7. Temporal trend of the number of Ginkgoales genera and the percent of total plant species that are angiosperms and 'other seed plants' (Ginkgoales, Bennettitales, Caytoniales, cycads, Czekanowskiales and Gnetales). Ginkgoales data from Tralau (1968); other data from Ligard and Crane (1990).

Perhaps the most surprising fact is that *Ginkgo* survives to the present at all. The persistence of the *Ginkgo* lineage may relate to its occurrence in mid- and high latitude areas. Wing and Boucher (1998) argued that lower temperatures and less light at higher latitudes make it more difficult for fast-growing competitive-ruderal trees to prosper, and that this explained the delayed increase in angiosperm diversity and dominance at higher latitudes during the Cretaceous (e.g., Lupia et al., 1999). If so, *Ginkgo* may have persisted because it was a mid- to high-latitude lineage for most of its existence; with growth limited by light and temperature, a relatively slow-growing competitive-ruderal such as *Ginkgo* could persist in the face of angiosperm competitors with the potential for faster rates of growth.

2.5 Conclusions

Ginkgo is an extreme example of a geologically long-lived genus, with its one living species arguably having a temporal range of > 100 Myr. A quantitative survey of sedimentological and floral data from 51 *Ginkgo*-bearing fossil sites, spanning the latest Cretaceous to middle Miocene, indicate that *Ginkgo* was largely confined to disturbed stream margin and levee environments. Furthermore, the stability of *Ginkgo*'s sedimentological and floral associations through the time series parallels the morphological identity of the fossil species, *G. adiantoides*, and the living *G. biloba*. As many of *Ginkgo*'s life history traits (e.g., long life-span, large seeds, and late sexual maturity) are not considered advantageous today in highly disturbed habitats, it is possible

that *Ginkgo* represents the survival of a pre-angiospermous strategy for growth in well-drained, disturbed habitats.

CHAPTER 3

Paleobotanical evidence for near present-day levels of atmospheric CO₂ during part of the Tertiary

Understanding the link between the greenhouse gas carbon dioxide (CO₂) and Earth's temperature underpins much of paleoclimatology and our predictions of future global warming. Here, we use the inverse relationship between leaf stomatal indices and the partial pressure of CO₂ in modern *Ginkgo biloba* and *Metasequoia glyptostroboides* to develop a CO₂ reconstruction based on fossil *Ginkgo* and *Metasequoia* cuticles for the middle Paleocene to early Eocene and middle Miocene. Our reconstruction indicates that CO₂ remained between 300 and 450 parts per million by volume for these intervals with the exception of a single high estimate near the Paleocene/Eocene boundary. These results suggest that factors in addition to CO₂ are required to explain these past intervals of global warmth.

Atmospheric CO₂ concentration and temperature have been tightly correlated for the past four Pleistocene glacial-interglacial cycles (Petit et al., 1999). Various paleo-CO₂ proxy data (Ekart et al., 1999; Royer et al., 2001) and long-term geochemical carbon cycle models (Tajika, 1998; Berner and Kothavala, 2001; Wallmann, 2001) also suggest that CO₂-temperature coupling has, in general, been maintained for the entire Phanerozoic (Crowley and Berner, 2001). Recent CO₂ proxy data, however, indicate low CO₂ values during the mid-Miocene thermal maximum (Pagani et al., 1999; Pearson and Palmer, 2000), and results for the middle Paleocene to early Eocene, another interval of known

global warmth relative to today, are not consistent, ranging from ~300 to 3000 parts per million by volume (ppmv) (Ekart et al., 1999; Pearson and Palmer, 2000). Here we address this problem by developing and applying an alternative CO₂ proxy based on the inverse correlation between the partial pressure of atmospheric CO₂ and leaf stomatal index (SI), with the aim of reconstructing CO₂ for both intervals to determine its role in regulating global climate.

Most modern vascular C₃ plants show an inverse relationship between the partial pressure of atmospheric CO₂ and SI (Woodward, 1987; Woodward and Bazzaz, 1988; Chapter 1), a likely response for maximizing water-use efficiency (Woodward, 1987). SI is calculated as: $SI (\%) = (SD / [SD + ED]) \times 100$, where a stoma is defined as the stomatal pore and two flanking guard cells, *SD* = stomatal density and *ED* = non-stomatal epidermal cell density. Since SI normalizes for leaf expansion, it is largely independent of plant water stress, and is primarily a function of CO₂ (Woodward, 1987; Chapter 1). This plant-atmosphere response therefore provides a reliable paleobotanical approach for estimating paleo-CO₂ levels from SI measurements on Quaternary (Rundgren and Beerling, 1999) and pre-Quaternary fossil leaves (van der Burgh et al., 1993). Because stomatal responses to CO₂ are generally species-specific (Chapter 1), one is limited in paleo-reconstructions to species that are present both in the fossil record and living today. Fossils morphologically similar to living *Ginkgo biloba* and *Metasequoia glyptostroboides* extend back to the Early and Late Cretaceous, respectively, and many workers consider the living and fossil forms conspecific (Chaney, 1951; Tralau, 1968). In this study, we use *G. adiantoides* and *M. occidentalis*, the forms most closely resembling *G. biloba* and *M. glyptostroboides*, and also *G. gardneri*, which has more prominent papillae and less sinuous upper epidermal cells than *G. biloba* (Tralau, 1968).

Measurements of SI made on fossil *Ginkgo* and *Metasequoia* were calibrated with historical collections of *G. biloba* and *M. glyptostroboides* leaves from sites that developed during the anthropogenically driven CO₂ increase of the past 145 years and with saplings of *G. biloba* and *M. glyptostroboides* grown in CO₂-controlled greenhouses¹. These data show a strong linear reduction in SI for both species between 288 and 369 ppmv CO₂ and a nonlinear response at CO₂ concentrations above 370 ppmv (Fig. 3.1). Because SI responds to partial pressure, not mole fraction (Woodward and Bazzaz, 1988), the effects of elevation must be considered. All of the leaves measured for the training set grew at elevations <250 m where concentration \cong partial pressure, so a correction is not needed. Both nonlinear regressions are highly significant (Fig. 3.1); however, a discontinuity exists for *Ginkgo* between the experimental results above 350 ppmv and the rest of the calibration set. Many species require >1 growing season for SI to adapt to high CO₂ (Chapter 1), and so these experimental results likely represent maxima for a given CO₂ level. Nevertheless, due to this discontinuity as well as the small sample

¹ Training sets were largely derived from herbarium sheets, where three fields of view (0.1795 mm² each) from each of five leaves were measured using epifluorescence (for *Ginkgo*) and transmitted light (for *Metasequoia*) microscopy. Geographic origins of herbarium sheets range from the United States, Japan and China. During the growing seasons of 1999 (*Ginkgo* and *Metasequoia*) and 2000 (*Ginkgo*), measurements were made on living trees. Reference CO₂ values were taken from the Siple Station ice core (Friedli et al., 1986) for the period 1856-1957 and from direct Mauna Loa measurements for the period 1958-2000 (Keeling et al., 1995). Measurements were also made on *Ginkgo* saplings growing in CO₂-controlled greenhouses after two growing seasons at 350 and 560 ppmv (Beerling et al., 1998) and after one growing season for 6- (*Ginkgo*) and 1-year-old (*Ginkgo* and *Metasequoia*) saplings at 430 and 790 ppmv [see Beerling et al. (1998) for experimental methodology]. Fields of view were concentrated near the centers of leaves in the intercostals, which have been shown in other species to yield the least variation in SI (Chapter 1). Although stomata occur in rows in *Metasequoia*, the field of view chosen (0.1795 mm²) nearly spanned the distance between the midrib and margin, and thus the stomatal bands should not confound our results.

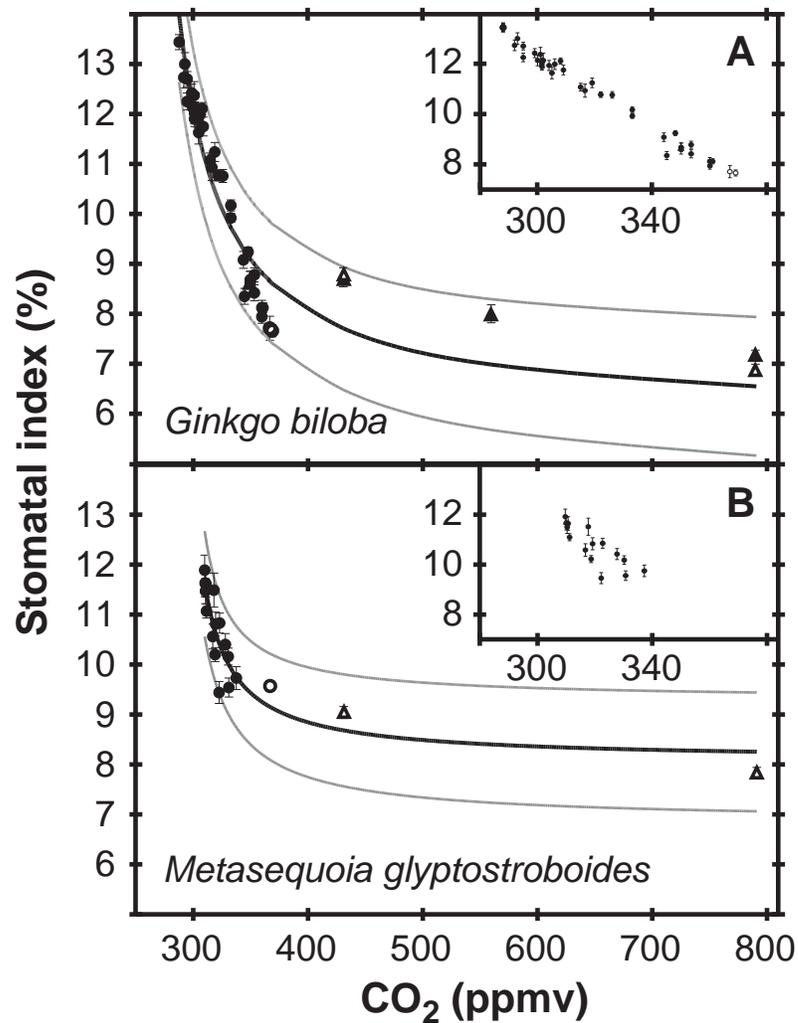


Fig. 3.1. Training sets for (A) *Ginkgo biloba* ($n = 40$) and (B) *Metasequoia glyptostroboides* ($n = 18$). Thick black lines represent regressions {*Ginkgo*: $r^2 = 0.91$, $F(1, 37) = 185$, $P < 0.0001$, $SI = [CO_2 - 194.4] / [(0.16784) \times CO_2 - 41.6]$; *Metasequoia*: $r^2 = 0.85$, $F(1,15) = 41$, $P < 0.0001$, $SI = [CO_2 - 274.5] / [(0.12373) \times CO_2 - 35.3]$ }. Gray lines represent $\pm 95\%$ prediction intervals. Inset graphs show the linear portions of both response curves in greater detail. Stomatal index determined from herbarium sheets (\bullet), fresh samples from living trees (\circ), and 6- (\blacktriangle) and 1-year-old (\triangle) saplings growing in CO_2 -controlled greenhouses. Error bars represent standard errors.

size and decreased sensitivity at high CO₂ for both *Ginkgo* and *Metasequoia*, paleo-CO₂ estimates >400 ppmv should be considered semi-quantitative.

To reconstruct atmospheric CO₂ changes, we measured the SI of fossil *Ginkgo* and *Metasequoia* cuticles from 24 localities in western North America and one from the Isle of Mull (Scotland), and then calibrated these data against the modern training set (Fig. 3.1) using inverse regression². Although not tightly constrained, the paleoelevations for all of the sites were probably <1000 m. This elevation difference could increase our estimates of CO₂ concentration by at most 10%, and so our conversion from partial pressure to concentration excludes any correction. Except for a single high CO₂ value near the Paleocene/Eocene boundary, all of our reconstructed CO₂ concentrations lie between 300 and 450 ppmv (Fig. 3.2; Table 3.1). These contrast with two other *Ginkgo*-based CO₂ estimates for the late Paleocene and middle Miocene that are very high (4500 and 2100 ppmv, respectively) (Retallack, 2001; see Footnote 3).

² Measurements of SI were made on fossil cuticles on each of 5 to 22 leaves per site (Table 1) using epifluorescence (for *Ginkgo*) and transmitted light (for *Metasequoia*) microscopy, with count replication and field sizes as in footnote 1. Inverted regression for *Ginkgo*: $CO_2 = [1 - (0.1564) \times SI] / [(0.00374 - (0.0005485) \times SI)]$, $r^2 = 0.84$, $F(1,37) = 91$, $P < 0.0001$; and for *Metasequoia*: $CO_2 = (SI - 6.672) / [(0.003883) \times SI - 0.02897]$, $r^2 = 0.98$, $F(1,15) = 336$, $P < 0.0001$. Standard errors in SI for fossil sites are ~0.20%. Fossil *Ginkgo* leaves preserved in environments that today host well-watered, open canopy forests (Chapter 2). Such conditions should minimize both the influence of confounding variables in the SI-CO₂ signal and the environmental differences between the training and fossil datasets. It is possible a species effect on SI exists between *G. gardneri* at Ardtun Head and *G. adiantoides* at the other sites, however a difference in SI of 6 (Table 1) is highly unlikely based on studies of intra-generic SI variation in modern plants (Chapter 1). All Bighorn Basin sites are dated using Age Model 2 of Wing et al. (2000), while the Ardtun Head site is dated assuming it is stratigraphically near the latest Paleocene carbon isotope excursion (55.2 Ma using Age Model 2). For details of dating methods for the other sites, see Chapter 4.

³ These two CO₂ estimates are based upon small sample sizes ($n = 4$ cuticle fragments each) and an extrapolation of a training set calibrated only up to 560 ppmv.

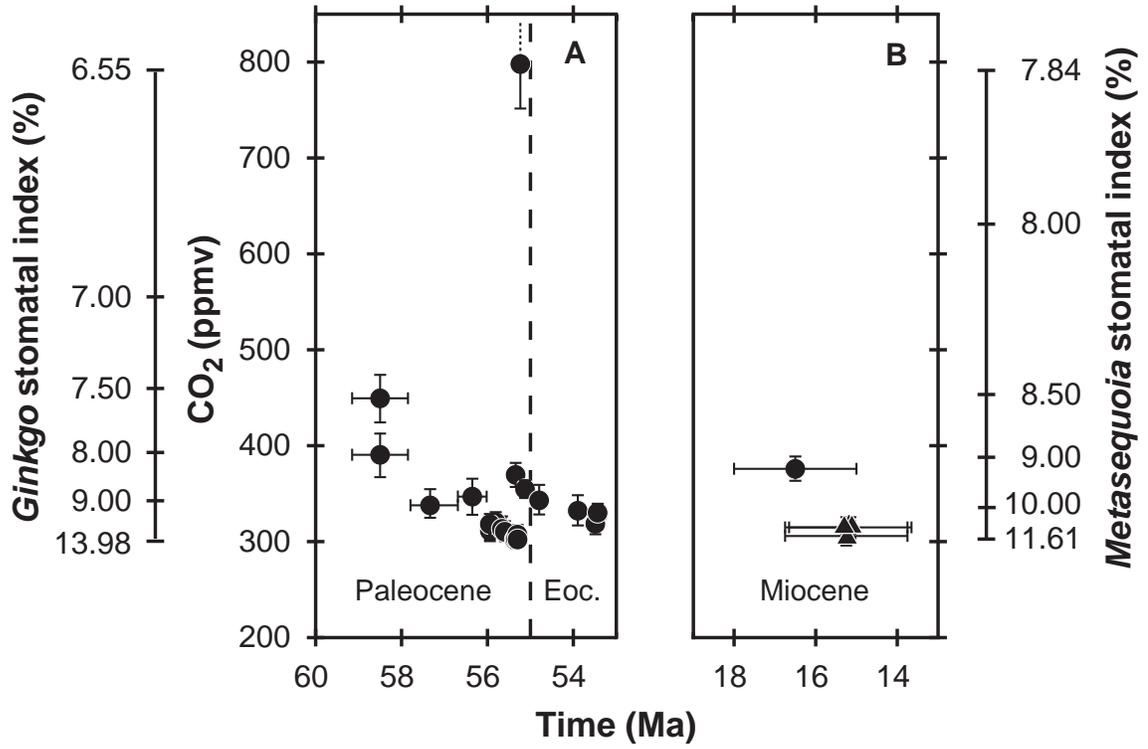


Table 3.1. Summary of fossil data. n = number of leaves measured for calculation of SI. A = south-central Alberta (Canada), BHB = Bighorn Basin (Wyoming and Montana, USA), M = Isle of Mull (UK), I = north-central Idaho (USA). Dashes indicate that no analyses were performed. BOM = bulk organic matter. $\delta^{13}\text{C}_{\text{om}} = \delta^{13}\text{C}$ of organic matter. See Footnote 4 for $\delta^{13}\text{C}$ methodology.

Site	Location	Age (Ma)	n	SI (%)	CO ₂ (ppmv)	$\delta^{13}\text{C}_{\text{om}}$ (V-PDB)
<i>Ginkgo</i>						
Burbank	A	58.5	7	7.55	450	–
Joffre Bridge	A	58.5	5	7.96	391	–
SLW 0025	BHB	57.3	7	9.01	340	–
LJH 7132	BHB	56.4	5	8.75	348	-23.91
SLW 991	BHB	55.9	5	10.97	314	–
SLW 992	BHB	55.9	8	10.80	316	-25.66
SLW 993	BHB	55.9	8	11.43	311	–
LJH 72141–1	BHB	55.8	12	10.63	317	–
SLW 9155	BHB	55.7	10	11.21	313	-29.80
SLW 9411	BHB	55.6	8	11.50	311	-23.77
SLW 9434	BHB	55.4	7	12.23	307	-22.53
SLW 9715	BHB	55.3	12	8.23	371	-24.17
SLW 9050	BHB	55.3	5	12.18	308	-26.99
SLW 9936	BHB	55.3	15	11.77	310	–
SLW 8612	BHB	55.3	7	12.41	307	–
Ardtun Head	M	55.2	13	6.54	798	-24.68
AR-2 (BOM)						-24.56
AR-6 (BOM)						-25.82
AR-8 (BOM)						-24.17
AR-10 (BOM)						-24.26
AR-15 (BOM)						-24.54
SLW 9812	BHB	55.1	22	8.53	356	-24.49
SLW 9915	BHB	54.8	8	8.83	345	–
SLW LB	BHB	53.9	5	9.29	334	–
SLW H	BHB	53.5	9	10.22	321	-26.93
LJH 9915	BHB	53.4	15	9.38	332	-26.33
Juliaetta (P6)	I	16.5	14	8.14	377	–
<i>Metasequoia</i>						
Clarkia (P33a)	I	15.3	6	11.59	307	–
Clarkia (P33b)	I	15.3	10	10.94	316	–
P37a	I	15.2	10	10.95	316	–

We have *Metasequoia*-derived CO₂ estimates only from the warm interval of the middle Miocene, but these are similar to coeval estimates derived from *Ginkgo* cuticles. The convergence of these two independent estimates increases our confidence that both species are reliably recording paleoatmospheric CO₂ levels. In addition, the measured SI values from most sites fall well inside the region of high CO₂ sensitivity in the training sets (Fig. 3.1; Table 3.1), and the 95% confidence intervals (± 50 ppmv or less) are over an order of magnitude lower than the errors associated with early Tertiary CO₂ estimates from geochemical models (Tajika, 1998; Berner and Kothavala, 2001; Wallmann, 2001) and other proxies (Ekart et al., 1999; Pearson and Palmer, 2000). Furthermore, middle Paleocene to early Eocene CO₂ reconstructions based on pedogenic carbonate (Ekart et al., 1999; see Footnote 4) and marine boron isotopes (Pearson and Palmer, 2000) show

⁴ In addition to the published estimates compiled in Ekart et al. (1999), we also made seven new CO₂ estimates based on our record of cuticle $\delta^{13}\text{C}$ (Table 3.1) and the $\delta^{13}\text{C}$ of pedogenic carbonate ($\delta^{13}\text{C}_{\text{cc}}$) from sites stratigraphically within 15 m (~30 ky) of our cuticle-bearing sites. This affords the rare opportunity to apply multiple CO₂ proxies to sediments in close geographic and stratigraphic proximity.

Stable carbon isotope analyses were conducted on *Ginkgo* cuticle fragments at all sites, as well as on bulk organic matter at Ardtun Head. Cuticle fragments were separated from host rock with either tweezers or by placing drops of 52% HF along the cuticle edges. Bulk organic matter was prepared by reacting finely ground host rock with 38% HCl for 6 hours, centrifuging, reacting with fresh HCl for an additional 24 hours, then repeating with 5:1 52% HF:38% HCl. Carbon dioxide was generated for isotopic analysis by combustion in sealed, evacuated VYCOR tubes at 910 °C for 1 hour in the presence of CuO and Cu metal. After cryogenic distillation, CO₂ was analyzed on a gas source mass spectrometer. Data are reported relative to V-PDB. Replicate analyses of a laboratory gelatin standard analyzed with these samples had a standard deviation of 0.05‰ ($n = 9$).

CO₂ calculated using Equation 1 in Ekart et al. (1999) and a planktonic foraminifera-derived atmospheric $\delta^{13}\text{C}$ value of -5.6‰ , and assuming the concentration of CO₂ contributed by soil = 5000 ppmv and soil temperature = 25 °C (Ekart et al., 1999). At sites SLW 826, 882, and 8822, we analyzed bulk cuticle, not just *Ginkgo* cuticle. The site names from which the cuticle and corresponding pedogenic carbonate were collected, $\delta^{13}\text{C}_{\text{cc}}$ values, and CO₂ estimates ($\pm \sim 500$ ppmv) are as follows: LJH 7132, SC 85 & 185, -7.95‰ , 1185 ppmv; SLW 992, SC 92, -7.82‰ , 2041 ppmv; SLW 9715, SC 22, -7.69‰ , 1448 ppmv; SLW 9812, SC 4, -8.36‰ , 1217 ppmv; SLW 826 (54.8 Ma, $\delta^{13}\text{C}_{\text{om}} = -$

large changes in CO₂ (≥ 2000 ppmv) over geologically brief periods of time [< 1 million years (My)] (Fig. 3.3) that cannot be readily explained. In contrast, the highly constrained error ranges and consistency among near time-equivalent estimates suggest that our SI-derived CO₂ reconstruction is presently the most reliable, particularly for the middle Paleocene to early Eocene.

A period of rapid climatic warming (~ 2 °C global mean rise within 10^4 years that lasted 10^5 years) near the Paleocene/Eocene boundary has been extensively documented (Kennett and Stott, 1991; Koch et al., 1992; Huber and Sloan, 1999). Although the leading hypothesis for the cause of most of this warming is the rapid release of methane from marine gas hydrates and its subsequent oxidation to CO₂ in the atmosphere and ocean (Dickens et al., 1995; Norris and Röhl, 1999), all previous attempts to resolve this possible atmospheric CO₂ spike have failed (Koch et al., 1992; Stott, 1992; Sinha and Stott, 1994). Our single high CO₂ estimate is based on *G. gardneri* cuticle from Ardtun Head, Isle of Mull. Anomalously low $\delta^{13}\text{C}_{\text{om}}$ values (-30‰), an influx of the marine dinocyst *Apectodinium*, and a thermophyllic flora (including *Caryapollentites veripites* and *Alnipollenites verus*) occur in stratigraphically equivalent sediments elsewhere on Mull. Together, these indicate a possible correlation with a section of a Paris Basin borehole that has been calibrated to this event (Thiry et al., 1998). At Ardtun Head, however, we failed to capture the negative carbon isotope excursion globally associated with this event, which ranges from 2.5‰ in the deep ocean (Kennett and Stott, 1991; Norris and Röhl,

28.65‰), YPM 200+4m, -11.88‰ , 1152 ppmv; SLW 882 (53.5 Ma, $\delta^{13}\text{C}_{\text{om}} = -27.71\text{‰}$) & SLW H & LJH 9915, YPM 320 & D1217+10m, -11.96‰ , 577 ppmv; SLW 8822 (52.8 Ma, $\delta^{13}\text{C}_{\text{om}} = -28.71\text{‰}$), Fern Q lower, -10.01‰ , 2034 ppmv.

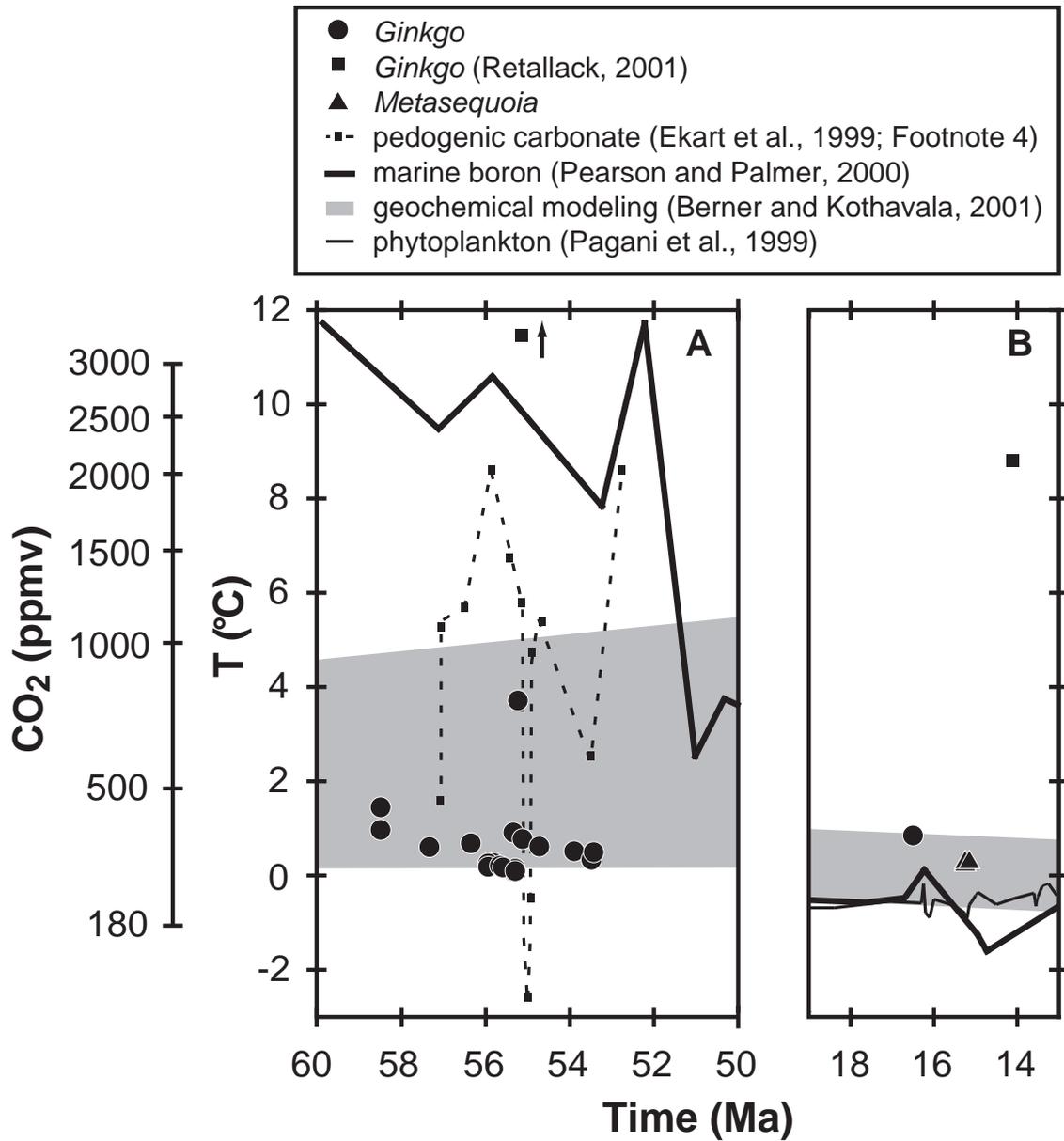


Fig. 3.3. Estimates of paleo-CO₂ concentration derived from a variety of methods and their corresponding model-determined temperature departures (T) of global mean surface temperature (GMST) from present day for the (A) middle Paleocene to early Eocene and (B) middle Miocene. Paleo-GMST calculated from paleo-CO₂ estimates using the CO₂-temperature sensitivity study of Kothavala et al. (1999). Present-day reference GMST calculated using the pre-industrial CO₂ value of 280 ppmv (14.7 °C). The error range of GMST predicted from the geochemical modeling-based CO₂ predictions of Berner and Kothavala (2001) corresponds to the model's sensitivity analysis

1999) to as much as 6‰ on land (Koch et al., 1992) (Table 3.1). Although the precise age of the Ardtun Head site remains uncertain, using a global carbon isotope mass balance model calibrated to Paleocene/Eocene conditions (Beerling, 2000), our reconstructed CO₂ increase (500 ppmv) is consistent with a release of 2522 Gt of methane-derived carbon, a value close to the estimate (2600 Gt C) calculated to account for the marine carbon isotopic excursion using methane as the carbon source (Dickens, 2001).

Carbon dioxide is an important greenhouse gas, and its effect on global mean surface temperature (GMST) can be quantified with general circulation models (GCMs) [e.g., Kothavala et al. (1999)]. Using the model output of Kothavala et al. (1999) we predicted GMST from our CO₂ results. The GCM used by Kothavala et al. is calibrated to the present day, which allows us to test the effect of CO₂ on GMST independent of any paleogeographic or vegetational changes. With the exception of the single value near the Paleocene/Eocene boundary, all predictions lie within 1.5 °C of the pre-industrial GMST (Fig. 3.3). These predictions contrast sharply with most paleoclimatic interpretations for these time intervals. For example, based on a synthesis of global late Paleocene and early Eocene δ¹⁸O-derived sea surface temperature data, Huber and Sloan (1999) estimated that GMST was 3° to 4 °C higher than today at this time, and δ¹⁸O-derived temperature estimates for the mid-Miocene thermal maximum [17 to 14.5 million years ago (Ma)] indicate that deep and high-latitude surface ocean temperatures were as much as 6 °C warmer than today (Savin et al., 1975).

As a cross-check on our results, we compared our GMST predictions with those based on a geochemical carbon cycle model and other CO₂ proxies for these same time periods. With the exception of the one stomatal-based CO₂ estimate by Retallack (2001),

there is very good agreement among the methods for the middle Miocene (Fig. 3.3), strongly suggesting that factors in addition to CO₂ are required to explain this brief warm period. In contrast, a large disagreement (10 °C or greater) exists for the middle Paleocene to early Eocene (Fig. 3.3). This discrepancy is largely driven by the high CO₂ estimates derived from marine boron isotopes (Pearson and Palmer, 2000); however, this proxy is probably less accurate than the other methods (Lemarchand et al., 2000; Royer et al., 2001). Nevertheless, even if the boron-based predictions are discounted, a large range still exists among the remaining three methods. This is striking considering that many of the pedogenic carbonate-derived CO₂ estimates are based on the same sediments as our stomatal-based estimates (Footnote 4); however, these estimates show a large temporal variability (-60 to 2040 ppmv) and are associated with relatively large error ranges (\pm 500 ppmv). If our low SI-based temperature predictions are correct, additional factors such as paleogeography, enhanced meridional heat transport, and high latitude vegetation feedbacks are required to explain this warm period, and new constraints for CO₂ levels are established for middle Paleocene to early Eocene and middle Miocene GCMs [e.g., Sloan and Rea (1995)]. Understanding the mechanisms of climate change will become increasingly important in the near future as atmospheric CO₂ levels climb to levels perhaps unprecedented for the last 60 My.

CHAPTER 4

Estimating Latest Cretaceous and Tertiary Atmospheric CO₂ Concentration from Stomatal Indices

4.1 Introduction

Globally averaged surface temperatures have risen sharply in the last century (~1 °C in the Northern Hemisphere) (Mann et al., 1998, 1999), concomitant with a 30% increase in the concentration of the greenhouse gas CO₂ (Friedli et al., 1986). It is now clear that CO₂ is largely responsible for this warming (Mann et al., 1998; Crowley, 2000; Barnett et al., 2001; Harries et al., 2001; Levitus et al., 2001; Mitchell et al., 2001). On the longer timescale of 10⁵ years, temperature and atmospheric CO₂ have been tightly coupled for at least the last four glacial-interglacial cycles (Petit et al., 1999), and it appears that CO₂ has also been driving these temperature variations (Shackleton, 2000).

On multimillion year timescales, proxy- and modeling-based estimates of CO₂ over the Phanerozoic largely correlate with geologic indicators of continent-scale glaciations, which are the most reliable index for the presence of ‘greenhouse’ or ‘icehouse’ climates (Crowley and Berner, 2001). This correlation is not present, however, during certain intervals. Proxy-based CO₂ estimates for the mid-Miocene thermal maximum (17-14.5 Ma), a brief interval of global warmth relative to today, suggest concentrations largely lower than the pre-industrial value of 280 ppmV (Pagani et al., 1999; Pearson and Palmer, 2000). In addition, CO₂ estimates derived from multiple

proxies for other time periods are not consistent. For example, early Tertiary (65-50 Ma) CO₂ estimates range from <300 ppmV to >3000 ppmV (Eckart et al., 1999; Pearson and Palmer, 2000).

Here I apply a CO₂-calibrated set of stomatal indices from *Ginkgo biloba* and *Metasequoia glyptostroboides* to reconstruct paleo-CO₂ levels for the very latest Cretaceous to early Eocene (66-53 Ma) and the middle Miocene (~15.5 Ma). This study expands on the data presented in Chapter 3 and Beerling et al. (2002).

4.2 The stomatal index method

Most modern vascular C₃ plants show an inverse relationship between the partial pressure of atmospheric CO₂ and stomatal index (Woodward, 1987; Beerling, 1999; Lake et al., 2001; Chapter 1). Stomatal index (SI) is the percentage of epidermal cells that are stomatal packages (guard cells + stomatal pore), and is defined as the following (Salisbury, 1927):

$$SI = \frac{SD}{SD + ED} \times 100$$

where SD = stomatal density (mm⁻²) and ED = non-stomatal epidermal cell density (mm⁻²). The mechanism(s) linking CO₂ to SI is not fully understood, but probably involves the strong selective pressure to maximize carbon fixation per unit of water transpired (water-use efficiency) (e.g., Woodward, 1987). For example, in a rising CO₂ regime plants can increase their water-use efficiency by reducing their stomatal conductance, thereby reducing water loss. Stomatal conductance is largely a function of

stomatal pore area, which in turn is controlled by the size of individual stomatal pores and the density of those pores. On the timescale of several growing seasons or longer, stomatal density is typically more sensitive to changes in atmospheric CO₂ than individual pore size (e.g., Chapter 1). Furthermore, on multimillion year timescales stomatal density and SI across all plant taxa inversely respond to CO₂ (Beerling and Woodward, 1997; McElwain, 1998; Chapter 1).

In contrast to stomatal density, SI is area independent, which normalizes for the effects of cell expansion. Stomatal index is essentially a measure of the number stomatal packages that develop per unit of non-stomatal epidermal cells. Since plant water-potential strongly influences cell expansion [e.g., water stressed plants generally reduce the size of their epidermal cells (Chapter 1)], but not the rate of stomatal initialization, changes in a plant water budget can affect stomatal density, but not stomatal index. Experimental studies show that SI is largely independent of water potential, irradiance, and temperature, and is primarily a function of CO₂ (e.g., Beerling, 1999; Chapter 1).

Stomatal indices (and stomatal densities) have been experimentally shown to respond to the partial pressure of CO₂, not mole fraction (Woodward and Bazzaz, 1988). This finding is corroborated by a positive correlation in many plants between elevation and stomatal density (Körner and Cochrane, 1985; Woodward, 1986; Woodward and Bazzaz, 1988; Beerling et al., 1992) and SI (Rundgren and Beerling, 1999). It is important, therefore, to control for elevation when estimating CO₂ from stomatal properties. The relationship between elevation and partial pressure in the lower atmosphere is roughly as follows:

$$P = -10.6E + 100$$

where E = elevation in kilometers and P = the percentage of partial pressure relative to sea level. If one does not control for elevation (i.e., assumes concentration = partial pressure), CO₂ concentration estimates will be underestimated by 3% for plants growing at 250 m elevation, for example, and 11% for plants at 1000 m.

4.2.1 Choice of species

Stomatal responses to atmospheric CO₂ are usually species-specific (Woodward and Kelly, 1995; Beerling and Kelly, 1997; Chapter 1). For example, based upon genera represented by >1 species in the compilation in Chapter 1 ($n = 7$), only one is internally consistent ($\alpha = 0.05$; chi-squared test) with respect to SI. In order to quantitatively reconstruct paleo-CO₂, then, the SI-CO₂ relationship in an extant species must be applied in the fossil record to the same species. This requirement has previously hindered the applicability of this method for fossil leaves pre-dating the late Miocene (10 Ma) (van der Burgh et al., 1993; Kürschner et al., 1996). Here I use the lineages represented today by *Ginkgo biloba* and *Metasequoia glyptostroboides*, which greatly extends the temporal applicability of the method.

Morphologically indistinguishable forms of *G. biloba* extend back to the early Cretaceous, leading many authors to consider the fossil and modern forms conspecific (Tralau, 1968; see Chapter 2). This fossil form is most commonly identified as *G. adiantoides*, however many other designations are considered equivalent, e.g., *G. spitsbergensis* (Tralau, 1968). In fact, the only Tertiary-aged ginkgo form that may deviate sufficiently from the *adiantoides-biloba* form to warrant a separate species class is *G. gardneri* (Manum, 1966; Tralau, 1968), which occurs at one early Paleogene site on

the Isle of Mull, Scotland (e.g., Boulter and Kvaček, 1989). Further evidence for conspecificity of the Tertiary-aged and modern forms comes from a paleoecological study of *Ginkgo* (Chapter 2). Based primarily upon the localities used in this study, the distribution of *Ginkgo*'s sedimentological contexts (e.g., stream margin, crevasse splay, backswamp) and floral associates remain largely unchanged throughout the Tertiary, suggesting a conservative phylogeny.

Chen et al. (2001) report that the SI in modern *G. biloba* is statistically independent of leaf size, long vs. short shoots, and male vs. female trees, and only minor differences (<5%) due to canopy position and the timing of leaf development are present. These results further suggest that the SI's in *Ginkgo* are reliable indicators of atmospheric CO₂ concentration.

Ginkgo foliage is very distinctive and readily identified. A *Ginkgo* leaf consists of a petiole and fan-shaped dichotomously veined lamina (Fig. 4.1). The veins only very rarely anastomose (Arnott, 1959). Vein spacing is typically ~1 mm. Stomata are found only on the abaxial side of the leaf (hypostomatous), and are distributed randomly in the intercostal regions. Subsidiary cells are moderately papillose, which partially cover the stomatal pore. The leaf is usually divided into two lobes, whose margins are wavy. The petiole consists of two vascular strands, each of which vascularize one of the lobes (Chamberlain, 1935; Gifford and Foster, 1989). Mucilage cavities are common and conspicuous, even in fossils. They are 1-5 mm in length, and are oriented parallel to venation (Chamberlain, 1935). The *adiantoides-biloba* form is differentiated from other ginkgo-forms (which largely occur earlier in the rock record) by its less dissected lamina (Tralau, 1968), however deeply lobed leaves do occur occasionally in *G. biloba* (Critchfield, 1970). The only other plants with open dichotomizing venation and bilobed

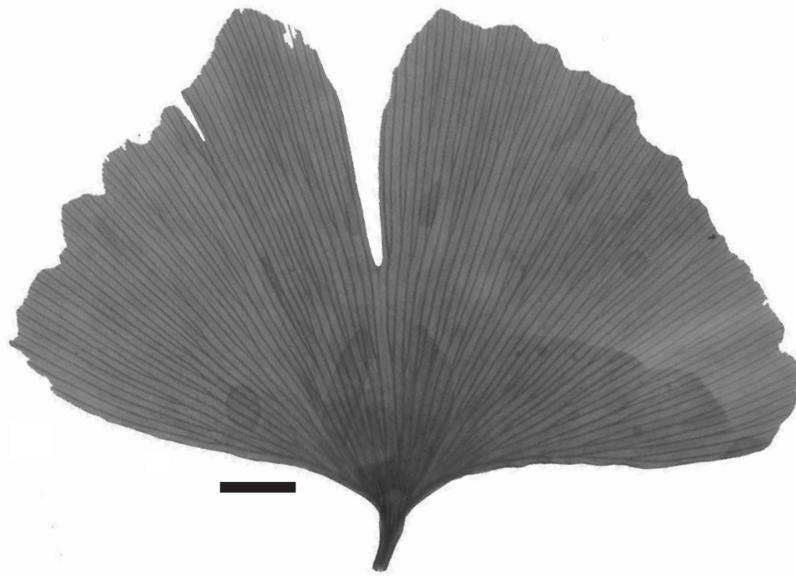


Fig. 4.1. A leaf of *Ginkgo biloba*. Collected from New Haven, Connecticut (USA). Scale bar = 1 cm.

lamina are ferns such as *Adiantum* (e.g., Gifford and Foster, 1989). These can be distinguished from *Ginkgo* by the single vascular strand in their petioles, the presence of sori abaxially near the tips of their leaves, and their more closely spaced veins.

Metasequoia also has a long fossil record of indistinguishable forms back to the Late Cretaceous (Liu et al., 1999). Many workers consider this fossil form, *M. occidentalis*, very closely related to, or conspecific with the modern *M. glyptostrobooides* (Chaney, 1951; Christophel, 1976; Liu et al., 1999). *Metasequoia* needles are flat and ~15-20 mm in length and ~2 mm in width. The leaf morphology of *Metasequoia* is very similar to *Sequoia* and *Taxodium*, however *Metasequoia* needles are arranged decussately opposite while the needles in the other two genera are spirally alternate. *Metasequoia* also tends to have more rounded tips, a higher angle of divergence between the leaf and shoot (~90°), and a more highly twisted base than *Sequoia* and *Taxodium* (Chaney, 1951).

The fossil records of *Ginkgo* and *Metasequoia* indicate that both were components of mid- to high latitude temperate forests (Tralau, 1967; Mai, 1994). In contrast to multi-story tropical forests, intracanalopy CO₂ gradients are small in temperate forests and should not bias stomatal-based CO₂ estimates (Chapter 1).

4.2.2 Construction of training set

Leaves from dated herbarium sheets of *Ginkgo biloba* and *Metasequoia glyptostrobooides* were used to assess the response of their SI's to the anthropogenic rise in atmospheric CO₂ concentration over the last 145 years. Because both *Ginkgo* and *Metasequoia* have deciduous leaf habits, all leaves are assumed to have grown only during the year recorded on the sheets. The CO₂ concentrations that the leaves developed

in were taken from ice core-derived (Friedli et al., 1986) and direct atmospheric (Keeling et al., 1995) measurements. The geographic origins of the leaves for both species range from the United States, China, and Japan. Most material came from cultivated individuals. All trees grew at <250 m in elevation, with 84% of them growing at elevations <75 m. Therefore, no correction is needed to convert CO₂ partial pressure to concentration. For each herbarium sheet, five mature leaves were randomly selected. Stomatal index was calculated for three fields of view (0.1795 mm² each) on every leaf. Measurements were made in the intercostal regions near the centers of the leaves, which in other species show the least variation in SI (Salisbury, 1927; Chapter 1). A combination of acetate peels and cleared leaves using light microscopy, and unaltered leaf tissue using epifluorescence microscopy was employed. No systematic variation in SI among the three preparation techniques was noted. During the growing season of 1999 (and 2000 for *Ginkgo*), SI was measured from trees growing in New Haven (Connecticut, USA), Ann Arbor (Michigan, USA), and London (UK). Sampling details are given in Table 4.1.

Stomatal indices were also measured on saplings growing in greenhouses at four discrete CO₂ concentrations. For each CO₂ treatment, three potted saplings were placed in each of four independent greenhouses [see Beerling et al. (1998) and Beerling and Osborne (2002) for details]. For the 430 / 445 ppmV and 790 / 800 ppmV treatments, SI was measured after a CO₂ exposure time of both one and two growing seasons. The *Metasequoia* saplings were one-year-old after their first season of CO₂ treatments. Two sets of *Ginkgo* saplings were used, aged one- and six-years-old after their first season of CO₂ treatments. For the 350 and 560 ppmV treatments, SI was determined from three-year-old *Ginkgo* saplings after a CO₂ exposure time of two growing seasons (Beerling et al., 1998). Sampling details are given in Table 4.1.

TABLE 4.1. DETAILS OF SAMPLING FOR TRAINING SETS
A. HISTORICAL COLLECTIONS

Species	Year(s) Collected	CO ₂ (ppmV)	Mode of Preparation*	Trees†	Leaves†	Field of view‡
<i>G. biloba</i>	1856-1996	288-361	E	1	5	15
	1999	367	A	6	13	46
	2000	369	C	3	10	30
<i>M. glyptostroboides</i>	1947-1980	310-338	A, C	1	5	15
	1999	367	A	4	21	46

B. GREENHOUSE EXPERIMENTS

Species	Exposure to CO ₂ (years)	CO ₂ (ppmV)	Mode of Preparation*	Trees†	Leaves†	Field of view‡
<i>G. biloba</i>	2	350§	A	8	16	160
	1	430#	C	8	16	48
	1	430**	C	8	8	24
	2‡	445#	C	8	8	24
	2‡	445**	C	8	8	24
	2	550§	A	8	16	160
	1	790#	C	8	16	48
	1	790**	C	7	7	21
	2‡	800#	C	8	8	24
	2‡	800**	C	8	8	24
<i>M. glyptostroboides</i>	1	430**	C	8	16	48
	2‡	445**	C	7	7	21
	1	790**	C	8	16	48
	2‡	800**	C	8	8	24

* A = acetate peels (light microscopy); C = cleared leaves (light microscopy); E = unaltered leaves (epifluorescence microscopy).

† Total number per CO₂ value or herbarium sheet.

§ From Beerling, McElwain, and Osborne (1998). Three year old saplings.

Six-year-old (for 430 and 790 ppmV treatments) and seven-year-old (for 445 and 800 ppmV treatments) saplings.

** One-year old (for 430 and 790 ppmV treatments) and two-year-old (for 445 and 800 ppmV treatments) saplings.

‡ Includes the exposure time from the previous growing season (either 430 or 790 ppmV).

4.2.3 Application of fossil cuticles

Stomatal indices were calculated from 27 very latest Cretaceous to middle Miocene-aged *Ginkgo* cuticle-bearing sites and three middle Miocene-aged *Metasequoia* cuticle-bearing sites. The application of both *Ginkgo* and *Metasequoia* cuticles from near coeval middle Miocene-aged sites provides a check on the reliability of the method. As with the training set material, SI was determined for three field of views on each fossil leaf. At least five leaves were measured per site (see Table 4.2). This ensures that at least as many leaves per site were used to reconstruct paleo-CO₂ than were used per herbarium sheet to establish the training sets. For *Ginkgo*, epifluorescence microscopy on unaltered cuticle was employed. Fossil *Metasequoia* cuticle failed to fluoresce, and so leaves were first cleared and then analyzed using light microscopy. Voucher specimen information for both the training and fossil datasets are available from the author.

The paleoelevations of all regions sampled in this study are most likely <1000 m. For example, the late Cretaceous Hell Creek Formation in the Williston Basin contains marine and brackish facies (Frye, 1969; Murphy et al., 1999), indicating paleoelevations near sea level. In the Bighorn Basin, which produced the largest number of fossil cuticle sites, there are few constraints on paleoelevation [see discussion in Chase et al. (1998)]. The most liberal estimates for the basin during this time range from 1500 to 2500 m (Chase et al., 1998), however other workers consider these estimates to be too high because they think it unlikely that climate at these higher elevations would be warm enough to support the large number of subtropical and frost intolerant plants that are known from basinal deposits (S. L. Wing, personal communication, 2001). Due to the lack of reliable estimates for the Bighorn Basin, and the inferred low paleoelevations for

TABLE 4.2. SAMPLING DETAILS AND CO₂ RECONSTRUCTION FOR FOSSIL DATA
A. *GINKGO*

Site	Location*	Depository†	Age (Ma)	n§	SI (%)	CO ₂ (ppmV)
DMNH 566	ND	DMNS	65.5	31	8.32	385
LJH 7423	BHB	USNM	64.5	5	9.48	339
LJH 7659	BHB	YPM	64.5	15	9.32	344
DMNH 2360	DB	DMNS	64.1	5	9.90	329
Basilika	S	PMO	64.0	8	9.42	341
Burbank	A	UA	58.5	7	7.55	451
Joffre Bridge	A	UA	58.5	5	7.96	409
SLW 0025	BHB	USNM	57.3	7	9.01	353
LJH 7132	BHB	YPM	56.4	5	8.75	363
SLW 991	BHB	USNM	55.9	5	10.97	312
SLW 992	BHB	USNM	55.9	8	10.80	314
SLW 993	BHB	USNM	55.9	8	11.43	307
LJH 72141-1	BHB	USNM, YPM	55.8	12	10.63	317
SLW 9155	BHB	USNM	55.7	10	11.21	309
SLW 9411	BHB	USNM	55.6	8	11.50	306
SLW 9434	BHB	USNM	55.4	7	12.23	299
SLW 9715	BHB	USNM	55.3	12	8.23	390
SLW 9050	BHB	USNM	55.3	5	12.18	300
SLW 9936	BHB	USNM	55.3	15	11.77	303
SLW 8612	BHB	USNM	55.3	7	12.41	298
Ardtun Head	M	BNHM, PMO, YPM	55.2	13	6.54	826
SLW 9812	BHB	USNM	55.1	22	8.53	373
SLW 9915	BHB	USNM	54.8	8	8.83	360
SLW LB	BHB	USNM	53.9	5	9.29	345
SLW H	BHB	USNM	53.5	9	10.22	323
LJH 9915	BHB	YPM	53.4	15	9.38	342
Juliaetta (P6)	I	YPM	16.5	14	8.14	396

B. *METASEQUOIA*

Clarkia (P33a)	I	YPM	15.3	6	11.59	310
Clarkia (P33b)	I	YPM	15.3	10	10.94	316
Emerald Creek (P37a)	I	YPM	15.2	10	10.95	316

* ND = southwestern North Dakota (Williston Basin); BHB = Bighorn Basin; DB = Denver Basin; S = south-central Spitsbergen; A = south-central Alberta; M = Isle of Mull; I = north-central Idaho.

† Depository for cuticle preparates: DMNS = Denver Museum of Nature and Science; USNM = National Museum of Natural History, Smithsonian Institution; YPM = Yale Peabody Museum; PMO = Oslo Paleontological Museum; UA = University of Alberta; BNHM = British Natural History Museum.

§ Number of leaves measured for calculation of SI.

the other sites used in this study, no corresponding corrections were applied to the CO₂ estimates.

Most (88%) *Ginkgo*-bearing sites used in this study have been interpreted as representing streamside and crevasse splay environments (Chapter 2). These highly disturbed environments usually host well-watered, open canopy forests. Mesic conditions should only improve the fidelity of the SI-CO₂ signal, while open canopies should remove any potential intra-canopy CO₂ gradients and provide a closer analog to the open grown trees used to construct the training set.

The following sources were used to date the fossil sites: Williston Basin (Hell Creek Formation)—Hicks et al. (2002); Bighorn Basin (Fort Union and Willwood Formations)—Wing et al. (1995) and Age Model 2 in Wing et al. (2000); Denver Basin (Dawson Formation)—Raynolds et al. (2001, p. 25); Spitsbergen (Firkanten Formation)—Manum (1963) and Kvaček et al. (1994); Alberta (Paskapoo Formation)—Fox (1990); Isle of Mull (Staffa Group)—Chapter 3; Idaho (Latah Formation)—Reidel and Fecht (1986).

4.2.4 *Stomatal ratio method*

An alternative, semi-quantitative, stomatal-based CO₂ proxy compares the SI in a fossil species to its nearest living equivalent (NLE) (McElwain and Chaloner, 1995, 1996; McElwain, 1998; McElwain et al., 1999). NLE's are defined ecologically and structurally, not taxonomically. The stomatal ratios of the extant:fossil species are directly translated into CO₂ estimates in an inverse linear fashion, such that a stomatal ratio of three equates to a CO₂ estimate of 1050 ppmV (assuming the leaves of the extant trees grew in a 350

ppmV CO₂ atmosphere). This method is not species-specific, and so can be applied to sediments dating back to the early Devonian (McElwain and Chaloner, 1995), however it assumes that the SI's in the related sets of species respond to CO₂ in the following hyperbolic fashion: $\text{CO}_2 \propto (\text{SI})^{-1}$.

To provide an independent check on the early Paleogene *Ginkgo*-derived CO₂ estimates, the stomatal indices of *Platanus guillelmae* from the *Ginkgo*-bearing site SLW H were compared to modern *P. occidentalis* and *P. orientalis* (and their hybrid *P. × acerfolia*) using the stomatal ratio method. *P. occidentalis* and *P. orientalis* are morphologically similar to the broadly trilobed *P. guillelmae*, and both the modern and fossil species prefer disturbed streamside environments (Chapter 2).

4.3 Results and discussion

The stomatal indices in both *Ginkgo* and *Metasequoia* show a strong linear response to the anthropogenic increases in atmospheric CO₂ over the last 145 years ($r^2 = 0.98$, $P < 0.001$ for *Ginkgo*; $r^2 = 0.68$, $P < 0.001$ for *Metasequoia*). For CO₂ concentrations above present-day levels, the responses are nonlinear (Fig. 4.2; Table 4.3). This type of nonlinear response is predicted by plant physiological principles (Kürschner et al., 2001) and supported by recent genetic work (Gray et al., 2000). Overall, both nonlinear regressions are highly significant (see Fig. 4.2).

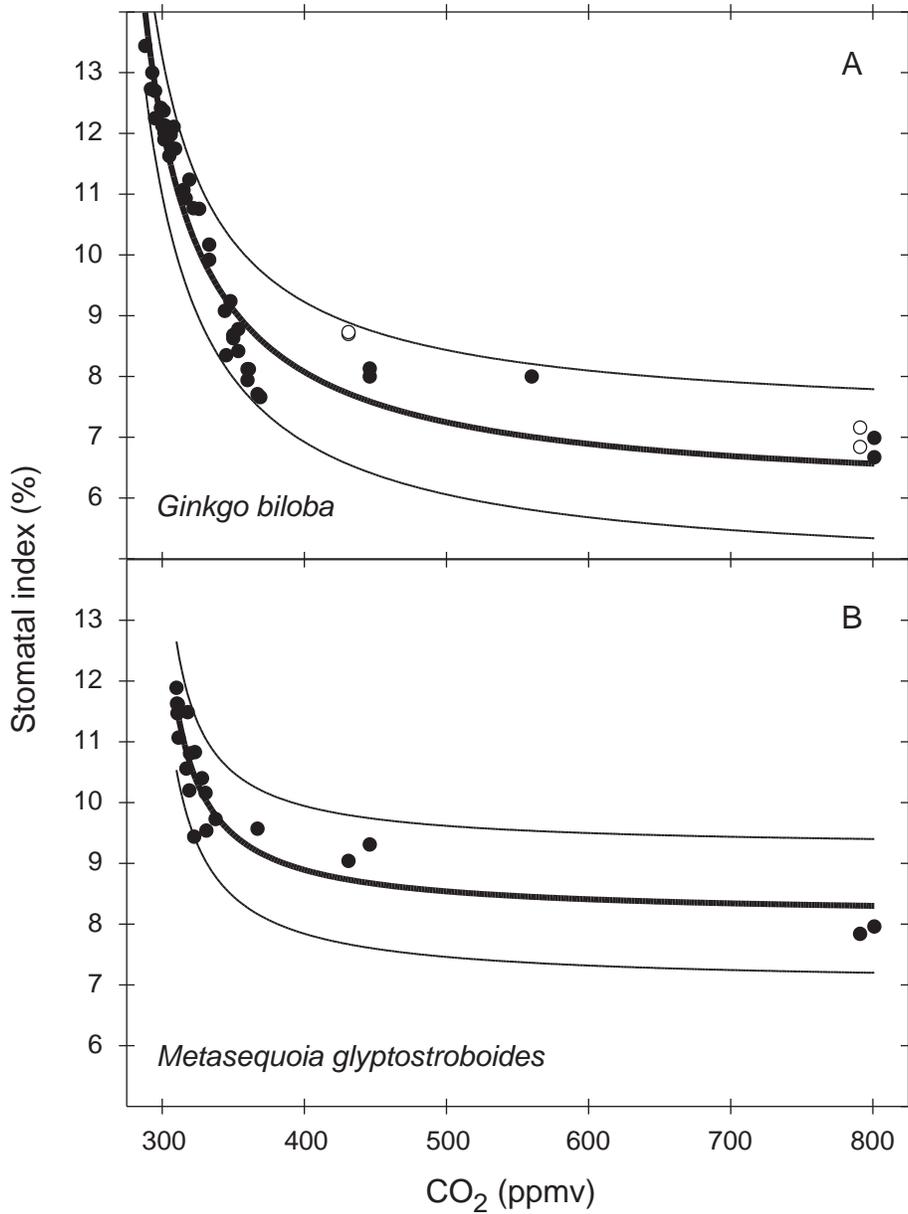


Fig. 4.2. Training sets (●,○) for (A) *Ginkgo biloba* ($n = 44$) and (B) *Metasequoia glyptostroboides* ($n = 20$). Thick lines represent regressions {*Ginkgo*: $r^2 = 0.93$, $F(1,37) = 243$, $P < 0.0001$, $SI = 2000 \times [CO_2 - 196.1] / [333.7 \times CO_2 - 83000]$; *Metasequoia*: $r^2 = 0.86$, $F(1,17) = 54$, $P < 0.0001$, $SI = 10000 \times [CO_2 - 273.7] / [1230.1 \times CO_2 - 350000]$ }. Thin lines represent $\pm 95\%$ prediction intervals. For *Ginkgo*, experimental results based upon a CO_2 exposure time of one growing season (○) were not used in regression (see text).

TABLE 4.3. RAW DATA FOR TRAINING SETS
A. *GINKGO*

Date	CO ₂ (ppmV)	SI (%)	Herbarium*	Source
1856	288	13.44	YPM	N.D.
1880	292	12.73	YPM	Hartford, CT
1883	293	13.00	MBG	Botanic Garden, Cambridge, MA
1900	295	12.25	NA	Washington, DC
1900	295	12.70	YPM	Yale Forestry School, New Haven, CT
1910	299	12.42	BBG	Orange, NJ
1915	300	12.13	MBG	Hannibal, MO
1918	301	12.37	BBG	Arnold Arboretum, Boston, MA
1920	302	11.90	MGB	Nanking, Kiangsu Province, China
1921	302	12.02	MGB	Missouri Botanical Garden, St. Louis, MO
1922	302	12.13	BBG	Purdys, NY
1928	304	11.93	NA	Maymont Park, Richmond, VA
1932	305	11.63	MA	Arnold Arboretum?, Boston, MA
1937	306	11.98	MBG	Japan
1942	308	12.11	NA	Berkeley, CA
1943	309	11.75	MBG	Japan
1958	315	11.07	MBG	Tower Grove Park, St. Louis, MO
1960	317	10.93	MBG	Peking, China
1965	319	11.24	MA	Arnold Arboretum, Boston, MA
1968	322	10.77	NA	Arnold Arboretum, Boston, MA
1971	326	10.76	MA	Knox College, Galesburg, IL
1977	333	9.92	NA	U.S. Capitol, Washington, DC
1977	333	10.17	NA	Blandy Experimental Farm, Boyce, VA
1984	344	9.08	MBG	China (Yellow Plateau Team)
1985	345	8.35	BBG	Brooklyn Botanical Garden, NYC, NY
1987	348	9.24	NA	Morton Arboretum, Lisle, IL
1988	350	8.68	MBG	Clemson University, Clemson, SC
N.A.	350	8.63	exp	3 year old saplings in greenhouses† (2§)
1990	354	8.42	MBG	Missouri Botanical Garden, St. Louis, MO
1990	354	8.78	MBG	Oomoto Kameyama Botanical Garden; Kyoto, Japan
1995	360	7.94	YPM	Yale Peabody Museum, New Haven, CT
1995	360	8.12	MBG	Missouri Botanical Garden, St. Louis, MO
1996	361	8.12	MBG	Oomoto Kameyama Botanical Garden; Kyoto, Japan
1999	367	7.71	modern	New Haven, CT
2000	369	7.66	modern	New Haven, CT
N.A.	431	8.70	exp	6 year old saplings in greenhouses (1§)
N.A.	431	8.73	exp	1 year old saplings in greenhouses (1§)
N.A.	446	8.13	exp	7 year old saplings in greenhouses (2§)
N.A.	446	8.00	exp	2 year old saplings in greenhouses (2§)
N.A.	560	8.00	exp	3 year old saplings in greenhouses† (2§)
N.A.	791	7.16	exp	6 year old saplings in greenhouses (1§)
N.A.	791	6.84	exp	1 year old saplings in greenhouses (1§)
N.A.	801	6.99	exp	7 year old saplings in greenhouses (2§)
N.A.	801	6.67	exp	2 year old saplings in greenhouses (2§)

TABLE 4.3 CONTINUED. RAW DATA FOR TRAINING SETS
B. *METASEQUOIA*

Date	CO ₂ (ppmV)	SI (%)	Herbarium*	Source
1947	310	11.89	UC	E Szechuan / NW Hubei, China
1948	310	11.63	UC	E Szechuan / NW Hubei, China
1949	311	11.47	UC	Berkeley, CA
1950	311	11.62	UC	Bar Harbor, ME
1953	312	11.07	UC	San Rafael, CA
1961	317	10.56	UC	Tallahassee, FL
1962	318	11.49	YPM	Bethany, CT
1964	319	10.20	MA	PA
1965	320	10.81	UC	Washington, DC
1968	323	9.44	UC	Fukuchiyama-city, Japan
1969	323	10.83	BBG	Brooklyn Botanical Garden, NY
1973	328	10.40	NA	Turlock, CA
1975	331	10.16	UC	University of Oregon, Eugene, OR
1976	331	9.54	UC	University of Oregon, Eugene, OR
1980	338	9.73	UC	Western Hubei, China
1999	367	9.57	modern	New Haven, CT; London, UK
N.A.	431	9.04	exp	1 year old saplings in greenhouses (1§)
N.A.	446	9.31	exp	2 year old saplings in greenhouses (2§)
N.A.	791	7.84	exp	1 year old saplings in greenhouses (1§)
N.A.	801	7.96	exp	2 year old saplings in greenhouses (2§)

Notes: N.D. = no data; N.A. = not applicable.

* YPM = Peabody Museum of Natural History (Yale University); MGB = Missouri Botanical Gardens; NA = U.S. National Arboretum; BBG = Brooklyn Botanic Gardens; MA = Morris Arboretum (University of Pennsylvania); UC = University Herbarium (University of California, Berkeley); modern = collections of fresh leaves; exp = collections from greenhouses.

† From Beerling, McElwain, and Osborne (1998).

§ Number of growing seasons saplings were exposed to CO₂ treatments.

A discontinuity exists in the *Ginkgo* training set between the experimentally-derived SI measurements at elevated CO₂ (>370 ppmV) and the rest of the training set (which includes one experimentally-derived measurement at 350 ppmV) (Fig. 4.2). Plants often require >1 growing season for their SI's to respond to CO₂ concentrations above present-day levels (Chapter 1). The stomatal indices in *Ginkgo* saplings exposed to CO₂ treatments for two growing seasons are indeed slightly lower than the same plants after one growing season (see Fig. 4.2; Table 4.3), and in the case of the 440 ppmV CO₂ treatment the difference is significant [$F(1,118) = 10.61$; $P < 0.001$; one-way ANOVA]. While not conclusive, this suggests that the discontinuity is an artifact of insufficient CO₂ exposure time.

The CO₂ reconstruction based upon 30 *Ginkgo*- and *Metasequoia*-bearing fossil sites is shown in Fig. 4.3 and Table 4.2. Except for one estimate near the Paleocene/Eocene boundary, all reconstructed CO₂ concentrations (300-450 ppmV) are close to present-day values. Estimates based upon <5 cuticle fragments are less reliable (Beerling and Royer, 2002a), however in general they conform with their more reliable coeval estimates (Table 4.4). It is interesting to note that the estimates from the two oldest very latest Cretaceous sites both suggest CO₂ levels in excess of 500 ppmV, and Upper Pliocene cuticles register CO₂ concentrations of 300 ppmV (Table 4.4).

There is some stratigraphic evidence that the site associated with the one high CO₂ estimate (Arduin Head, Isle of Mull, Scotland) dates to the Paleocene/Eocene Thermal Maximum (PETM; previously known as LPTM) (Chapter 3). The PETM was brief (~10⁵ years; Kennett and Stott, 1991; Bains et al., 1999; Norris and Röhl, 1999) but significant [global mean surface temperatures rose ~2 °C (Huber and Sloan, 1999)]. Concomitant with warming, there was a worldwide negative excursion in δ¹³C, ranging in magnitude

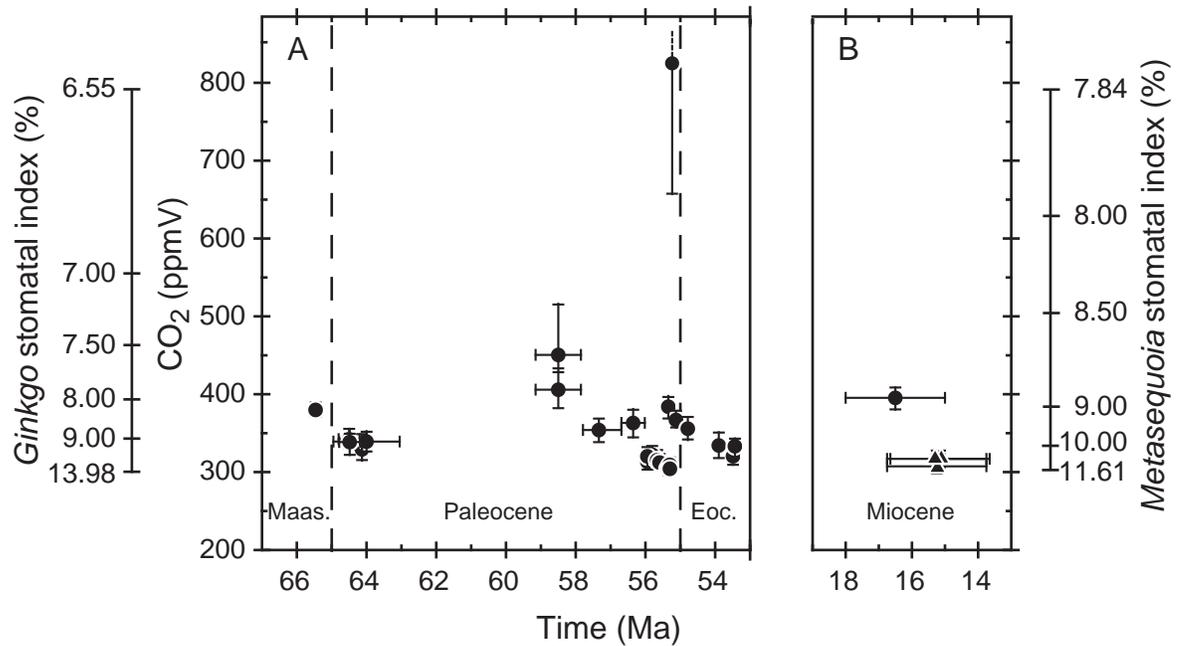


Fig. 4.3. Reconstruction of paleo-CO₂ for the (A) very latest Cretaceous to early Eocene and (B) middle Miocene based upon SI measurements of *Ginkgo* (●) and *Metasequoia* (▲) fossil cuticles. Errors represent ±95% confidence intervals. Regressions used to solve for CO₂ were rewritten from the training set-derived regressions [*Ginkgo*: $CO_2 = 2000 \times (415 \times SI - 1961) / (3337 \times SI - 20000)$; *Metasequoia*: $CO_2 = 70000 \times (50 \times SI - 391) / (12301 \times SI - 100000)$]

TABLE 4.4. SAMPLING DETAILS AND CO₂ RECONSTRUCTION FOR FOSSIL GINKGO DATA BASED UPON SITES WITH <5 CUTICLE FRAGMENTS

Site	Location*	Depository†	Age (Ma)	n§	SI (%)	CO ₂ (ppmV)
DMNH 571	ND	DMNS	65.9	1	7.03	554
KJ 87134	ND	YPM	65.8	3	7.09	536
DMNH 1489	ND	DMNS	65.4	2	8.42	379
N.D.	BHB	YPM	~63	3	8.60	370
UF 18255	PRB	FMNH	~58	3	11.24	309
DMNH 907	ND	DMNS	56	2	8.89	358
LJH 8411	E	YPM	55	2	8.29	386
Klärbecken	G	BNHM	~ 2	3	12.29	299

Notes: N.D. = no data.

* ND = southwestern North Dakota (Williston Basin); BHB = Bighorn Basin; PRB = Powder River Basin; E = Ellesmere Island; G = Germany.

† Depository for cuticle preparates: DMNS = Denver Museum of Nature and Science; YPM = Yale Peabody Museum; FMNH = Florida Museum of Natural History; BNHM = British Natural History Museum.

§ Number of leaves measured for calculation of SI.

from -2.5‰ in the benthic marine record (Kennett and Stott, 1991; Bains et al., 1999) to -6‰ in the terrestrial record (Koch et al., 1992; Sinha and Stott, 1994; Bowen et al., 2001). The leading hypothesis to explain this event involves the dissociation of isotopically light ($\sim -60\text{‰}$) methane hydrates along continental margins (Dickens et al., 1995; Bains et al., 1999; Katz et al., 1999) and their subsequent oxidation to carbon dioxide. However, all previous attempts to discern this hypothesized atmospheric CO_2 spike using other CO_2 proxies have failed (Koch et al., 1992; Stott, 1992; Sinha and Stott, 1994). While it is tempting to correlate Ardtun Head and its corresponding 500 ppmV spike in atmospheric CO_2 to the PETM, further stratigraphic work is required to resolve this issue. If it is not of PETM age, then it is recording another high CO_2 event that is probably not associated with massive methane hydrate dissociation.

4.3.1 *Ginkgo and Metasequoia cuticle as reliable recorders of atmospheric CO_2*

There are numerous factors indicating that the CO_2 reconstruction presented here is accurate. As discussed above, both genera are restricted in the fossil record to mid- and high latitude temperate forests, and, in addition, *Ginkgo* prefers disturbed riparian settings. These characteristics should strengthen the applicability of the fossil cuticles to their respective training sets. Also, both *Ginkgo* and *Metasequoia* are very phylogenetically conservative, and in the case of *Ginkgo* the slope of the response in experimental treatments after one growing season is similar to that derived from a multimillion year sequence of fossil SI measurements and coeval pedogenic carbonate-derived CO_2 estimates (Beerling and Royer, 2002b). This suggests that: 1) The SI- CO_2 relationships in *Ginkgo* and *Metasequoia* have remained unchanged throughout the Cenozoic; and 2) The

short-term (largely phenotypic) stomatal responses to CO₂ captured in the training sets are similar to the long-term (largely genotypic) responses reflected in the fossils.

At least as many leaves per fossil site were used to reconstruct CO₂ as were used per data point to construct the bulk of the training sets (compare Table 4.1 with Table 4.2). This contrasts with other studies, where <5 fossil leaves are commonly measured per fossil site (e.g., Retallack, 2001). In addition, it is likely that fossil cuticles measured for a given site stemmed from multiple trees, whereas SI's from only one tree were normally measured per data point in the training sets. In this sense, then, the sampling methodology for the fossil cuticles was at least as conservative than for the modern cuticles.

I have measured *Metasequoia* cuticles from only the middle Miocene, but these CO₂ estimates agree reasonably well with their near-coeval *Ginkgo*-based estimates (Fig. 4.3; Table 4.2). This provides further support that both species are reliably recording atmospheric CO₂. As a cross-check on the early Paleogene *Ginkgo*-derived estimates, I compared the SI's of *Platanus guillelmae* from site SLW H with the SI's of its nearest living equivalents *P. occidentalis* and *P. orientalis* using the stomatal ratio method (see above). While this method can only generate semi-quantitative estimates of CO₂, all of the estimates for this site fall between 350 and 390 ppmV (Table 4.5). This range compares favorably with the *Ginkgo*-derived CO₂ estimate (323 ppmV), indicating, again, that *Ginkgo* faithfully records CO₂.

The SI's of nearly all the measured fossil material fall within the range of high CO₂ sensitivity in the training sets (Fig. 4.2; Table 4.2). This contrasts with the study of Retallack (2001), where the stomatal indices in 82% of the fossil sites fall outside the range captured in his training set, and so these CO₂ estimates are less reliable. In addition, I have intensively sampled several intervals, for example 12 sites from the last 1 m.y. of

TABLE 4.5. CO₂ RECONSTRUCTION FROM STOMATAL RATIOS IN *PLATANUS*A. FOSSIL *PLATANUS*

Species	Site	Age (Ma)	<i>n</i> *	SI (%)
<i>P. guillelmae</i>	SLW H	53.5	7	12.89

B. MODERN *PLATANUS*

Species	Site	Date	<i>n</i> *	SI (%)	Stomatal ratio†	Paleo-CO ₂ (ppmV)§
<i>P. occidentalis</i>	North Carolina, USA	1891	6	15.35	1.19	350
<i>P. orientalis</i>	Iraq	1975	5	15.22	1.18	390
<i>P. acerfolia</i>	Connecticut, USA	2001	5	13.28	1.03	381
<i>P. occidentalis</i>	Connecticut, USA	2001	6	13.44	1.04	386

* Number of leaves measured for calculation of SI.

† Stomatal ratio = $SI_{\text{modern}} / SI_{\text{fossil}}$

§ CO₂ reconstructions for site SLW H. These estimates should be considered semi-quantitative only. Values calculated by multiplying the stomatal ratio by the corresponding CO₂ concentration in which the leaves developed. See text for details.

the Paleocene. These characteristics allow for more precise determinations of paleo-CO₂ relative to other CO₂ proxies, and, because SI's respond to CO₂ on the timescale of 10⁰-10² years, the ability to discern rapid shifts in atmospheric CO₂ (Royer et al., 2001).

A few of the fossil SI's fall outside the densely sampled regions in the training sets, and in the case of the Ardtun Head site, nearly outside the range captured in the entire *Ginkgo* training set. This suggests that the SI's in *G. biloba* and *G. adiantoides* are not constrained to a limited range (8-12%) irrespective of atmospheric CO₂ levels. Additional evidence that the SI's in *Ginkgo* are highly adaptable comes from Paleozoic and Mesozoic studies where SI's in *Ginkgo* are as low as 2.6% (McElwain and Chaloner, 1996; McElwain et al., 1999; Chen et al., 2001; Retallack, 2001).

The only possible confounding factor that I have discerned in my fossil dataset is latitude (Fig. 4.4). Except for the Spitsbergen site, there is a negative correlation between latitude and the SI in *Ginkgo*. Even within the densely sampled Bighorn Basin, there is a small (slope = -2.2 using SI as the dependent variable) but moderately significant ($r^2 = 0.43$; $P = 0.002$) correlation. Differences in irradiance (Lake et al., 2001) or temperature (Wagner, 1998) may be driving this response, however more data are required before any firm conclusions can be drawn.

4.3.2 Paleoclimatic implications

Fig. 4.5 is a compilation of most published proxy- and modeling-based CO₂ estimates for the last 66 m.y. There is very good agreement among the methods for the Neogene, strongly indicating that CO₂ levels were not much different from the present-

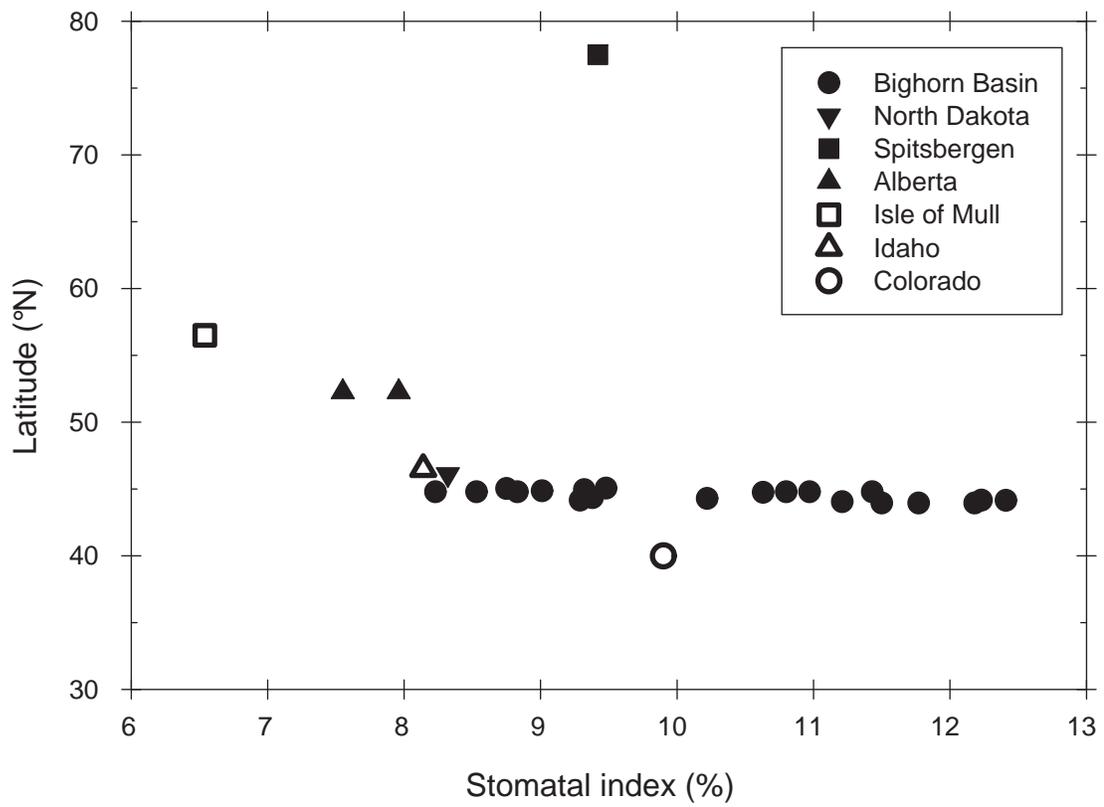


Fig. 4.4. Correlation between latitude and stomatal index for fossil sites.

Present-day latitudes are plotted, but should reflect their corresponding paleolatitudes to within 5° (Schettino and Scotese, 2001).

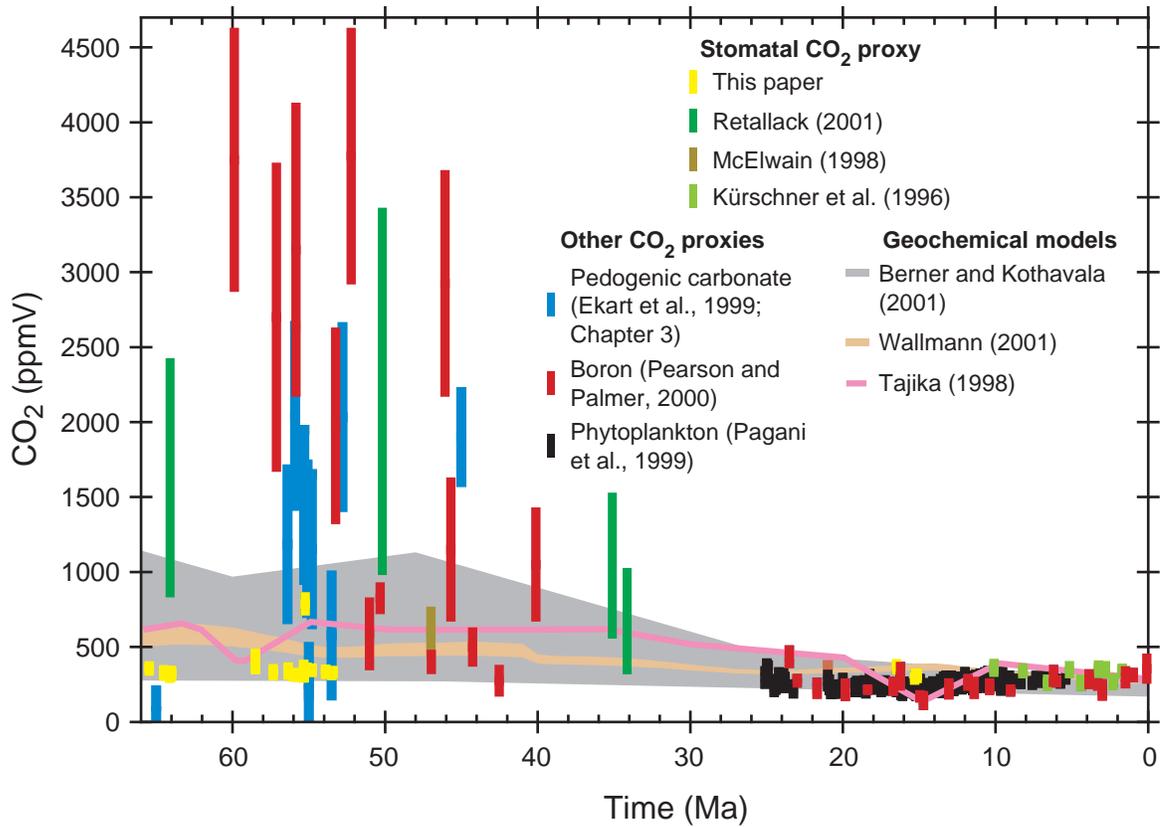


Fig. 4.5. A compilation of CO₂ reconstructions for the Tertiary. In the case of Retallack (2001), only those sites for which > 4 cuticle fragments were counted are plotted. No pedogenic carbonate-derived estimates are plotted for the Neogene because their margins of error are much larger relative to the other methods (Royer et al., 2001).

day. CO₂ reconstructions for the Paleogene, however, are inconsistent, ranging from <300 ppmV to >3000 ppmV. These inconsistencies are largely stratified by method, with the highest CO₂ estimates derived from the boron proxy and the lowest from this stomatal study (Fig. 4.5).

What do these CO₂ reconstructions mean in terms of paleoclimate? Assuming that we understand the role of CO₂ as a greenhouse gas in regulating global temperature, it is possible to convert a given concentration of atmospheric CO₂ into a prediction of global mean surface temperature (GMST). I have done this for the late Cretaceous to early Paleogene (69-50 Ma) and middle Miocene (19-13 Ma) CO₂ reconstructions, the two intervals for which I have stomatal data, using the general circulation model (GCM)-based CO₂-GMST sensitivity study of Kothavala et al. (1999). This GCM study was calibrated to present-day conditions. While it may be instructive to use sensitivity studies calibrated to early Paleogene and middle Miocene conditions, the CO₂-GMST relationship is most accurately known for present-day conditions, and applying a sensitivity study calibrated to the present-day allows testing the sole effects of CO₂ on GMST. In other words, if the CO₂-derived GMST predictions do not match the geologic indicators of GMST for a given period, then factors in addition to CO₂ such as paleogeography and vegetative feedbacks are required to explain the GMST for that period.

The CO₂-derived GMST predictions are shown in Fig. 4.6. As with the raw CO₂ data, there is general agreement for the middle Miocene, with most predictions within 1 °C of the present-day GMST. Predictions for the early Paleogene are not consistent, ranging from near present-day values to over 10 °C warmer than the present-day GMST.

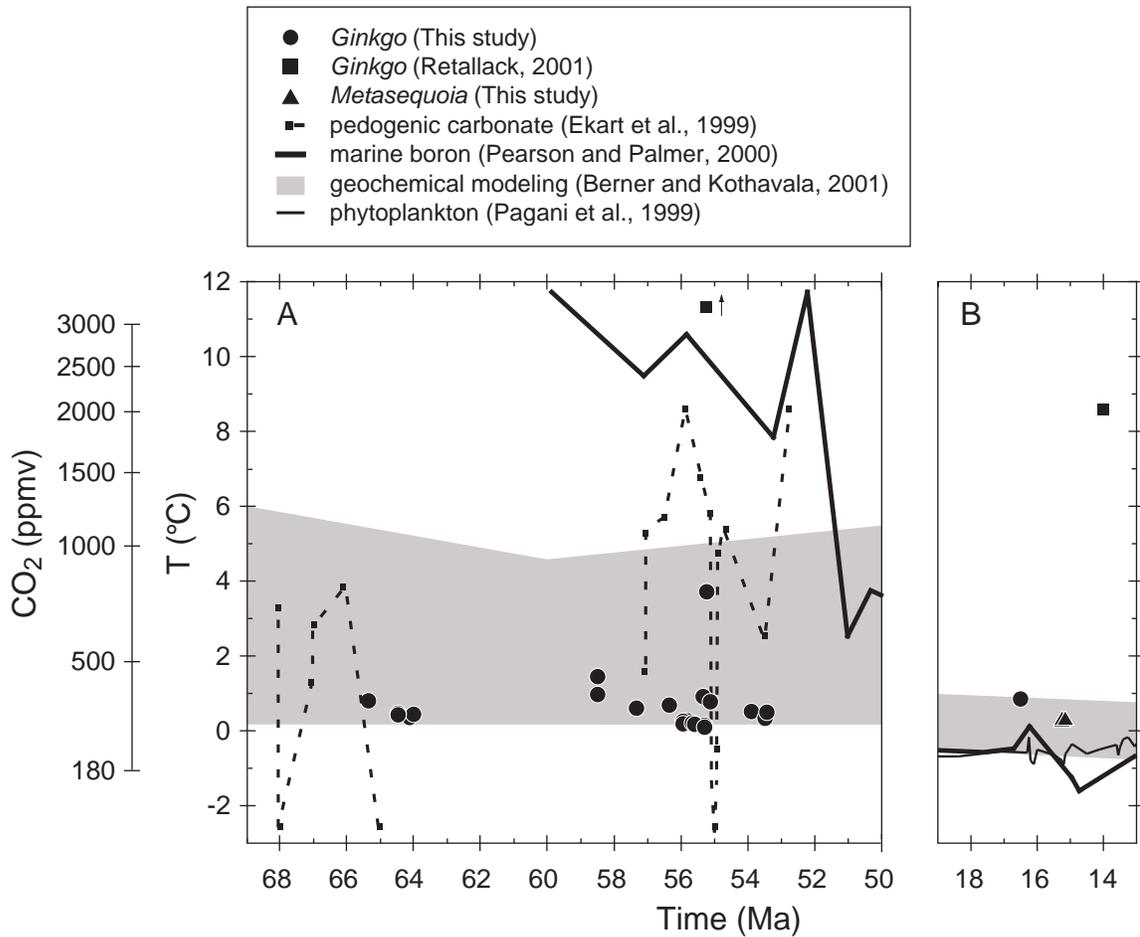


Fig. 4.6. Estimates of CO₂ from Fig. 4.5 and their corresponding model-determined temperature departures (ΔT) of global mean surface temperature (GMST) from present day for the (A) very latest Cretaceous to early Eocene and (B) middle Miocene. Paleo-GMST calculated using the CO₂-temperature sensitivity study of Kothavala et al. (1999). Present-day reference GMST calculated using the pre-industrial CO₂ value of 280 ppmv (14.7 °C). The error range of GMST predicted from the geochemical modeling-based CO₂ predictions of Berner and Kothavala (2001) corresponds to the model's sensitivity analysis.

Both the early Paleogene and middle Miocene are intervals of global warmth relative to today. The physiognomic characters of early Paleogene fossil floras indicate temperatures much higher than today at mid and high latitudes (e.g., Hickey 1980; Spicer and Parrish, 1990; Wing and Greenwood, 1993; Wolfe, 1994; Greenwood and Wing, 1995; Wilf, 2000). The comparison of these fossil plants with their Nearest Living Relatives (NLRs) also indicates warmer temperatures (e.g., Hickey, 1977; Spicer and Parrish, 1990). For example, frost intolerant palms occur at paleolatitudes up to 20° north of their current northern limit in North America (Greenwood and Wing, 1995). Animals also yield paleoclimatic information; for example, the early Paleogene distributions of large tortoises (Hutchison, 1982) and crocodylians (Markwick, 1994, 1998) in the United States indicate temperatures higher than the present-day.

While the above data constrain continental temperatures, $\delta^{18}\text{O}$ data from calcitic marine shells provide temperature information for the oceans. Oceans are the largest reservoirs of heat in the biosphere, and therefore provide the best proxy for globally averaged temperatures. Based upon the $\delta^{18}\text{O}$ compilations of Zachos et al. (1994) and Zachos et al. (2001), benthic temperatures were 8-12 °C warmer than the present-day during the early Paleogene, and high latitude near surface temperatures were upwards of 15 °C warmer. Using late Paleocene near surface paleotemperature data and present-day levels of atmospheric CO_2 as input into a GCM, O'Connell et al. (1996) calculated a GMST 3 °C warmer than the present-day. During the mid-Miocene thermal maximum (~17-14.5 Ma), benthic waters warmed 1-2 °C to temperatures 6 °C warmer than the present-day (Lear et al., 2000; Zachos et al., 2001). Plant distributions (White et al., 1997) and leaf physiognomic characters (Wolfe, 1994) also indicate substantial warming in

North America. If my stomatal-based CO₂ reconstruction is correct, then, factors in addition to carbon dioxide are required to explain these two intervals of global warmth, and these intervals may not serve as good analogs for understanding future climate change. It is crucial to emphasize that low concentrations of atmospheric CO₂ during intervals of global warmth do not counter the theory of the greenhouse effect (for this, high CO₂ levels are required during globally cool intervals); instead, they indicate that additional forcings are required.

Traditionally, GCM modelers interested in warm climates have struggled to match their output with the geologic data (Barron, 1987; Sloan and Barron, 1990, 1992; Sloan et al., 1995), particularly when using low CO₂ levels (Sloan and Rea, 1995). Recent work, however, has begun to resolve these discrepancies (Otto-Bliesner and Upchurch, 1997; Sloan and Pollard, 1998; DeConto et al., 1999; Upchurch et al., 1999; Sewall et al., 2000; Sloan et al., 2001). It will be crucial to understand the various roles CO₂ plays during globally warm periods, particularly as atmospheric CO₂ rises to levels in the very near future that are perhaps unprecedented for the past 65 m.y.

APPENDIX 1.1

Experimental Stomatal Responses

Experiment Length (days)	CO ₂ Levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
?	↑ 300%	<i>Phaseolus vulgaris</i>	abaxial	*↓ 9%	-	O'Leary and Knecht, 1981 ^b
			adaxial	↔	-	
14	↑2067%	<i>Marsilea vestita</i>	abaxial	*↓ 91%	-	Bristow and Looi, 1968 ^b
			adaxial	↔	-	
	↑~10 ⁵ %	<i>Marsilea vestita</i>	abaxial	*↓ 99%	-	
			adaxial	↔	-	
15	↑ 100%	<i>Populus euroamericana</i>	-	↑ 38%	↔	Gaudillère and Mousseau, 1989 ^b
20	↑ 80%	<i>Phaseolus vulgaris</i>	abaxial	↔	↔	Ranasinghe and Taylor, 1996 ^b
			adaxial	↔	↔	
21	↑ 86%	<i>Tradescantia (fluminensis?)</i>	abaxial	↔	↔	Boetsch et al., 1996 ^b
21	↓ 29%	<i>Vaccinium myrtillus</i>	abaxial	↔	↔	Woodward, 1986 ^b
			adaxial	*↑ 548%	*↑ 424%	
	↑ 29%	<i>Vaccinium myrtillus</i>	abaxial	↔	↔	
			adaxial	↔	↔	
21	↓ 34%	<i>Acer pseudoplatanus</i>	abaxial	*↑ 220%	*↑ 122%	Woodward and Bazzaz, 1988 ^b
		<i>Geum urbanum</i>	abaxial	*↑ 31%	*↑ 18%	
			adaxial	*↑ 214%	*↑ 191%	
		<i>Quercus robur</i>	abaxial	*↑ 131%	*↑ 81%	
		<i>Rhamnus catharticus</i>	abaxial	*↑ 117%	*↑ 100%	
		<i>Rumex crispus</i>	abaxial	*↑ 71%	*↑ 31%	
			adaxial	*↑ 150%	*↑ 400%	
	↑ 100%	<i>Amaranthus retroflexus</i>	abaxial	*↓ 35%	*↓ 23%	
			adaxial	*↓ 38%	*↓ 26%	
		<i>Ambrosia artemisiifolia</i>	abaxial	*↓ 11%	↔	

Experiment Length (days)	CO ₂ Levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
		<i>Setaria faberii</i>	adaxial	*↓ 24%	*↓ 25%	
			abaxial	↔	*↓ 21%	
			adaxial	*↓ 22%	*↓ 21%	
21-35	↑ 94%	<i>Lolium perenne</i>	adaxial	↔	-	Ryle and Stanley, 1992 ^b
26	↑ 86%	<i>Lycopersicon esculentum</i>	abaxial	*↓ 17%	↔	Madsen, 1973 ^b
			adaxial	*↓ 14%	↔	
	↑ 814%	<i>Lycopersicon esculentum</i>	abaxial	*↓ 23%	↔	
			adaxial	*↓ 36%	↔	
~28	↑ 33%	<i>Lolium temulentum</i>	adaxial	↔	-	Gay and Hauck, 1994 ^b
28	↑ 100%	<i>Phaseolus vulgaris</i>	abaxial	↔	↔	Radoglou and Jarvis, 1992 ^c
			adaxial	↔	↔	
~40	↑ 91%	<i>Raphanus raphanistrum</i>	abaxial	↔	↔	Case et al., 1998 ^b
45	↑ 71%	<i>Anthyllis vulneraria</i>	abaxial	*↓ 32%	*↓ 17%	Ferris and Taylor, 1994 ^b
			adaxial	↔	↔	
		<i>Lotus corniculatus</i>	abaxial	↑ 60%	↔	
			adaxial	↑ 40%	↔	
		<i>Plantago media</i>	abaxial	*↓ 20%	↔	
			adaxial	*↓ 36%	*↓ 12%	
		<i>Sanguisorba minor</i>	abaxial	↑ 175%	↑ 36%	
			adaxial	↑ 150%	↑ 213%	
45	↑ 100%	<i>Vicia faba</i>	abaxial	↔	↔	Radoglou and Jarvis, 1993 ^c
			adaxial	↔	↔	
45	↑ 168%	<i>Glycine max</i>	abaxial	↑ 38%	↔	Thomas and Harvey, 1983 ^c
			adaxial	↔	↔	
		<i>Liquidambar styraciflua</i>	abaxial	↔	↑ 30%	
		<i>Zea mays</i> (C ₄)	abaxial	↔	↔	
			adaxial	↔	↔	

Experiment Length (days)	CO ₂ Levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
~50	↑ 68%	<i>Anthyllis vulneraria</i>	abaxial	*↓ 23%	↔	Bryant et al., 1998 ^c
		<i>Sanguisorba minor</i>	abaxial	↔	↔	
		<i>Bromopsis erecta</i>	abaxial	↔	↔	
50	↓ ~32%	<i>Avena sativa</i>	abaxial	↔	-	Malone et al., 1993 ^b
		<i>Prosopis glandulosa</i>	adaxial	↔	-	
			abaxial	↔	-	
		<i>Schizachyrium scoparium</i> (C ₄)	adaxial	↔	-	
			abaxial	↔	-	
		<i>Triticum aestivum</i>	abaxial	↔	-	
adaxial	↔	-				
54	↑ 93%	<i>Boehmeria cylindrica</i>	-	↔	-	Woodward and Beerling, 1997 ^b
56	↑ 100%	<i>Coleus blumei</i>	abaxial	*↓ 9%	*↓ 4%	Beerling and Woodward, 1995 ^b
		<i>Tropaeolum major</i>	abaxial	*↓ 4%	*↓ 10%	
59	↑ 186%	<i>Pelargonium hortorum</i>	abaxial	↔	-	Kelly et al., 1991 ^b
			adaxial	*↓ 50%	-	
60	↓ 29%	<i>Salix herbacea</i>	combined	*↑ 28%	-	Beerling et al., 1995 ^b
	↑ 100%		combined	*↓ 41%	-	
60	↑ 93%	<i>Ochroma lagopus</i>	abaxial	↔	-	Oberbauer et al., 1985 ^b
			adaxial	↔	-	
63	↑ 157%	<i>Panicum tricanthum</i>	abaxial	*↓ 22%	-	Tipping and Murray, 1999 ^b
		<i>Panicum antidotale</i> (C ₄)	abaxial	↑ 28%	-	
<66	↑ 100%	<i>Gossypium hirsutum</i>	abaxial	↔	↔	Reddy et al., 1998 ^b
			adaxial	↔	↔	
72	↑ 89%	<i>Lolium perenne</i>	adaxial	↑	*↓	Ferris et al., 1996 ^c
80	↑ 100%	<i>Betula pendula</i>	abaxial	↔	↔	Wagner, 1998 ^b

Experiment Length (days)	CO ₂ Levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
90	↑ 100%	<i>Quercus ilex</i>	abaxial	*↓ 27%	-	Paoletti et al., 1997 ^b
90-120	↑ 100%	<i>Andropogon gerardii</i> (C ₄)	abaxial	*↓ 28%	-	Knapp et al., 1994 ^c
			adaxial	↑ 75%	-	
		<i>Salvia pitcheri</i>	abaxial	↑ 40%	-	
			adaxial	↑ 125%	-	
92	↑ 100%	<i>Populus trichocarpa</i>	abaxial	↔	↔	Radoglou and Jarvis, 1990 ^b
			adaxial	↔	↔	
93	↓ 52%	<i>Oryza sativa</i>	abaxial	↓ 29%	-	Rowland-Bamford et al., 1990 ^b
			adaxial	↓ 17%	-	
	↑ 173%	<i>Oryza sativa</i>	abaxial	↔	-	
			adaxial	↔	-	
105	↑ 186%	<i>Pelargonium hortorum</i>	abaxial	↔	-	Kelly et al., 1991 ^b
			adaxial	↔	-	
114	↑ 87%	<i>Arachis hypogaea</i>	abaxial	*↓ 12%	↔	Clifford et al., 1995 ^b
			adaxial	*↓ 16%	*↓ 8%	
120	↑ 757%	<i>Rhizophora mangle</i>	abaxial	*↓ 14%	-	Beerling, 1994 ^b
			abaxial	*↓ 31%	-	
		<i>Musa apiculata</i>	abaxial	↔	-	
			adaxial	↔	-	
120	↑ 100%	<i>Populus trichocarpa</i>	abaxial	*↓ 19%	*↓ 31%	Ceulemans et al., 1995 ^c
			adaxial	↔	↔	
		<i>Populus deltoides</i>	abaxial	*↓ 27%	*↓ 36%	
			adaxial	*↓ 33%	↔	
120	↑ 100%	<i>Quercus petraea</i>	abaxial	*↓ 25%	*↓ 14%	Kürschner et al., 1998 ^b
123	↑ 93%	<i>Pentaclethra macroloba</i>	abaxial	*↓ 7%	-	Oberbauer et al., 1985 ^b
			adaxial	↔	-	

Experiment Length (days)	CO ₂ Levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
125	↑ 49%	<i>Triticum aestivum</i>	abaxial adaxial	↔ ↔	↔ ↔	Estiarte et al., 1994 ^c
135	↑ 100%	<i>Prunus avium</i>	abaxial	↔	-	Centritto et al., 1999 ^c
150	↑ 100%	<i>Chlorophytum picturatum</i> <i>Hedera helix</i> <i>Hypoestes variegata</i>	abaxial abaxial abaxial	*↓ 7% *↓ 10% *↓ 9%	*↓ 23% *↓ 29% *↓ 6%	Beerling and Woodward, 1995 ^b
217	↑ 100%	<i>Maranthus corymbosa</i>	abaxial	*↓ 14%	-	Eamus et al., 1993 ^b
~240	↑ 98%	<i>Picea sitchensis</i>	abaxial	↔	-	Barton and Jarvis, 1999 ^b
270	↑ 114%	<i>Pinus banksiana</i>	-	↔	-	Stewart and Hoddinott, 1993 ^b
300	↑ 97%	<i>Eucalyptus tetrodonta</i>	abaxial	*↓ 20%	-	Berryman et al., 1994 ^{b,c}
~365	↑ 71%	<i>Rumex obtusifolius</i>	abaxial adaxial	*↓ 8% ↔	- -	Pearson et al., 1995 ^b
~400	↑ 100%	<i>Rhizophora mangle</i>	abaxial	*↓ 16%	↔	Farnsworth et al., 1996 ^b
~425	↑ 71%	<i>Bromus erectus</i> <i>Plantago media</i> <i>Sanguisorba minor</i>	abaxial adaxial abaxial adaxial abaxial	↔ ↔ ↔ ↔ ↔	↔ ↔ ↔ ↔ ↔	Lauber and Körner, 1997 ^c
570	↑ 100%	<i>Prunus avium</i> <i>Quercus robur</i>	abaxial abaxial	↔ ↑ ~150%	- -	Atkinson et al., 1997 ^b
600	↑ 97%	<i>Pinus palustris</i>	-	↔	-	Pritchard et al., 1998 ^c
~730	↑ 100%	<i>Picea abies</i> <i>Quercus rubra</i>	- abaxial	↔ ↑ 8%	- -	Dixon et al., 1995 ^c
730	↑ 60%	<i>Tussilago farfara</i>	abaxial	-	*↓ 26%	Beerling and Woodward, 1997 ^b

Experiment Length (days)	CO ₂ Levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
750	↑ 97%	<i>Mangifera indica</i>	abaxial	*↓ 17%	-	Goodfellow et al., 1997 ^b
~840	↑ 99%	<i>Scirpus olneyi</i>	-	↔	-	Drake, 1992 ^c
3 years	↑ 60%	<i>Pinus sylvestris</i>	abaxial	*↓ 16%	-	Beerling, 1997 ^b
			adaxial	*↓ 18%	-	
3 years	↑ 60%	<i>Ginkgo biloba</i>	abaxial	*↓ 20%	*↓ 7%	Beerling et al., 1998a ^b
1155	↑ 50%	<i>Pseudotsuga menziesii</i>	abaxial	↔	-	Apple et al., 2000 ^b
~5 years	↑ ~82%	<i>Citrus aurantium</i>	abaxial	↔	↔	Estiarte et al., 1994 ^c
meta-analysis		43 species (60% showed stomatal density reductions)		*↓ (9.0± 3.3% s.e.)	-	Woodward and Kelly, 1995

* Response inversely relates ($P < 0.05$) to CO₂ concentration

↔ No significant change ($P > 0.05$)

- Not reported

^a Typically between 340 and 360 ppmV

^b Plants grown in enclosed greenhouses or chambers

^c Plants grown in open-top chambers (OTCs)

APPENDIX 1.2

Subfossil Stomatal Responses

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
#	↓ 5%	<i>Salix herbacea</i>	abaxial adaxial	↔ *↑ 83%	- -	Beerling et al., 1992
#	↓ 13%	<i>Eucalyptus pauciflora</i>	combined	*↑ 26%	-	Körner and Cochrane, 1985
#	↓ 6%	<i>Griselinia littoralis</i>	combined	↔	-	Körner et al., 1986
	↓ 13%	<i>Nothofagus menziesii</i>	abaxial	*↑ 21%	-	
	↓ 8%	<i>Ranunculus grahamii</i>	combined	↔	-	
#	↓ 10%	<i>Vaccinium myrtillus</i>	abaxial adaxial	↓ 20% *↑ 425%	- -	Woodward, 1986
#	↓ 6%	<i>Nardus stricta</i>	abaxial adaxial	↔ *↑ 19%	- -	Woodward and Bazzaz, 1988
@	↑ 194%	<i>Tussilago farfara</i>	abaxial	-	*↓ 65%	Beerling and Woodward, 1997
@	↑ 100%	<i>Scirpus lacustris</i>	-	*↓ 19%	-	Bettarini et al., 1997
@	↑ 100%	<i>Allium sphaerocephalon</i>	abaxial	↔	-	Bettarini et al., 1998
		<i>Buxus sempervirens</i>	abaxial	↔	↔	
		<i>Convolvulus arvensis</i>	abaxial	↔	↑ 26%	
		<i>Convolvulus cantabrica</i>	abaxial	↔	↔	
		<i>Conyza canadensis</i>	abaxial	*↓ 26%	↑ 21%	
		<i>Fraxinus ornus</i>	abaxial	*↓ 35%	↔	
		<i>Geranium molle</i>	abaxial	↔	↔	
		<i>Globularia punctata</i>	abaxial adaxial	↔ ↔	↔ ↔	
		<i>Hypericum perforatum</i>	abaxial	↔	↔	
		<i>Plantago lanceolata</i>	abaxial adaxial	↔ ↔	↔ ↔	

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
		<i>Potentilla reptans</i>	abaxial	↔	↔	
		<i>Pulicaria sicula</i>	abaxial	↔	↔	
		<i>Ruscus aculeatus</i>	abaxial	↔	-	
		<i>Scabiosa columbaria</i>	abaxial	↔	↔	
		<i>Silene vulgaris</i>	abaxial	↔	↔	
		<i>Stachys recta</i>	abaxial	*↓ 11%	↔	
		<i>Trifolium pratense</i>	abaxial	↔	↔	
@	↑~130%	<i>Bauhinia multinervia</i>	abaxial	↑ 62%	↑ 41%	Fernández et al., 1998
			adaxial	*↓ 71%	*↓ 73%	
		<i>Spathiphyllum cannifolium</i>	abaxial	↔	↔	
			adaxial	*↓ 72%	*↓ 85%	
@	↑ 40%	<i>Quercus pubescens</i>	abaxial	↔	↔	Miglietta and Rasci, 1993
@	↑ 515%	<i>Arbutus unedo</i>	abaxial	*↓ 29%	*↓ 20%	Jones et al., 1995
@	↑ 114%	<i>Quercus ilex</i>	abaxial	*↓ 26%	-	Paoletti et al., 1998
@	↑ 50%	<i>Boehmeria cylindrica</i>	-	↔	-	Woodward and Beerling, 1997
@	↑ ~71%	<i>Phragmites australis</i>	abaxial	↔	-	van Gardingen et al., 1997
			adaxial	*↓ 45%	-	
37	↑ 15% ^b	<i>Metasequoia glyptostroboides</i>	abaxial	↔	*↓ 17%	D.L. Royer, unpublished data
43	↑ 15% ^b	<i>Betula pendula</i>	abaxial	*↓ 30%	*↓ 32%	Wagner et al., 1996
70	↑ 18% ^c	<i>Acer campestre</i>	abaxial	↔	-	Beerling and Kelly, 1997
		<i>Acer pseudoplatanus</i>	abaxial	↔	-	
		<i>Alliaria petiolata</i>	abaxial	↑ 22%	-	
		<i>Allium ursinum</i>	abaxial	↔	-	
		<i>Alnus glutinosa</i>	abaxial	↑ 132%	-	
		<i>Anemone nemorosa</i>	abaxial	↔	-	
		<i>Arum maculatum</i>	abaxial	*↓ 61%	-	

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
			adaxial	*↓ 80%	-	
		<i>Betula pendula</i>	abaxial	*↓ 39%	-	
		<i>Betula pendula</i>	abaxial	*↓ 43%	-	
		<i>Betula pubescens</i>	abaxial	*↓ 56%	-	
		<i>Carpinus betulus</i>	abaxial	↑ 13%	-	
		<i>Castanea sativa</i>	abaxial	*↓ 24%	-	
		<i>Chamaenerion angustifolium</i>	abaxial	↔	-	
		<i>Circaea lutetiana</i>	abaxial	*↓ 25%	-	
		<i>Cirsium palustre</i>	abaxial	*↓ 22%	-	
		<i>Cornus sanguinea</i>	abaxial	*↓ 16%	-	
		<i>Corylus avellana</i>	abaxial	*↓ 50%	-	
		<i>Crataegus monogyna</i>	abaxial	*↓ 36%	-	
		<i>Dipsacus fullonum</i>	abaxial	↑ 54%	-	
			adaxial	↑ 550%	-	
		<i>Epilobium montanum</i>	abaxial	*↓ 28%	-	
			adaxial	↔	-	
		<i>Fagus sylvatica</i>	abaxial	↑ 33%	-	
		<i>Fagus sylvatica</i>	abaxial	↔	-	
		<i>Fraxinus excelsior</i>	abaxial	↑ 39%	-	
		<i>Geranium dissectum</i>	abaxial	↔	-	
		<i>Geranium robertianum</i>	abaxial	*↓ 58%	-	
			adaxial	↑	-	
		<i>Geum urbanum</i>	abaxial	*↓ 21%	-	
			adaxial	↔	-	
		<i>Glechoma hederacea</i>	abaxial	*↓ 23%	-	
		<i>Hedera helix</i>	abaxial	↑ 101%	-	
		<i>Heracleum sphondylium</i>	abaxial	*↓ 14%	-	
			adaxial	↔	-	
		<i>Hyacinthoides non-scripta</i>	abaxial	↑ 56%	-	
			adaxial	↔	-	

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
		<i>Hypericum hirsutum</i>	abaxial	*↓ 11%	-	
		<i>Hypericum perforatum</i>	abaxial	*↓ 56%	-	
		<i>Ilex aquifolium</i>	abaxial	↑ 31%	-	
		<i>Lamiaestrum galeobdolon</i>	abaxial	↔	-	
		<i>Lathyrus pratensis</i>	abaxial	↔	-	
			adaxial	*↓ 38%	-	
		<i>Ligustrum vulgare</i>	abaxial	*↓ 67%	-	
		<i>Lonicera periclymenum</i>	abaxial	*↓ 27%	-	
		<i>Luzula sylvatica</i>	abaxial	*↓ 44%	-	
		<i>Lysimachia nummularia</i>	abaxial	*↓ 56%	-	
			adaxial	*↓ 67%	-	
		<i>Mercurialis perennis</i>	abaxial	*↓ 17%	-	
		<i>Oxalis acetosella</i>	abaxial	↔	-	
		<i>Populus nigra</i>	abaxial	↑ 46%	-	
		<i>Primula vulgaris</i>	abaxial	*↓ 14%	-	
		<i>Prunella vulgaris</i>	abaxial	*↓ 47%	-	
			adaxial	*↓ 55%	-	
		<i>Prunus avium</i>	abaxial	*↓ 20%	-	
		<i>Pteridium aquilinum</i>	abaxial	↔	-	
		<i>Quercus petraea</i>	abaxial	*↓ 14%	-	
		<i>Quercus robur</i>	abaxial	↔	-	
		<i>Ranunculus ficaria</i>	abaxial	*↓ 21%	-	
			adaxial	↔	-	
		<i>Rosa canina</i>	abaxial	*↓ 28%	-	
		<i>Sambucus nigra</i> (sun)	abaxial	↔	-	
		<i>Sambucus nigra</i> (shade)	abaxial	↔	-	
		<i>Scrophularia nodosa</i>	abaxial	*↓ 18%	-	
		<i>Silene dioica</i>	abaxial	↑ 49%	-	
			adaxial	*↓	-	
		<i>Sorbus aucuparia</i>	abaxial	↔	-	

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
		<i>Stellaria holostea</i>	abaxial	*↓ 28%	-	
		<i>Taxus baccata</i>	abaxial	↔	-	
		<i>Tilia cordata</i>	abaxial	*↓ 34%	-	
		<i>Ulmus glabra</i>	abaxial	↔	-	
		<i>Vaccinium myrtillus</i>	abaxial	↔	-	
			adaxial	↔	-	
		<i>Vicia cracca</i>	abaxial	*↓ 57%	-	
			adaxial	*↓ 20%	-	
		<i>Vicia sepium</i>	abaxial	*↓ 43%	-	
		<i>Viola odorata</i>	abaxial	↔	-	
91	↑ 20% ^c	<i>Betula nana</i>	abaxial	*↓ 29%	-	Beerling, 1993
98	↑ 24% ^c	<i>Salix herbacea</i>	combined	-	*↓ 21%	Rundgren and Beerling, 1999
110	↑ 25% ^c	<i>Betula pubescens</i>	abaxial	*↓ 45%	*↓ 35%	Kürschner, 1996
118	↑ 24% ^c	<i>Quercus petraea</i>	abaxial	-	*↓ 34%	van der Burgh et al., 1993
126	↑ 14% ^c	<i>Salix herbacea</i>	combined	*↓ 22%	-	Beerling et al., 1993
~127	↑ 24% ^c	<i>Salix cinerea</i>	abaxial	*↓ 22%	*↓ 17%	McElwain et al., 1995
144	↑ 23% ^c	<i>Salsola kali</i> (C ₄)	abaxial	-	↔	Raven and Ramsden, 1988
144	↑ 27% ^c	<i>Ginkgo biloba</i>	abaxial	↔	*↓ 44%	D.L. Royer, unpublished data
150	↑ 14% ^c	<i>Salix herbacea</i>	combined	*↓ 26%	-	Beerling et al., 1995
151	↑ 27% ^c	<i>Quercus robur</i>	abaxial	*↓ 23%	-	Beerling and Chaloner, 1993b
173	↑ 25% ^c	<i>Olea europaea</i>	abaxial	*↓ 24%	-	Beerling and Chaloner, 1993c
181	↑ 26% ^c	<i>Fagus sylvatica</i>	abaxial	*↓ 43%	-	Paoletti and Gellini, 1993
		<i>Quercus ilex</i>	abaxial	*↓ 28%	-	
190	↑ 27% ^c	<i>Quercus petraea</i>	abaxial	*↓ 40%	*↓ 31%	Kürschner et al., 1996

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
200	↑ 24% ^c	<i>Acer pseudoplatanus</i> , <i>Carpinus betulus</i> , <i>Fagus sylvatica</i> , <i>Populus nigra</i> , <i>Quercus petraea</i> , <i>Q. robur</i> , <i>Rhamnus catharticus</i> , <i>Tilia cordata</i>	abaxial	*↓ 40% (mean)	-	Woodward, 1987
240	↑ 25% ^c	<i>Alnus glutinosa</i> , <i>Amaranthus caudatus</i> , <i>Betula pendula</i> , <i>Buxus sempervirens</i> , <i>Ceratonia siliqua</i> , <i>Cynodon dactylon</i> , <i>Gentiana alpina</i> , <i>Helleborus foetidus</i> , <i>Juniperus communis</i> , <i>Papaver alpinum</i> , <i>Pinus pinea</i> , <i>P. uncinata</i> , <i>Pistacia lentiscus</i> , <i>Rhododendron ferrugineum</i>	combined	*↓ 17% (mean)	↔ (mean)	Peñuelas and Matamala, 1990
3318	↑ 22% ^d	<i>Olea europaea</i>	abaxial	*↓ 33%	-	Beerling and Chaloner, 1993c

* Response inversely relates ($P < 0.05$) to CO₂ concentration

↔ No significant change ($P > 0.05$)

- Not reported

^a Typically between 340 and 360 ppmV; for herbarium studies, control corresponds with oldest material

^b From direct measurements from Mauna Loa Observatory, Hawaii and South Pole (Keeling et al., 1995)

^c From Siple Station ice core (Neftel et al., 1985; Friedli et al., 1986)

^d From Taylor Dome ice core (Indermühle et al., 1999)

Data from an altitudinal study; thus, the 'age' is however long the population has existed at the sampled altitudes

@ Data from a natural CO₂ spring area; thus, the 'age' is however long the population has existed at the location, assuming constant CO₂ emissions

APPENDIX 1.3

Fossil Stomatal Responses

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
9000	↓ 25% ^{b,c}	<i>Salix herbacea</i>	combined	-	*↑ 55%	Rundgren and Beerling, 1999
9190	↓ 27% ^b	<i>Salix cinerea</i>	abaxial	*↑ 57%	*↑ 32%	McElwain et al., 1995
9800	↑ 20% ^{d,j}	<i>Betula pubescens</i> , <i>B. pendula</i>	abaxial	-	*↓ 32% (mean)	Wagner et al., 1999
10750 (Allerød/Y. Dryas)	↓ 25% ^{c,k}	<i>Salix herbacea</i>	combined	*↑ 27%	-	Beerling et al., 1995
11500	↓ 24% ^b	<i>Salix herbacea</i>	combined	↓ 46%	↔	Beerling et al., 1992
13000	↓ 29% ^b	<i>Betula nana</i>	abaxial	*↑ 60%	-	Beerling, 1993
16500	↓ 47% ^b	<i>Salix herbacea</i>	combined	*↑ 54%	*↑ 25%	Beerling et al., 1993
28000	↓ 46% ^b	<i>Pinus flexilis</i>	-	*↑ 31%	-	van de Water et al., 1994
140,000	↓ 47% ^b	<i>Salix herbacea</i>	combined	*↑ 73%	*↑ 39%	Beerling et al., 1993
2.5 m.y.	↑ 4% ^e	<i>Quercus petraea</i>	abaxial	-	*↓ 10%	van der Burgh et al., 1993; Kürschner et al., 1996
6.5 m.y.	↓ 20% ^e	<i>Quercus petraea</i>	abaxial	-	*↑ 55%	van der Burgh et al., 1993;
	↓ 24% ^{e,l}	<i>Fagus attenuata</i>	abaxial	-	*↑ 41%	Kürschner et al., 1996
6.5 m.y.	↓ 20% ^d	<i>Betula subpubescens</i>	abaxial	*↑ 72%	*↑ 45%	Kürschner, 1996
10 m.y.	↑ 4% ^e	<i>Quercus petraea</i>	abaxial	-	*↓ 9%	van der Burgh et al., 1993; Kürschner et al., 1996
10 m.y.	↔ ^d	<i>Betula subpubescens</i>	abaxial	*↔	*↔	Kürschner, 1996

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source
15.5 m.y.	↑ ^f	<i>Chamaecyparis linguaefolia</i> , <i>Cunninghamia chaneyi</i> , <i>Metasequoia occidentalis</i> , <i>Pinus harneyana</i> , <i>Pinus</i> sp., <i>Taxodium dubium</i>	combined	*↓ (mean)	-	Huggins, 1985
44-50 m.y. (M. Eocene)	↑ 43% ^g	<i>Lindera cinnamomifolia</i> , <i>Lindera</i> sp. ⁿ	abaxial	*↓ 36% (mean)	*↓ 47% (mean)	McElwain, 1998
		<i>Litsea bournensis</i> , <i>L. edwardsii</i> , <i>L. hirsuta</i> ⁿ	abaxial	*↓ 27% (mean)	*↓ 38% (mean)	
160-185 m.y. (M. Jurassic)	↑ 149% ^g	<i>Brachyphyllum crucis</i> ⁿ	abaxial	*↓ 54%	*↓ 39%	McElwain and Chaloner, 1996
		<i>B. mamillare</i> ⁿ	abaxial	*↓ 39%	*↓ 52%	
		<i>Ginkgo huttonii</i> ⁿ	abaxial	*↓ 32%	-	
160-185 m.y. (M. Jurassic)	↑ 149% ^g	<i>Baeira furcata</i> ⁿ	abaxial	*↓ 44%	-	McElwain, 1998
			adaxial	*↓ 67%	-	
		<i>Ctenis exilis</i> , <i>C. kaneharai</i> , <i>C. sulcaulis</i> ⁿ	abaxial	*↓ 46% (mean)	↑ 14% (mean)	
		<i>Pagiophyllum kurrii</i> , <i>P. maculosum</i> , <i>P. ordinatum</i> ⁿ	abaxial	*↓ 36% (mean)	*↓ 39% (mean)	

Age of Material (years)	CO ₂ levels (relative to controls ^a)	Species Used	Side of Leaf	Stomatal Density Response	Stomatal Index Response	Source	
~205 m.y. (Latest Triassic)	↑ 69% ^h	<i>Baeira boeggildiana</i> ⁿ	abaxial	-	*↓ 44%	McElwain et al., 1999	
		<i>B. minuta</i> ⁿ	abaxial	-	*↓ 49%		
		<i>B. paucipartata</i> ⁿ	abaxial	-	*↓ 25%		
		<i>Baeira</i> sp. ⁿ	abaxial	-	*↓ 36%		
		<i>Ctenis minuta</i> , <i>C. nilssonii</i> ⁿ	abaxial	-	*↓ 43%		
		(mean)					
		<i>C. nilssonii</i> ⁿ	abaxial	-	*↓ 21%		
~205 m.y. (Earliest Jurassic)	↑ 567% ^h	<i>Ginkgo acosmica</i> ⁿ	abaxial	-	*↓ 26%		
		<i>G. obovatus</i> ⁿ	abaxial	-	*↓ 57%		
		<i>Baeira longifolia</i> ⁿ	abaxial	-	*↓ 60%		
		<i>B. spectabilis</i> ⁿ	abaxial	-	*↓ 71%		
		<i>Nilssonia polymorpha</i> ⁿ	abaxial	-	*↓ 70%		
		<i>Stenopteris dinosaurensis</i> ⁿ	abaxial	-	*↓ 11%		
285-290 m.y. (E. Permian)	↔ ^g	<i>Lebachia frondosa</i> ⁿ	abaxial	↑ 120%	*↔	McElwain and Chaloner, 1995	
290-303 m.y. (L. Penn.)	↑ ^l	<i>Neuropteris ovata</i>	abaxial	*↓ 40%	*↓ 27%	Cleal et al., 1999	
310 m.y. (L. Penn.)	↔ ^g	<i>Swillingtonia denticulata</i> ⁿ	abaxial	↑ 460%	*↔	McElwain and Chaloner, 1995	
388-373 m.y. (M. Devonian)	↓ 61% ^{g,m}	<i>Drepanophycus spinaeformis</i>	-	*↑ 42%	*↑ 38%	Edwards et al., 1998	
390-403 m.y. (E. Devonian)	↑ 657% ^g	<i>Aglaophyton major</i> ⁿ	combined	*↓ 99%	*↓ 84%	McElwain and Chaloner, 1995	
		<i>Sawdonia ornata</i> ⁿ	combined	*↓ 98%	*↓ 78%		

* Response inversely relates ($P < 0.05$) to CO₂ concentration

↔ No significant change ($P > 0.05$)

- Not reported

^a Typically between 340 and 360 ppmV

^b From Vostok (Barnola et al., 1987) and Taylor Dome (Indermühle et al., 1999) ice cores

- ^c From stomatal response of recent *Salix herbacea*, where CO₂ concentrations are known; values match ice core data^b
- ^d From stomatal responses of recent *Betula pubescens* and *Betula pendula*, where CO₂ concentrations are known
- ^e From stomatal response of recent *Quercus petraea*, where CO₂ concentrations are known; values correlate with temperature curve
- ^f From Freeman and Hayes, 1992; Cerling et al., 1997 (c.f. Pagani et al., 1999a)
- ^g From 'best estimate' of Berner (1994, 1998)
- ^h From stomatal ratios (McElwain and Chaloner, 1995, 1996; McElwain, 1998)
- ⁱ This likely CO₂ spike at the Westphalian/Stephanian boundary is corroborated by a warming interval in Gondwanan and Anfaran areas; the control group is the Westphalian material
- ^j The control group is prior to the CO₂ spike (260 ppmV CO₂^d)
- ^k The control group is the late Allerød material, prior to CO₂ drop (273 ppmV CO₂^c)
- ^l The control group is the 10 Ma material (370 ppmV CO₂^e)
- ^m The control group is the 388 Ma material (2600 ppmV CO₂^g)
- ⁿ Stomatal responses compared with corresponding Nearest Living Equivalents (NLEs); method described in text

APPENDIX 2.1

Summary of sedimentology at ginkgo-bearing sites. W = Williston Basin (western North and South Dakota, USA); BHB = Bighorn Basin (Wyoming and Montana, USA); DB = Denver Basin (Colorado, USA); S = southwestern Spitsbergen (Norway); A = south-central Alberta (Canada); IM = Isle of Mull (Scotland); E = southwestern Ellesmere Island (Canada); ID = north-central Idaho (USA); MT = western Montana (USA). Sedimentological contexts with question marks not included in analysis.

Site #	Site name	Location	Age (Ma)	Sedimentary features	Interpretation
1	DMNH 571	W	65.9	sandstone-mudstone couplets	relief channel
2	DMNH 1492	W	65.9	tabular sand with root traces on top	crevasse splay
3	DMNH 572	W	65.8	sandstone-mudstone couplets	relief channel
4	DMNH 425	W	65.6	cross-bedded sandstone	channel
5	DMNH 2087	W	65.5	sandstone-mudstone couplets	relief channel
6	DMNH 568	W	65.5	cross-bedded sandstone	channel
7	DMNH 566	W	65.5	gleyed mudstone with abundant root traces	distal floodplain
8	DMNH 1489	W	65.4	sandstone-mudstone couplets	relief channel
9	KJ 83101 / 8415	BHB	64.5	tabular sand	crevasse splay
10	LJH 7423	BHB	64.5	sandstone-shale-lignite upward sequence; Ginkgo in sand	crevasse splay
11	LJH 7424c	BHB	64.5	ledgey cross-bedded sandstone	crevasse splay
12	LJH 7425 / 7653	BHB	64.5	ledgey cross-bedded sandstone	crevasse splay?
13	LJH 7659	BHB	64.5	siltstone (relief channel) at base of clay plug (abandoned cutoff)	relief channel
14	DMNH 2360	DB	64.1	gray claystone w/occasional fine/med sand partings	crevasse splay
15	Basilika	S	64	~8 m siltstones/sandstones resting on ~30 cm pebble/cobble conglom.	relief channel
16	LJH 7861	BHB	64	siltstone/sandstone overlying lignite; Ginkgo in sand	crevasse splay
17	Joffre Bridge	A	58.5	carbonaceous mudstone; Ginkgo in hard mollusc layer	abandoned channel
18	Burbank	A	58.5	coarse sandstone bounded by shales	crevasse splay
19	SLW 0029	BHB	57.8	grey siltstone above carbonaceous shale	crevasse splay
20	SLW 0025	BHB	57.3	siltstone-mudstone couplets; Ginkgo mainly in siltstone	relief channel
21	LJH 72127	BHB	57	lignite-siltstone-sand sequence	crevasse splay
22	SLW 0019	BHB	56.7	cross-bedded medium to fine sandstone; large-scale crosscuts	channel
23	Chance	BHB	56.4	coarsening upwards sequence; Ginkgo in sand	crevasse splay
24	Almont	W	56	siliceous, iron-stained shale containing fish scales overlying fine sand	abandoned channel
25	SLW 993	BHB	55.9	base of channel fill directly above basal lag deposits (pebbles)	abandoned channel
26	SLW 992	BHB	55.9	interlaminated fine/coarse siltstones	abandoned channel
27	SLW 991	BHB	55.9	laminated gray siltstone	abandoned channel
28	SLW 0051	BHB	55.9	laminated siltstone and fine sandstone	crevasse splay
29	LJH 72141-1	BHB	55.8	3 m above brown fissile lignite	?

Site #	Site name	Location	Age (Ma)	Sedimentary features	Interpretation
30	SLW 9155	BHB	55.7	interlaminated siltstone-sandstone	crevasse splay
31	SLW 9427 / 9437	BHB	55.5	thinly bedded sandstone	crevasse splay
32	SLW 9411	BHB	55.5	interbedded siltstone and sandstone above a carbonaceous shale	crevasse splay
33	SLW 9412	BHB	55.5	laminated fine sand	distal floodplain
34	SLW 9438	BHB	55.5	thinly bedded sandstone	crevasse splay
35	SLW 9434	BHB	55.4	interlaminated cross-bedded siltstone-sandstone; Ginkgo in siltstone	crevasse splay
36	SLW 9050	BHB	55.3	interlaminated siltstone-sandstone	crevasse splay
37	SLW 9715 / 981	BHB	55.3	laminated fining upwards sequences; basal lag contains molluscs	relief channel
38	SLW 9936	BHB	55.3	siltstone overlying sandstone	?
39	SLW 8612	BHB	55.3	carbonaceous shale	backswamp
40	Ardtun Head	IM	55.2	white/grey clay in a clay/sandstone sequence interbedded with basalt	relief channel
41	SLW 9812	BHB	55.1	gray shale-mudstone couplets	relief channel
42	Stenkul Fiord	E	55	medium to fine cross-bedded sandstone with climbing ripples	channel
43	SLW 9911	BHB	54.8	mudstone/siltstone couplets; green sand at base	relief channel
44	SLW 9915	BHB	54.8	sandstone-carbonaceous shale-siltstone unit; Ginkgo in top siltstone	relief channel
45	SLW 841	BHB	54.3	carbonaceous shale	backswamp
46	SLW LB	BHB	53.9	coarse carbonaceous siltstone	relief channel
47	SLW H	BHB	53.5	sandy siltstone	relief channel
48	SLW near YPM 67	BHB	53.5		relief channel
49	LJH 9915	BHB	53.4	thin carbonaceous lenticular paper shale sandwiched between soils	relief channel
50	Beaver Creek	MT	36		lake
51	Juliaetta	ID	16.5	coarse sand; delta foresets	lake

APPENDIX 2.2

Summary of flora at ginkgo-bearing sites. Only those species that occur at ≥ 4 sites are tabulated here. n = number of occurrences for a given species. See Appendix 2.1 for full description of sites.

n	Taxon	Site name															
		DMNH 571	DMNH 572	DMNH 425	DMNH 2087	DMNH 568	DMNH 566	DMNH 1489	KJ 83101	LJH 7423	LJH 7424c	LJH 7425	LJH 7659	Basilika	LJH 7861	Joffre Bridge	Burbank
Site #	Age (Ma)	1	3	4	5	6	7	8	9	10	11	12	13	15	16	17	18
Species richness		65.9	65.8	65.6	65.5	65.5	65.5	65.4	64.5	64.5	64.5	64.5	64.5	64	64	58.5	58.5
		15	9	12	11	13	15	12	13	7	6	10	11	4	19	5	4
28	<i>Cercidiphyllum genatrix</i>		X		X				X	X	X	X	X	X	X		X
21	<i>Metasequoia occidentalis</i>	X	X				X		X	X	X	X	X	X	X	X	
20	<i>Platanus raynoldsii / guillelmae</i>								X	X	X	X	X		X		
14	<i>Glyptostrobus europaeus</i>	X							X			X	X		X	X	
8	<i>Dryophyllum subfalcata</i>	X	X	X	X	X	X	X							X		
8	<i>Menispermites parvareolatus</i>																
8	<i>Platanus nobilis / brownii</i>															X	
7	<i>Ampelopsis acerifolia</i>								X			X			X		
6	Betulaceae sp. 1																
6	<i>Ficus artocarpoides</i>								X			X	X		X		
6	<i>Ficus planicostata</i>										X						
6	<i>Erlingdorgia montana</i>	X	X	X		X	X	X									
6	<i>Leepierceia preartocarpoides</i>	X	X	X		X	X	X									
6	<i>Rhamnus cleburni</i>			X	X	X	X	X							X		
6	<i>Taxodium olrikii</i>	X		X		X	X	X					X				
6	<i>Vitis stantoni</i>	X	X		X		X				X				X		
5	<i>Corylus insignis</i>														X		
5	<i>Trochodendroides nebrascensis</i>	X		X		X	X	X									
5	<i>Viburnum cupanioides</i>														X		
5	<i>Zingiberopsis isonervosa</i>																
4	<i>Averrhoites affinis</i>																
4	<i>Aesculus hickeyi</i>														X		
4	<i>Nyssidium arcticum</i>	X		X			X										
4	<i>Paranymphea crassifolia</i>								X			X	X		X		
4	Platanaceae	X	X	X		X											

Appendix 2.2 continued

		Site name															
		SLW 0029	SLW 0025	LJH 72127	SLW 0019	Chance	Almont	SLW 993	SLW 992	SLW 991	SLW 0051	LJH 72141	SLW 9155	SLW 9427	SLW 9411	SLW 9412	SLW 9438
Site #	Age (Ma)	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34
Species richness		57.8	57.3	57	56.7	56.4	56.0	55.9	55.9	55.9	55.9	55.8	55.7	55.5	55.5	55.5	55.5
<i>n</i>	Taxon	3	3	10	8	11	11	6	4	5	5	12	4	8	4	4	11
28	<i>Cercidiphyllum genatrix</i>	X	X	X	X	X		X		X	X	X		X			X
21	<i>Metasequoia occidentalis</i>			X	X	X		X				X					X
20	<i>Platanus raynoldsii / guillelmae</i>		X		X	X		X			X		X	X		X	X
14	<i>Glyptostrobus europaeus</i>			X	X	X						X					
8	<i>Dryophyllum subfalcatum</i>																
8	<i>Menispermites parvareolatus</i>				X			X		X							
8	<i>Platanus nobilis / brownii</i>													X		X	
7	<i>Ampelopsis acerifolia</i>	X		X		X											X
6	Betulaceae sp. 1												X	X			X
6	<i>Ficus artocarpoides</i>										X			X			
6	<i>Ficus planicostata</i>													X	X	X	X
6	<i>Erlingdorffia montana</i>																
6	<i>Leopierceia preartocarpoides</i>																
6	<i>Rhamnus cleburni</i>																
6	<i>Taxodium olrikii</i>																
6	<i>Vitis stantoni</i>																
5	<i>Corylus insignis</i>							X	X	X							
5	<i>Trochodendroides nebrascensis</i>								X	X							
5	<i>Viburnum cupanioides</i>			X					X	X							
5	<i>Zingiberopsis isonervosa</i>											X					
4	<i>Averrhoites affinis</i>								X								
4	<i>Aesculus hickeyi</i>			X								X					
4	<i>Nyssidium arcticum</i>						X										
4	<i>Paranymphea crassifolia</i>																
4	Platanaceae																

Appendix 2.2 continued

	Site name	SLW 9434	SLW 9050	SLW 9715	SLW 9936	SLW 8612	Ardtun Head	SLW 9812	SLW 9911	SLW 9915	SLW 841	SLW LB	SLW H	YPM 67	LJH 9915	Julietta
	Site #	35	36	37	38	39	40	41	43	44	45	46	47	48	49	50
	Age (Ma)	55.4	55.3	55.3	55.3	55.3	55.2	55.1	54.8	54.8	54.3	53.9	53.5	53.5	53.4	16.5
	Species richness	3	5	5	1	4	8	7	6	10	5	22	22	3	3	2
<i>n</i>	Taxon															
28	<i>Cercidiphyllum genatrix</i>							X	X	X	X	X	X			X
21	<i>Metasequoia occidentalis</i>		X			X		X		X						
20	<i>Platanus raynoldsii / guillelmae</i>			X						X		X	X	X		
14	<i>Glyptostrobus europaeus</i>							X	X	X		X				
8	<i>Dryophyllum subfalcata</i>															
8	<i>Menispermites parvareolatus</i>			X				X	X		X	X				
8	<i>Platanus nobilis / brownii</i>	X		X				X	X				X			
7	<i>Ampelopsis acerifolia</i>															
6	Betulaceae sp. 1		X									X	X			
6	<i>Ficus artocarpoides</i>															
6	<i>Ficus planicostata</i>	X														
6	<i>Erlingdorgia montana</i>															
6	<i>Leepierceia preartocarpoides</i>															
6	<i>Rhamnus cleburni</i>															
6	<i>Taxodium olrikii</i>															
6	<i>Vitis stantoni</i>															
5	<i>Corylus insignis</i>			X												
5	<i>Trochodendroides nebrascensis</i>															
5	<i>Viburnum cupanioides</i>							X								
5	<i>Zingiberopsis isonervosa</i>										X	X	X			X
4	<i>Averrhoites affinis</i>							X			X	X				
4	<i>Aesculus hickeyi</i>					X										
4	<i>Nyssidium arcticum</i>															
4	<i>Paranymphaea crassifolia</i>															
4	Platanaceae															

APPENDIX 2.3

Complete list of species names with authors used in Chapter 2.

Acer silberlingii Brown

Aesculus hickeyi Manchester

Amentotaxus formosana Li

Amentotaxus gladifolia (Ludwig) Ferguson, Jähnichen, & Alvin

Archaeampelos acerifolia (Newberry) McIver & Basinger

Athyrium Felix-femina Linnaeus

Averrhoites affinis (Newberry) Hickey

Beringeaphyllum cupanoides (Newberry) Manchester, Crane & Golovneva

Betulaceae sp. 1

Cercidiphyllum genatrix (Newberry) Hickey

Cercidiphyllum japonicum Siebold & Zuccarini

Corylus insignis Heer

Dryophyllum subfalcata Lesquereux

Erlingdorgia montana Johnson

"Ficus" artocarpoides Lesquereux

"Ficus" planicostata Lesquereux

Ginkgo adiantoides (Unger) Heer

Ginkgo beckii Scott, Barghoorn & Prakash

Ginkgo biloba Linnaeus

Ginkgo coriacea Florin

Ginkgo digitata (Brongniart) Heer

Ginkgo gardnerii Florin

Ginkgo huttonii (Sternberg) Heer

Ginkgo spitzbergensis Manum

Ginkgo tigrensis Archangelsky

Ginkgo yimaensis Zhou & Zhang

Glyptostrobus europaeus (Brongniart) Heer

Leepierceia preartocarpoides Johnson

Metasequoia glyptostroboides H. H. Hu & Cheng

Metasequoia occidentalis (Newberry) Chaney

Nyssidium arcticum (Heer) Iljinskaya

Paranymphea crassifolia (Newberry) Berry

Platanaceae

Platanus X acerifolia Willdenow

Platanus brownii (Berry) MacGinitie

Platanus guillelmae Geoppert

Platanus mexicana Moricand

Platanus nobilis Newberry

Platanus occidentalis Linnaeus

Platanus orientalis Linnaeus

Platanus racemosa Nuttall

Platanus raynoldsii Newberry

Platanus wrightii Watson

"Rhamnus" cleburni Lesquereux

Taxodium olrikii (Heer) Brown

Trochodendroides nebrascensis (Newberry) Dorf

Vitis stantonii (Knowlton) Brown

Wardiaphyllum daturaefolia (Ward) Hickey

Zingiberopsis isonervosa Hickey

Zizyphoides flabella (Newberry) Crane, Manchester & Dilcher

APPENDIX 3.1

Voucher specimen information for historical collections used to construct training sets. All catalogued specimens are deposited at the Yale Peabody Museum (YPM) Herbarium. Permanent slides of *Ginkgo* were not prepared (see Table 4.3 for herbaria information).

Date	CO ₂ (ppmV)	SI (%)	Source herbarium ^a	Source location of leaves	Cuticle accession number
<i>Metasequoia glyptostroboides</i>					
1947	310	11.89	UC	E Szechuan / NW Hubei, China	dlr 00289-00293
1948	310	11.63	UC	E Szechuan / NW Hubei, China	dlr 00294-00298
1949	311	11.47	UC	Berkeley, CA	dlr 00299-00303
1950	311	11.62	UC	Bar Harbor, ME	dlr 00304-00308
1953	312	11.07	UC	San Rafael, CA	dlr 00309-00313
1961	317	10.56	UC	Tallahassee, FL	dlr 00314-00318
1962	318	11.49	YPM	Bethany, CT	(acetate peels) dlr 9924-9928
1964	319	10.20	MA	PA	(acetate peels) dlr 9931-9935
1965	320	10.81	UC	Washington DC	dlr 00319-00323
1968	323	9.44	UC	Fukuchiyama-city, Japan	dlr 00324-00328
1969	323	10.83	BBG	Brooklyn Botanical Garden, NY	(acetate peels) dlr 999-9913
1973	328	10.40	NA	Turlock, CA	(acetate peels) dlr 0064-0068
1975	331	10.16	UC	U of Oregon, Eugene, OR	dlr 00329-00333
1976	331	9.54	UC	U of Oregon, Eugene, OR	dlr 00334-00338
1980	338	9.73	UC	Western Hubei, China	dlr 00339-00343
1999	367	9.57	modern	New Haven, CT; London, UK	(various acetate peels)
<i>Platanus</i> sp.					
1891	294	15.35	YPM	Stanley Co., NC (<i>P. occidentalis</i>)	dlr 0129-0134
1975	330	15.22	YPM	Gali Ali Beg, Iraq (<i>P. orientalis</i>)	dlr 0124-0128
2001	370	13.28	modern	New Haven, CT (<i>P. x acerfolia</i>)	dlr 0135-0138
2001	370	13.44	modern	New Haven, CT (<i>P. occidentalis</i>)	dlr 0139-0144

^a YPM = Peabody Museum of Natural History (Yale University); NA = U.S. National Arboretum; BBG = Brooklyn Botanic Gardens; MA = Morris Arboretum (University of Pennsylvania); UC = University Herbarium (University of California, Berkeley); modern = collections of fresh leaves

APPENDIX 3.2

Voucher specimen information for experimental collections used to construct training sets. All catalogued specimens are deposited at the Yale Peabody Museum (YPM) Herbarium.

CO ₂ (ppmV)	SI (%)	Plant age (yr)	Cuticle accession number
<i>Ginkgo biloba</i>			
431	8.70	6	dlr 00203-00218
	8.73	1	dlr 00226-00233
446	8.13	7	dlr 0172-0179
	8.00	2	dlr 0164-0171
791	7.16	6	dlr 00187-00202
	6.84	1	dlr 00219-00225
801	6.99	7	dlr 0156-0163
	6.67	2	dlr 0148-0155
<i>Metasequoia glyptostroboides</i>			
431	9.04	1	dlr 00349-00364
446	9.31	2	dlr 0180-0183
791	7.84	1	dlr 00365-00380
801	7.96	2	dlr 0184-0187

Age (Ma)	SI (%)	Site	Depository ^a	Rock accession number ^b	Cuticle accession number
65.4	8.42	^d Wet Butte (DMNH 1489)	DMNS	DMNH 20613(1) DMNH 20610(1)	DMNH 20613(1) DMNH 20610(1)
64.5	9.48	Ginkgo (LJH 7423)	USNM		USNM 519346-519350
64.5	9.32	Debeya (LJH 7659)	YPM		YPM 40794-40800 YPM 45166-45173
64.1	9.90	Eat My Ginkgo (DMNH 2360)	DMNS	DMNH 23071-23075	DMNH 23071-23075
64	9.42	Spitsbergen (Basilika Fm.)	PMO		PMO PA164.991 PMO PA164.992 PMO PA164.998 PMO PA165.003 PMO PA167.556 PMO PA167.557
			YPM		YPM 45128
63	8.60	^d ? (coll. by E. Dorf)	YPM		YPM 45127
58.5	7.55	Burbank (Paskapoo Fm.)	UA	S17775 S17778 S17777 S24573B S17764 S17779 S51314	dlr 00129 dlr 00130 dlr 00131 dlr 00132 dlr 00133 dlr 00134 dlr 00135
58.5	7.96	Joffre Bridge (Paskapoo Fm.)	UA	S39561 S39559 S39552 S39555 S39549	dlr 00136 dlr 00137 dlr 00138 dlr 00139 dlr 00140
58	11.24	^d Linch (UF 18255)	FMNH	UF 25310 UF 25313-25314	UF 25310 UF 25313-25314
57.3	9.01	SLW 0025	USNM		USNM 519351-519357
56.4	8.75	Chance (LJH 7132/72118; USNM 14184)	YPM	USNM 511364	USNM 511364 YPM 45196 YPM 45914 YPM 45197-45198
56	8.86	^d LB 2373 (coll. By E.L. Simons)	YPM		YPM 40013
56	8.89	^d Almont/Sentinel Butte (DMNH 907)	DMNS		dlr 0057-0058

Age (Ma)	SI (%)	Site	Depository ^a	Rock accession number ^b	Cuticle accession number
55.9	10.97	SLW 991	USNM		USNM 519358-519362
55.9	10.80	SLW 992	USNM		dlr 00115-00117 USNM 519363-519367
55.9	11.43	SLW 993	USNM		USNM 516616-516622
55.8	10.63	Powell dump / J.P. Reiss (LJH 72141-1)	YPM		YPM 45129 YPM 45130
			USNM		dlr 9952-9954 dlr 9945-9946
55.7	11.21	SLW 9155	USNM		USNM 519368-519377
55.6	11.5	SLW 9411	USNM		USNM 519378-519385
55.4	12.23	SLW 9434	USNM		USNM 519386-519392
55.3	8.23	SLW 9715/981	USNM		dlr 0071-0075 dlr 0081-0085 dlr 00100-00101
55.3	12.18	SLW 9050	USNM		dlr 00110-00114
55.3	11.77	SLW 9936	USNM		dlr 9960-9964 dlr 9939-9942 dlr 9955-9959
55.3	12.41	SLW 8612	USNM		dlr 9947-9951 dlr 9943-9944
55.2	6.54	Isle of Mull, Scotland [<i>G. gardneri</i>]	PMO		PMO PA165.018 PMO PA165.020 PMO PA165.022
			YPM		YPM 45126
			BNHM		PA 2997 PA 2999
55.1	8.53	SLW 9812	USNM		dlr 0076-0080 dlr 0096-0097 dlr 00104 dlr 0086-0090 dlr 00102-00103 dlr 0091-0095 dlr 0098-0099
55	8.29	^d Stenkul Fiord (LJH 8411)	YPM		YPM 45199 YPM 48913

- ^a PMO = Oslo Paleontological Museum; DMNS = Denver Museum of Nature and Science; YPM = Yale Peabody Museum; USNM = National Museum of Natural History, Smithsonian Institution; FMNH = Florida Museum of Natural History; UA = University of Alberta; BNHM = British Natural History Museum; UI = University of Idaho
- ^b Accession number of rock from which cuticle was removed
- ^c *Ginkgo adiantoides* unless noted otherwise
- ^d Data not used in CO₂ reconstruction (i.e., Figs. 3.2 and 4.3)

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